IN THE UNITED STATES PATENT AND TRADEMARK OFFICE



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BEN SH	EN, WEN LIU, STEVEN D.)	
	TENSON and SCOTT STANDAGE)	
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For:	GENE CLUSTER FOR)	
]	PRODUCTION OF THE ENEDIYNE)	
	ANTITUMOR ANTIBIOTIC C-1027	Ś	
		Ś	San Francisco, Californ

Patent Application Assistant Commissioner for Patents Washington, D.C. 20231

By Express Mail No: EL160743652US

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PATENT APPLICATION TRANSMITTAL

Sir:

Transmitted herewith for filing is the patent application of inventor(s) Ben Shen, Wen Liu, Steven D. Christenson and Scott Standage, for "GENE CLUSTER FOR PRODUCTION OF THE ENEDIYNE ANTITUMOR ANTIBIOTIC C-1027." Enclosed are:

- 1. 64 pages of the specification, including 71 claims and an abstract.
- 2. 11 sheets of drawings.
- 3. 79 pages of Sequence Listing.
- 4. An oath or declaration of the inventors (unsigned).

The filing fee is being deferred at this time.

Dated: January 5, 2000.

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In the United States Patent and Trademark Office U.S. Patent Application For

GENE CLUSTER FOR PRODUCTION OF THE ENEDIYNE ANTITUMOR ANTIBIOTIC C-1027

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GENE CLUSTER FOR PRODUCTION OF THE ENEDIYNE ANTITUMOR ANTIBIOTIC C-1027

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims benefit under 35 U.S.C. §119 of provisional application USSN 60/115,434, filed on January 6, 1999, which is herein incorporated by reference in its entirety for all purposes.

STATEMENT AS TO RIGHTS TO INVENTIONS MADE UNDER FEDERALLY SPONSORED RESEARCH AND DEVELOPMENT

This work was supported in part by a grant from the Cancer Research Coordinating Committee, University of California, the National Institutes of Health grant CA78747, and the Searle Scholars Program/The Chicago Community Trust. The Government of the United States of America may have certain rights in this invention.

FIELD OF THE INVENTION

This invention relates to the field of enediyne antibiotics. In particular this invention elucidates the gene cluster controlling the biosynthesis of the C-1027 enediyne.

BACKGROUND OF THE INVENTION

The enediyne antibiotics are currently the focus of intense research activity in the fields of chemistry, biology, and medical sciences, because of their unique molecular architecture, biological activities, and modes of actions (Doyle and Borders (1995) *Enediyne antibiotics as antitumor agents*. Marcel-Dekker, New York, Thorson *et al.* (1999) *Bioorg. Chem.*, 27: 172-188). Since the unveiling of the structure of neocarzinostatin chromophore (Edo *et al.* (1985) *Tetrahedron Lett.* 26: 331-340) in 1985, the enediyne family has grown steadily. Thus far, there have been three basic groups within the enediyne antibiotic family: (a) the calicheamicin/esperamicin type, which includes the calicheamicins, the esperamicins, and namenamicin, (b) the dynemicin type, and (c) the chromoprotein type, consisting of an apoprotein and an unstable enediyne chromophore. The latter group includes neocarzinostatin, kedarcidin, C-1027 (Fig. 1), and maduropeptin, whose enediyne chromophore structures have been established, as well as several others whose enediyne chromophore structures are yet to be determined due to their instability (Thorson *et al.*

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(1999) *Bioorg. Chem.*, 27: 172-188). N1999A2, in contrast to the other chromoproteins, exists as an enediyne chromophore alone despite the fact that its structure is very similar to the other chromoprotein chromophore (*Ando et al.*(1998) *Tetra. Letts.*, 39: 6495-6480).

As a family, the enediyne antibiotics are the most potent, highly active antitumor agents ever discovered. Some members are 1000 times more potent than adriamycin, one of the most effective, clinically used antitumor antibiotics (Zhen et al. (1989) J. Antibiot. 42: 1294-1298). All members of this family contain a unit consisting of two acetylenic groups conjugated to a double bond or incipient double bond within a nine or ten-membered ring; i.e., the enediyne core as exemplified by C-1027 in Fig. 1. As the consequence of this structural feature, these compounds share a common mechanism of action: the enediyne core undergoes an electronic rearrangement to form a transient benzenoid diradical, which is positioned in the minor groove of DNA so as to damage DNA by abstracting hydrogen atoms from deoxyriboses on both strands (Fig. 1). Reaction of the resulting deoxyribose carbon-centered radicals with molecular oxygen initiates a process that results in both single-strand and double-strand DNA cleavages (Doyle and Borders (1995) Enediyne antibiotics as antitumor agents. Marcel-Dekker, New York; Ikemoton et al. (1995) Proc. Natl. Acad. Sci. USA 92:10506-10510; Myers et al. (1997) J. Am. Chem. Soc. 119: 2965-2972; Stassinopoulos et al. (1996) Science 272: 1943-1946; Thorson et al. (1999) Bioorg, Chem., 27: 172-188; Xu et al. (1997) J. Am. Chem. Soc. 119: 1133-1134). This novel mechanism of DNA damage has important implications for their application as potent cancer chemotherapeutic agents (Doyle and Borders (1995) supra.; Sievers et al. (1999) Blood 93: 3678-3684).

As an alternative to making structural analogs of microbial metabolites by chemical synthesis, manipulations of genes governing secondary metabolism offer a promising alternative allowing preparation of these compounds biosynthetically (Cane *et al.* (1998) *Science* 282: 63-68; Hutchinson and Fujii. (1995) *Ann. Rev. Microbiol.* 49: 201-38; Katz and Donadio (1993) *Ann. Rev. Microbiol.* 47: 875-912). The success of the latter approach depends critically on the availability of novel genetic systems and on genes encoding novel enzyme activities. The enediynes offer a distinct opportunity to study the biosynthesis of their unique molecular scaffolds and the mechanism of self-resistance to extremely cytotoxic natural products. Elucidation of these aspects provides access to rational engineering of enediyne biosynthesis for novel drug leads and makes it possible to construct enediyne overproducing strains by de-regulating the biosynthetic machinery. In

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addition, elucidation of an enediyne gene cluster contributes to the general field of combinatorial biosynthesis by expanding the repertoire of novel polyketide synthase (PKS) and deoxysugar biosynthesis genes as well as other genes uniquely associated with enediyne biosynthesis, leading to the making of novel enediynes via combinatorial biosynthesis.

SUMMARY OF THE INVENTION

This invention provides nucleic acid sequences and characterization of the gene cluster responsible for the biosynthesis of the enediyne C-1027 (produced by *Streptomyces globisporus*). In particular structural and functional characterization is provided for the 50 open reading frames (ORFs) comprising this gene cluster. Thus, in one embodiment, this invention provides an isolated nucleic acid comprising a nucleic acid selected from the group consisting of a nucleic acid encoding any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA), a nucleic acid encoding a polypeptide encoded by any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA); and a nucleic acid amplified by polymerase chain reaction (PCR) using primer pairs that amplify any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA). In one embodiment, preferred nucleic acids comprise a nucleic acid encoding at least two (more preferably at least three or more) open reading frames (ORFs) selected from the group consisting of ORF-1 through ORF 42, excluding ORF 9 (cagA).

In another embodiment this invention provides an isolated nucleic acid comprising a nucleic acid that specifically hybridizes under stringent conditions to an open reading frame (ORF) of the C-1027 biosynthesis gene cluster, excluding ORF 9 (cagA), and can substitute for the ORF to which it specifically hybridizes to direct the synthesis of an enediyne. In certain embodiments this also includes nucleic acids that would stringently hybridizes indicated above, but for, the degeneracy of the nucleic acid code. In other words, if silent mutations could be made in the subject sequence so that it hybridizes to he indicated sequence(s) under stringent conditions, it would be included in certain embodiments.

Particularly preferred nucleic acids comprises a nucleic acid that specifically hybridizes under stringent conditions to a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 10, ORF 11, ORF 12, ORF 13, ORF 14, ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF

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26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42. Particularly preferred isolated nucleic acid comprises a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 10, ORF 11, ORF 12, ORF 13, ORF 14, ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42. The nucleic acid may comprises a nucleic acid that is a single nucleotide polymorphism (SNP) of a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 81, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 81, ORF 13, ORF 14, ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42.

This invention also provides an isolated gene cluster comprising open reading frames encoding polypeptides sufficient to direct the assembly of a C-1027 enediyne or a C-1027 enediyne analogue. The gene cluster may be present in a cell, more preferably in a bacterial cell (e.g. Actinomycetes, Actinoplanetes, Actinomadura, Micromonospora, or Streptomycetes). Particular preferred bacterial cells include, but are not limited to Streptomyces globisporus, Streptomyces lividans, Streptomyces coelicolor, Micromonospora echinospora spp. calichenisis, Actinomadura verrucosopora, Micromonospora chersina, Streptomyces carzinostaticus, and Actinomycete L585-6. The gene cluster may contain one or more open reading frames is operatively linked to a heterologous promoter (e.g. a constitutive or an inducible promoter).

This invention also provides for an polypeptide encoded by any one or more of the nucleic acids described herein.

Also provided are host cell(s) (e.g. eukaryotic cells or bacterial cells as described herein) transformed with one or more of the expression vectors described herein. Preferred host cells are transformed with an exogenous nucleic acid comprising a gene cluster encoding polypeptides sufficient to direct the assembly of a C-1027 enediyne or a C-1027 enediyne analogue. In certain embodiments, heterologous nucleic acid may comprise only a portion of the gene cluster, but the cell will still be able to express an enediyne.

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This invention also provides methods of chemically modifying a biological molecule. The methods involve contacting a biological molecule that is a substrate for a polypeptide encoded by a C-1027 biosynthesis gene cluster open reading frame, with a polypeptide encoded by a C-1027 biosynthesis gene cluster open reading frame whereby the polypeptide chemically modifies the biological molecule. In one preferred embodiment, the polypeptide is an enzyme selected from the group consisting of a hydroxylase, a homocysteine synthase, a dNDP-glucose dehydrogenase, a citrate carrier protein, a C-methyl transferase, an N-methyl transferase, an aminotransferase, a CagA apoprotein, an NDPglucose synthase, an epimerase, an acyl transferase, a coenzyme F390 synthase, and epoxidase hydrolase, an anthranilate synthase, a glycosyl transferase, a monooxygenase, a type II condensation protein, an aminomutase, a type II adenylation protein, an O-methyl transferase, a P-450 hydroxylase, an oxidoreductase, and a proline oxidase. In a preferred embodiment the method involves contacting the biological molecule with at least two (preferably at least three or more) different polypeptides encoded by C-1027 biosynthesis gene cluster open reading frames. The contacting may be in a host cell (e.g. a eukaryotic cell or a bacterial cell) or the contacting can be ex vivo. The biological molecule can be an endogenous metabolite produced by said host cell or an exogenous supplied metabolite. In preferred embodiments, the host cell is a bacterial cell or eukaryotic cell (e.g., a mammalian cell, a yeast cell, a plant cell, a fungal cell, an insect cell, etc.). In certain preferred embodiments, the host cell synthesizes sugars and glycosylates the biological molecule. In other preferred embodiments, the host cell synthesizes deoxysugars. The method can further involve contacting the biological molecule with a polyketide synthase or a non-ribosomal polypeptide synthetase. The contacting can be in a cell (e.g., a bacterial cell) or ex vivo. In one preferred embodiment the method comprises contacting the biological molecule with at substantially all of the polypeptides encoded by C-1027 biosynthesis gene cluster open reading frames and said method produces an enediyne or enediyne analogue. In another preferred embodiment, the biological molecule is a fatty acid and the biological molecule is contacted with a C-1027 orf polyeptide selected from the group consisting of an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase. In certain embodiments, the biological molecule is a fatty acid and said biological molecule is contacted with a plurality of C-1027 orf polypeptides comprising an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase. In one especially preferred

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embodiment, the biological molecule is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38. In another especially preferred embodiment, the biological molecule is contacted with polypeptides encoded by ORF 15, ORF 16, ORF 28, ORF3, ORF 14, and ORF 13, and, in certain embodiments, ORF 4 and ORF 3 as well.

In certain embodiments, the method may comprise contacting a sugar with one or more C-1027 open reading frame polypeptides selected from the group consisting of a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase. Particularly preferred variant of this method comprise contacting a dNDP-glucose with a plurality of C-1027 open reading frame polypeptides comprising a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase.

In certain other embodiments, the method comprises contacting an amino acid with one or one or more C-1027 open reading frame polypeptides selected from the group consisting of a hydroxylase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein. These methods may involve contacting an amino acid with a plurality of C-1027 open reading frame polypeptides comprising a hydroxylase, a halogenase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein. In particularly preferred embodiments, the amino acid is a tyrosine.

This invention also provides a method of synthesizing a chromaprotein type enediyne core, said method comprising contacting a fatty acid with one or more C-1027 orf polypeptides selected from the group consisting of an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase. In preferred embodiments, the fatty acid may be contacted with a plurality of C-1027 orf polypeptides comprising an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase. In particularly preferred embodiments, the fatty acid is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38.

In still yet another embodiment, this invention provides a method of synthesizing a deoxysugar. This method involves contacting a sugar with one or more C-1027 open reading frame polypeptides selected from the group consisting of a dNDP-glucose

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synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase. In preferred embodiments, this method involves contacting a dNDP-glucose with a plurality of C-1027 open reading frame polypeptides comprising a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase. In particularly preferred embodiments, the dNDP-glucose is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38.

This invention also provides methods of synthesizing a beta amino acid by contacting an amino acid with one or one or more C-1027 open reading frame polypeptides selected from the group consisting of a hydroxylase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein. The method preferably comprises contacting an amino acid with a plurality of C-1027 open reading frame polypeptides comprising a hydroxylase, a halogenase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein. Particularly preferred embodiments comprise contacting the amino acid (*e.g.* tyrosine) with polypeptides encoded by ORF 4, ORF11, ORF24, ORF25, and ORF26.

Also provided are methods of synthesizing an enediyne or an enediyne analogue. These methods involve culturing a cell (e.g. a eukaryotic cell or a bacterium) comprising a recombinantly modified C-1027 gene cluster under conditions whereby said cell expresses said enediyne or enediyne analogue; and recovering the enediyne or enediyne analogue. In preferred embodiments, the gene cluster is present in a bacterium (e.g., Actinomycetes, Actinoplanetes, Actinomadura, Micromonospora, or Streptomycetes). Particularly preferred bacteria include, but are not limited to Streptomyces globisporus, Streptomyces lividans, Streptomyces coelicolor, Micromonospora echinospora spp. calichenisis, Actinomadura verrucosopora, Micromonospora chersina, Streptomyces carzinostaticus, and Actinomycete L585-6. In another preferred embodiment, the gene cluster is present in a eukaryotic cell (e.g. a mammalian cell, a yeast cell, a plant cell, a fungal cell, an insect cell, etc.). The host cell can be one that synthesizes sugars and glycosylates the enediyne or enediyne analogue. The host can be one that synthesizes deoxysugars.

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This invention also provides a method of making a cell (*e.g.*, a bacterial or eukaryotic cell) resistant to an enediyne or an enediyne metabolite. This method involves expressing in the cell one or more isolated C-1027 open reading frame nucleic acids that encode a protein selected from the group consisting of a CagA apoprotein, a SgcB transmembrane efflux protein, a transmembrane transport protein, a Na+/H+ transporter, an ABC transport, a glycerol phosphate transporter, and a UvrA-like protein. In preferred embodiments, the isolated C-1027 open reading frame nucleic acids are selected from the group consisting of ORF 9, ORF2, ORF 27, ORF 0, ORF 1 c-terminus, ORF 2, and ORF 1 N-terminus. Certain embodiments exclude cagA (ORF 9).

In one embodiment, this invention specifically excludes one or more of open reading frames -7 through 42. In particular, in one embodiment this invention excludes cagA (ORF 9), and/or sgcA (ORF 1), and/or sgcB (ORF 2).

DEFINITIONS

The terms "C-1027 open reading frame", and "C-1027 ORF" refer to an open reading frame in the C-1027 biosynthesis gene cluster as isolated from *Streptomyces globisporus*. The term also embraces the same open reading frames as present in other enediyne-synthesizing organisms (*e.g.* other strains and/or species of *Streptomyces*, *Actinomyces*, and the like). The term encompasses allelic variants and single nucleotide polymorphisms (SNPs). In certain instances the C-1027 ORF is used synonymously with the polypeptide encoded by the C-1027 ORF and may include conservative substitutions in that polypeptide. The particular usage will be clear from context.

The terms "isolated" "purified" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native state. With respect to nucleic acids and/or polypeptides the term can refer to nucleic acids or polypeptides that are no longer flanked by the sequences typically flanking them in nature.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers. The term also includes variants on the traditional peptide linkage joining the amino acids making up the polypeptide.

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in physiological environments.

The terms "nucleic acid" or "oligonucleotide" or grammatical equivalents herein refer to at least two nucleotides covalently linked together. A nucleic acid of the present invention is preferably single-stranded or double stranded and will generally contain phosphodiester bonds, although in some cases, as outlined below, nucleic acid analogs are included that may have alternate backbones, comprising, for example, phosphoramide (Beaucage et al. (1993) Tetrahedron 49:1925) and references therein; Letsinger (1970) J. Org. Chem. 35:3800; Sprinzl et al. (1977) Eur. J. Biochem. 81: 579; Letsinger et al. (1986) Nucl. Acids Res. 14: 3487; Sawai et al. (1984) Chem. Lett. 805, Letsinger et al. (1988) J. Am. Chem. Soc. 110: 4470; and Pauwels et al. (1986) Chemica Scripta 26: 141 9), phosphorothioate (Mag et al. (1991) Nucleic Acids Res. 19:1437; and U.S. Patent No. 5,644,048), phosphorodithioate (Briu et al. (1989) J. Am. Chem. Soc. 111:2321, Omethylphophoroamidite linkages (see Eckstein, Oligonucleotides and Analogues: A Practical Approach, Oxford University Press), and peptide nucleic acid backbones and linkages (see Egholm (1992) J. Am. Chem. Soc. 114:1895; Meier et al. (1992) Chem. Int. Ed. Engl. 31: 1008; Nielsen (1993) Nature, 365: 566; Carlsson et al. (1996) Nature 380: 207). Other analog nucleic acids include those with positive backbones (Denpcy et al. (1995) Proc. Natl. Acad. Sci. USA 92: 6097; non-ionic backbones (U.S. Patent Nos. 5,386,023, 5,637,684, 5,602,240, 5,216,141 and 4,469,863; Angew. (1991) Chem. Intl. Ed. English 30: 423; Letsinger et al. (1988) J. Am. Chem. Soc. 110:4470; Letsinger et al. (1994) Nucleoside & Nucleotide 13:1597; Chapters 2 and 3, ASC Symposium Series 580, "Carbohydrate Modifications in Antisense Research", Ed. Y.S. Sanghui and P. Dan Cook; Mesmaeker et al. (1994), Bioorganic & Medicinal Chem. Lett. 4: 395; Jeffs et al. (1994) J. Biomolecular NMR 34:17; Tetrahedron Lett. 37:743 (1996) and non-ribose backbones, including those described in U.S. Patent Nos. 5,235,033 and 5,034,506, and Chapters 6 and 7, ASC Symposium Series 580, Carbohydrate Modifications in Antisense Research, Ed. Y.S. Sanghui and P. Dan Cook. Nucleic acids containing one or more carbocyclic sugars are also included within the definition of nucleic acids (see Jenkins et al. (1995), Chem. Soc. Rev. pp169-176). Several nucleic acid analogs are described in Rawls, C & E News June 2, 1997 page 35. These modifications of the ribose-phosphate backbone may be done to facilitate the addition of additional moieties such as labels, or to increase the stability and half-life of such molecules

The term "heterologous" as it relates to nucleic acid sequences such as coding sequences and control sequences, denotes sequences that are not normally associated with a

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region of a recombinant construct, and/or are not normally associated with a particular cell. Thus, a "heterologous" region of a nucleic acid construct is an identifiable segment of nucleic acid within or attached to another nucleic acid molecule that is not found in association with the other molecule in nature. For example, a heterologous region of a construct could include a coding sequence flanked by sequences not found in association with the coding sequence in nature. Another example of a heterologous coding sequence is a construct where the coding sequence itself is not found in nature (e.g., synthetic sequences having codons different from the native gene). Similarly, a host cell transformed with a construct which is not normally present in the host cell would be considered heterologous for purposes of this invention.

A "coding sequence" or a sequence which "encodes" a particular polypeptide (e.g. a PKS, an NRPS, etc.), is a nucleic acid sequence which is ultimately transcribed and/or translated into that polypeptide in vitro and/or in vivo when placed under the control of appropriate regulatory sequences. In certain embodiments, the boundaries of the coding sequence are determined by a start codon at the 5' (amino) terminus and a translation stop codon at the 3' (carboxy) terminus. A coding sequence can include, but is not limited to, cDNA from procaryotic or eucaryotic mRNA, genomic DNA sequences from procaryotic or eucaryotic DNA, and even synthetic DNA sequences. In preferred embodiments, a transcription termination sequence will usually be located 3' to the coding sequence.

Expression "control sequences" refers collectively to promoter sequences, ribosome binding sites, polyadenylation signals, transcription termination sequences, upstream regulatory domains, enhancers, and the like, which collectively provide for the transcription and translation of a coding sequence in a host cell. Not all of these control sequences need always be present in a recombinant vector so long as the desired gene is capable of being transcribed and translated.

"Recombination" refers to the reassortment of sections of DNA or RNA sequences between two DNA or RNA molecules. "Homologous recombination" occurs between two DNA molecules which hybridize by virtue of homologous or complementary nucleotide sequences present in each DNA molecule.

The terms "stringent conditions" or "hybridization under stringent conditions" refers to conditions under which a probe will hybridize preferentially to its target subsequence, and to a lesser extent to, or not at all to, other sequences. "Stringent hybridization" and "stringent hybridization wash conditions" in the context of nucleic acid

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hybridization experiments such as Southern and northern hybridizations are sequence dependent, and are different under different environmental parameters. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes part I chapter 2 Overview of principles of hybridization and the strategy of nucleic acid probe assays*, Elsevier, New York. Generally, highly stringent hybridization and wash conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Very stringent conditions are selected to be equal to the T_m for a particular probe.

An example of stringent hybridization conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or northern blot is 50% formamide with 1 mg of heparin at 42°C, with the hybridization being carried out overnight. An example of highly stringent wash conditions is 0.15 M NaCl at 72°C for about 15 minutes. An example of stringent wash conditions is a 0.2x SSC wash at 65°C for 15 minutes (see, Sambrook et al. (1989) Molecular Cloning - A Laboratory Manual (2nd ed.) Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor Press, NY, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1x SSC at 45°C for 15 minutes. An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6x SSC at 40°C for 15 minutes. In general, a signal to noise ratio of 2x (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids which do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides which they encode are substantially identical. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

Expression vectors are defined herein as nucleic acid sequences that are direct the transcription of cloned copies of genes/cDNAs and/or the translation of their mRNAs in an appropriate host. Such vectors can be used to express genes or cDNAs in a variety of hosts such as bacteria, bluegreen algae, plant cells, insect cells and animal cells. Expression vectors include, but are not limited to, cloning vectors, modified cloning vectors, specifically designed plasmids or viruses. Specifically designed vectors allow the shuttling of DNA

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between hosts, such as bacteria-yeast or bacteria-animal cells. An appropriately constructed expression vector preferably contains: an origin of replication for autonomous replication in a host cell, a selectable marker, optionally one or more restriction enzyme sites, optionally one or more constitutive or inducible promoters. In preferred embodiments, an expression vector is a replicable DNA construct in which a DNA sequence encoding a one or more PKS and/or NRPS domains and/or modules is operably linked to suitable control sequences capable of effecting the expression of the products of these synthase and/or synthetases in a suitable host. Control sequences include a transcriptional promoter, an optional operator sequence to control transcription and sequences which control the termination of transcription and translation, and so forth.

The term "conservative substitution" is used in reference to proteins or peptides to reflect amino acid substitutions that do not substantially alter the activity (specificity or binding affinity) of the molecule. Typically conservative amino acid substitutions involve substitution one amino acid for another amino acid with similar chemical properties (e.g. charge or hydrophobicity). The following six groups each contain amino acids that are typical conservative substitutions for one another: 1) Alanine (A), Serine (S), Threonine (T); 2) Aspartic acid (D), Glutamic acid (E); 3) Asparagine (N), Glutamine (Q); 4) Arginine (R), Lysine (K); 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

The "group consisting of ORF-1 through ORF 42" refers to the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 81, ORF 1, ORF 12, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 9, ORF 10, ORF 11, ORF 12, ORF 13, ORF 14, ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42 as identified in Tables I and II. In certain embodiments ORF 9 (cagA) is excluded.

A "biological molecule that is a substrate for a polypeptide encoded by a enediyne (e.g., C-1027) biosynthesis gene" refers to a molecule that is chemically modified by one or more polypeptides encoded by open reading frame(s) of the C-1027 biosynthesis gene cluster. The "substrate" may be a native molecule that typically participates in the biosynthesis of an enediyne, or can be any other molecule that can be similarly acted upon by the polypeptide.

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A "polymorphism" is a variation in the DNA sequence of some members of a species. A polymorphism is thus said to be "allelic," in that, due to the existence of the polymorphism, some members of a species may have the unmutated sequence (*i.e.* the original "allele") whereas other members may have a mutated sequence (*i.e.* the variant or mutant "allele"). In the simplest case, only one mutated sequence may exist, and the polymorphism is said to be diallelic. In the case of diallelic diploid organisms, three genotypes are possible. They can be homozygous for one allele, homozygous for the other allele or heterozygous. In the case of diallelic haploid organisms, they can have one allele or the other, thus only two genotypes are possible. The occurrence of alternative mutations can give rise to trialleleic, *etc.* polymorphisms. An allele may be referred to by the nucleotide(s) that comprise the mutation.

"Single nucleotide polymorphism" or "SNPs are defined by their characteristic attributes. A central attribute of such a polymorphism is that it contains a polymorphic site, "X," most preferably occupied by a single nucleotide, which is the site of the polymorphism's variation (Goelet and Knapp U.S. patent application Ser. No. 08/145,145). Methods of identifying SNPs are well known to those of skill in the art (*see*, *e.g.*, U.S. Patent 5,952,174).

Abbreviations used herein include LB, Luria-Bertani; NGDH, dNDP-glucose 4,6-dehydratase; nt, nucleotide; ORF, open reading frame; PCR, polymerase chain reaction; PEG, polyethyleneglycol; PKS, polyketide synthase; RBS, ribosomal binding site; Apr, apramycin; R, resistant; Th, thiostrepton; WT, wild-type; and TS, temperature sensitive

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates the structures of C-1027 chromophore and the benzenoid diradical intermediate proposed to initiate DNA cleavage.

Figure 2 illustrates a scheme using C-1027 open reading frame polypeptides for the synthesis of deoxysugars.

Figure 3A illustrates a scheme using C-1027 open reading frame polypeptides for the synthesis of a β -amino acid.

Figure 3B illustrates a scheme using C-1027 open reading frame polypeptides for the synthesis of a benzoxazolinate.

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Figure 4 illustrates the synthesis of the enediyne core and final assembly of the C-1027 enediyne.

Figures 5A, 5B, and 5C illustrate the organization of the C-1027 enediyne biosynthetic gene cluster. Figure 5A shows a restriction map of the 75-kb sgc gene cluster from S. globisporus as represented by three cosmid clones. Figure 5B illustrates the genetic organization of the sgcA, sgcB, and cagA genes, showing that they are clustered in the sgc gene cluster. Probe 1, the 0.55-kb dNDP-glucose 4,6-dehydratase gene fragment from pBS1002. Probe 2, the 0.73-kb cagA fragment from pBS1003. A, ApaI; B, BamHI; E, EcoRI; K, KpnI, S, SacII; Sp, SphI. Figure 5C shows the genetic organization of the C-1027 biosynthesis gene cluster.

Figure 6 shows the DNA and deduced amino acid sequences of the 3.0-kb BamHI fragment from pBS1007, showing the sgcA and sgcB genes. Possible RBSs are boxed. The presumed translational start and stop sites are in boldface. Restriction enzyme sites of interest are underlined. The amino acids, according to which the degenerated PCR primer were designed for amplifying the dNDP-glucose 4,6-dehydratase gene from S. globisporus, are underlined.

Figure 7 shows the amino acid sequence alignment of SgcA with three other dNDP-glucose 4,6-dehydratases. Gdh, TDP-glucose 4,6-dehydratase of *S. erythraea* (AAA68211); MtmE, TDP-glucose 4,6-dehydratase in the mithramycin pathway of *S. argillaceus* (CAA71847); TylA2, TDP-glucose 4,6-dehydratase in the tylosin pathway of *S. fradiae* (S49054). Given in parentheses are protein accession numbers. The $\alpha\beta\alpha$ fold with the NAD⁺-binding motif of GxGxxG is boxed.

Figures 8A and 8B show disruption of *sgcA* by single crossover homologous recombination. Figure 8A shows construction of *sgcA* disruption mutant and restriction maps of the wild-type *S. globisporus* C-1027 and *S. globisporus* SB1001 mutant strains showing predicted fragment sizes upon *Bam*HI digestion. Figures 8B and 8C show a Southern analysis of *S. globisporus* C-1027 (lane 1) and *S. globisporus* SB1001 (lanes 2, 3, and 4, three individual isolates) genomic DNA, digested with *Bam*HI, using (Figure 8B) pOJ260 vector or (Figure 8C) the 0.75-kb *Sac*II/*Kpn*I fragment of *sgcA* from pBS1012 as a probe, respectively. B, *Bam*HI; K, *Kpn*I; S, *Sac*II.

Figures 9A, 9B, 9B, and 9D illustrate the determination of C-1027 production in various *S. globisporus* strains by assaying their antibacterial activity against *M. luteus*.

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Figure 9A:1, *S. globisporus*C-1027; 2,3, and 4, *S. globisporus* SB1001 (three individual isolates); 5, *S. globisporus* AF67; 6, *S. globisporus* AF40. Figure 9B: 1, *S. globisporus* C-1027; 2, *S. globisporus* SB1001 (pWHM3); 3 and 4, *S. globisporus* SB1001 (pBS1015) (two individual isolates). Both *S. globisporus* SB1001 (pWHM3) and *S. globisporus* SB1001 (pBS1015) were grown in the presence of 5 μg/mL thiostrepton. Figure 9C: 1, *S. globisporus* C-1027; 2, *S. globisporus* SB1001 (pBS1015); 3. *S. globisporus* SB1001; 4. *S. globisporus* SB1001 (pWHM3); 5. *S. globisporus* AF40; 6. *S. globisporus* AF44. All *S. globisporus* strains were grown in the absence of thiostrepton. Figure 9D: 1. *S. globisporus* (pKC1139); 2. *S. globisporus* (pBS1018).

DETAILED DESCRIPTION

This invention provides a complete gene cluster regulating the biosynthesis of C-1027, the most potent member of the enediyne antitumor antibiotic family. C-1027 is produced by *Streptomyces globisporus* C-1027 and consists of an apoprotein (encoded by the *cagA* gene) and a non-peptidic chromophore. The C-1027 chromophore could be viewed as being derived biosynthetically from a benzoxazolinate, a deoxyamino hexose, a β-amino acid, and an enediyne core. Adopting a strategy to clone the C-1027 biosynthesis gene cluster by mapping a putative dNDP-glucose 4,6-dehydratase (NGDH) gene to *cagA*, we localized 75 kb contiguous DNA from *S. globisporus* encoding a complete C-1027 gene cluster.

Initial sequencing of the cloned gene cluster revealed two genes, sgcA and sgcB, that encode an NGDH enzyme and a transmembrane efflux protein, respectively, and confirmed that the cagA gene resides approximately 14 kb upstream of the sgcA, B locus. The involvement of the cloned gene cluster in C-1027 biosynthesis was demonstrated by disrupting the sgcA gene to generate C-1027-nonproducing mutants and by complementing the sgcA mutants in vivo to restore C-1027 production.

Subsequent DNA sequence analysis provided the complete enediyne C-1027 gene cluster sequence (SEQ ID NOs: 1 and 2) revealing 50 open reading frames which are summarized in Tables I and II. These results represent the first cloning of a gene cluster for enediyne anti-tumor antibiotic biosynthesis.

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Table I. Summary of the C-1027 gene cluster open reading frames. Table 1. C-1027 gene cluster open reading frames (-7 to 26), primers for ORF amplification, and proposed functions

orf#	Size	Relative position			Prin	ners			Function	Seq ID No.
orf-	648	658-11	Fwd:	ATG	GGC	ATG	ACG	GGT	very weak	3
(-7)	bp		Rev:						homology to putative hydroxylase	4
orf-	549	1478-	Fwd:	ATG	CCG	CGG	ATT	CCC	Viral	5
(-6)	pp	930	Rev:	TCA	GCT	GTC	GAT	GTC	infectivity potentiator protein	6
orf-	1065	2713-	Fwd:	ATG	ACC	ATC	GCC	ACT	N-truncated	7
(-5)	bp	1649	Rev:	TCA	GAG	GCC	GAG	CAC	Methionine synthase (likely psuedogene)	8
orf-	387	3238-	Fwd:	ATG	AGC	TCG	CTA	CTG	Viral	9
(-4)	bp	2851	Rev:						transcription factor	10
orf-	1530	4971-	Fwd:						Viral Homolog	11
(-3)	bp	3442	Rev:						possibly primase	12
orf- (-2)	3027 bp	5982- 7478	Fwd: Rev:						Glycerol- Phosphate ABC Transporter (SnoX drug resistance)	13 14
orf-	2328	9900-	Fwd:						UvrA-like drug	15
(-1)	pp	7573	Rev:	TCA	ACC	CGC	CCT	GCG	resistance pump	16
orf- 0	1368 bp	11349- 9982	Fwd: Rev:						Na ⁺ /H ⁺ efflux pump	17 18
orf-	999	28590-	Fwd:	ATG	AGG	ATG	CTG	GTG	dNTP-glucose	19
1	bp	29588	Rev:	TCA	GCC	GAC	GGC	GTC	dehydratase	20
orf- 2	1566 bp	29632- 31197	Fwd: Rev:						Transmembrane efflux protein	21 22
orf-	1311	31280-	Fwd:	GTG	GAG	TAC	TGG	AAC	Coenzyme F390	23
3	bp	32590	Rev:						synthase phenylacetyl-CoA ligase	24
orf-	1584	32809-	Fwd:	GTG	CCC	CAC	GGT	GCA	phenol	25
4	bp	34392	Rev:	CTA	CAG	CCC	TCC	GAG	hydroxylase chlorophenol-4- monooxygenase	26
orf-	bp	35274-	Fwd:						citrate	27
5		34458	Rev:						transport protein	28
orf-	1272	17924-	Fwd:						C-methyl	29
6		16653	Rev:	TCA	GAC	GAG	CTC	CTT	transferase hydroxylase	30
orf-	735	16653-	Fwd:	ATG	GAG	TAC	GGC	CCC	N -	31

7	bp	15919	Rev: TCA TGC CGT GCG CAC methyltransferas e	32
orf-	1233	15922-	Fwd: ATG AGC GGC GGC CCG Aminotransferase	33
8	bp	14690	Rev: TCA CCT CGC CGG ACG	34
	<u>-</u> -			
orf-	432	14643-	Fwd: ATG TCG TTA CGT CAC CagA	35
9	bp	14212	Rev: TCA GCC GAA GGT CAG	36
_			The same of the sa	2.5
orf-	1068	13012-	Fwd: ATG AAG GCA CTT GTA dNTP-glucose Rev: TCA GGC CGC GAT CTC synthase	37 38
10	bp	14079	Rev: ICA GGC CGC GAI CIC Synthase	30
orf-	1485	12835-	Fwd: GTG GAC GTG TCA GCG Hydroxylase,	39
11	bp	11351	Rev: TCA GGA CCG CGC ACC Halogenase	40
orf-	579	25564-	Fwd: ATG AAG CCG ATC GGG dNTP-4-keto-6-	41
12	bp	24986	Rev: TCAGGA CGA CTT GTT deoxyglucose	42
onf	1127	24702	3,5-epimerase Fwd: ATG CCT TCC CCC TTC 3-0-	43
orf <i>-</i> 13	1137 bp	24702- 23566	Rev: TCA GGT GCG CTC GGC acyltransferase	44
13	υp	23300	Nev. 16A 001 000 CTC 000 acyteramprerape	
orf-	1455	22878-	Fwd: GTG AGA GAC GGC CGG Coenzyme F-390	45
14	bp	21424	Rev:TCA CGT GGT GAT GGC Synthase	46
			Phenylacetyl CoA	
_		0.1.4.0.	Ligase	47
orf-	1482	21407-	Fwd: ATG ACC GAC CAG TGC Anthranilate Rev: TCA CAG CAA CTC CTC Synthase I	47 48
15	bp	19926	Rev: ICA CAG CAA CIC CIC Synthase I	40
orf-	663	19929-	Fwd: GTG AGC TTG TGG TCT Anthranilate	49
16	bp	19267	Rev: TCA GGC CGG TTC GGC Synthase II	50
orf-	1161	19191-	Fwd: GTG CGT CCC TTC CGT epoxide	51
17	pp	18031	Rev: TCA GCG GAG CGG ACG hydrolase	52
orf-	423	35938-	Fwd: ATG CCA GCA CCG ACT Unknown	53
18	bp	35516	Rev: TCA GTC GTT GCC GCG	54
	-			
orf-	1380	27214-	Fwd: ATG CGG GTG ATG ATC glycosyl	55
19	bp	28593	Rev: TCA TCG GTC CGC CTC transferase	56
orf	1356	25815-	Fwd: ATG ACC AAG CAC GCC squalene	57
orf- 20	pb T320	27170	Rev: TCA TAC GGC GGC GCC monooxygenase	58
20	Dp.	21270	NOV. 1011 4110 000 000 000 messa 22, 3 assess	
orf-	672	23546-	Fwd: GTG AGC GCA CAA CTC hypothetical Fe-	59
21	bp	22875	Rev: TCA CGG CTG TGC CTG S flavoprotein	60
_	0.1.	25051	The A THE	61
orf-	816	35274-	Fwd: ATG TCT TCA ACC CGT haloacetate	61 62
22	pp	34458	Rev: TCA GCC GCG CAG GAA dehalogenase hydrolase	UZ
orf-	1380	37559-	Fwd: ATG ACG ACG TCC GAC peptide	63
23	bp	38938	Rev: TCA GGA GGT GAA GGG synthetase	64
	-			
orf-	1620	40986-	Fwd: ATG GCA TTG ACT CAA Histidine	65
24	bp	39367	Rev: TCA GCG CAG CTG GAT Ammonia lyase	66
orf-	1560	42611-	Fwd: ATG ACG CGG CCG GTG Type II	67
25	bp	41052	Rev: TCA GCG GGT GAG CCG adenylation	68
="	-	-	protein	
orf-	282	38983-	Fwd: GTG TCC ACC GTT TCC Type II peptidyl	
26	bp	39264	Rev: TCA CTG CGT TCC GGA carrier protein	70

Table II. C-1027 gene cluster open reading frames (27 to 42), primers for ORF amplification, and proposed functions

ORF	Relative	Primers	Function	SEQ
	Position			ID
				NO.
orf-	43945-46023	Fwd: GTG TGC CCG GTG ACA GAC	Antibiotic	71
27		Rev: TCA GCC CAC GGG CTG GGA	Transporter	72
orf-	46167-47171	Fwd: GTG TTG GGC GAT GAG GAC	O-	73
28		Rev: TCA GAC CGC GGA CAT CTG	methyltransfer ase	74
orf	47227-48485	Fwd: ATG GCC GGC CTG GTC ATG	p450	75
29	4,22, 40403	Rev: TCA GGA CCC GAG GGT CAC	hydroxylase	76
orf-	48610-49714	Fwd: GTG GAC CAG ACG TCT ACG	Oxidoreductase	77
30		Rev: TCA TGC AGG TGC AGC GTG		78
orf-	50350-51390	Fwd: ATG AGG CCG CTC GTT CGG	Unknown	79
31		Rev: TCA TCC CGG CCC GGC GGC	Protein	80
orf-	51420-52341	Fwd: ATG AGA ACG CGG CGA CGC	Oxidoreductase	81
32		Rev: TCA CGG CCG GAG GCG TAC		82
orf-	53241-54074	Fwd: GTG TAT CAG CCG GAC TGT	Unknown	83
33		Rev: CTA CTC ATT CCA GTT GTG	Protein	84
orf-	54230-55379	Fwd: ATG TCT ACG GGC TAT CTC	Unknown	85
34		Rev: TCA GCC GCC GGT GGC GCC	Protein	86
orf-	56027-56881	Fwd: ATG TTC TCC CCC GCC GCC	Oxidase/	87
35		Rev: TCA GTA CGC CTG GTG GGC	Dehydrogenase	88
orf-	56928-57730	Fwd: ATG AAT TCG CTC GAC GAC	Unknown	89
36		Rev : TCA GCT CCC GGT CGC CGC	Protein	90
orf-	57834-58304	Fwd: ATG ACC GCG ACG AAT CCT	Regulatory	91
37_		Rev: CTA GGC GGC GCG TCC CGC		92
orf-	58440-60091	Fwd: ATG AGC ACC ACG GCC GAG	Oxidoreductase	93
38		Rev: TCA GCC GCG CGC CGA CGG	D 7.4	94
orf-	60092-60622	Fwd: ATG ACC CTG GAG GCC TAC	Regulatory	95
39		Rev: TCA TGC GGG GCT CCC GGT	D7-E	96
orf-	60940-62020	Fwd: GTG AAA AGT GAC TCT GCC	Regulatory	97
40		Rev: TCA ACG GCG AGT TGG CTG	-	98
orf-	62045-62899	Fwd: GTG ACC ACG AAC ACC ATC	Regulatory	99
41		Rev: TCA CCC GCG ATC TCG ATC		100
orf-	62788-63164	-	p450	101
42		Rev: TCA CCT CGC CGT ACT CAC	hydroxylase	102

Surprisingly, sequence analysis failed to reveal any gene that resembles a polyketide synthase. The C-1027 open reading frames, however, encode polypeptides exhibiting a wide variety of enzymatic activities (*e.g.*, epoxide hydrase, monooxygenase, oxidoreductase, P-450 hydroxylase, *etc.*). The isolated C-1027 gene cluster can be used to

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synthesize C-1027 enediyne antibiotics and/or analogues thereof. The C-1027 gene cluster can be modified and/or augmented to increase C-1027 and/or C-1027 analogue production.

Alternatively, various components of the C-1027 gene cluster can be used to synthesize and/or chemically modify a wide variety of metabolites. Thus, for example, ORF 6 (C-methyltransferase) can be used to methylate a carbon, while ORF 12, an epimerase, can be used to change the conformation of a sugar. The ORFs can be combined in their native configuration or in modified configurations to synthesize a wide variety of biomolecules/metabolites. Thus, for example, various combinations of C-1027 open reading frames can be used to synthesize an enediyne core, to synthesize a deoxy sugar, to synthesize a β-amino acid, to make a benzoxazolinate, *etc* (*see, e.g.*, Figures 2, 3, and 4).

The native C-1027 gene cluster ORFs can be re-ordered, modified, and combined with other biosynthetic units (*e.g.* polyketide synthases (PKSs) or catalytic domains thereof and/or non-ribosomal polypeptide synthetases (NRPSs) or catalytic domains thereof) to produce a wide variety of molecules. Large chemical libraries can be produced and then screened for a desired activity.

The C-1027 gene cluster also includes a number of drug resistance genes (*see*, *e.g.*, Table 2) that confer resistance to C-1027 and/or metabolites involved in C-1027 biosynthesis thereby permitting the cell to complete the enediyne biosynthesis. These resistance genes can be used to confer enediyne resistance on a cell lacking such resistance or to augment the enediyne resistance of a cell that does tolerate enediynes. Such cells can be used to produce high levels of enediynes and/or enediyne metabolites, and/or enediyne analogues.

Table III. C-1027 cluster drug resistance genes.

ORF	Protein	Mechanism
ORF 9:	CagA apoprotein	Drug sequestering
ORF 2:	SgcB transmembrane efflux protein	Drug exporting
ORF 27	Transmembrane transport protein	Drug exporting
ORF 0	Na ⁺ /H ⁺ transporter	Drug exporting
ORF -1	ABC transport (C-terminus)	Drug exporting
ORF -2	Glycerol phosphate transporter	Drug exporting
ORF -1	UvrA-like protein (N-terminus)	DNA repairing

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I. Isolation, preparation, and expression of C-1027 nucleic acids.

The C-1027 gene cluster nucleic acids can be isolated, optionally modified, and inserted into a host cell to create and/or modify a metabolic (biosynthetic) pathway and thereby enable that host cell to synthesize and/or modify various metabolites. Alternatively the C-1027 gene cluster nucleic acids can be expressed in the host cell and the encoded C-1027 polypeptide(s) recovered for use as chemical reagents, *e.g.* in the *ex vivo* synthesis and/or chemical modification of various metabolites. Either application typically entails insertion of one or more nucleic acids encoding one or more isolated and/or modified C-1027 enediyne open reading frames in a suitable host cell. The nucleic acid(s) are typically in an expression vector, a construct containing control elements suitable to direct expression of the C-1027 polypeptides. The expressed C-1027 polypeptides in the host cell then act as components of a metabolic/biosynthetic pathway (in which case the synthetic product of the pathway is typically recovered) or the C-1027 polypeptides themselves are recovered. Using the sequence information provided herein, cloning and expression of C-1027 nucleic acids can be accomplished using routine and well known methods.

A) C-1027 nucleic acids.

The nucleic acids comprising the C-1027 gene cluster are identified in Tables I and are listed in the sequence listing provided herein. In particular, Table 1 identifies genes and functions of open reading frames (ORFs) in the C-1027 enedigne biosynthesis gene cluster and identifies primers suitable for the amplification/isolation of any one or more of the C-1027 open reading frames. Of course, using the sequence information provided herein, other primers suitable for amplification/isolation of one or more C-1027 open reading frames can be determined according to standard methods well known to those of skill in the art (*e.g.* using Vector NTI SuiteTM, InforMax, Gaithersberg, MD, USA).

Typically such amplifications will utilize the DNA or RNA of an organism containing the requisite genes (e.g. Streptomyces globisporus) as a template. Typical amplification conditions include the following PCR temperature program: initial denaturing at 94°C for 5 min, 24-36 cycles of 45 sec at 94°C, 1 min at 60°C, 2 min at 72°C, followed by additional 7 min at 72°C. One of skill will appreciate that optimization of such a protocol, e.g. to improve yield, etc. is routine (see, e.g., U.S. Patent No. 4,683,202; Innis (1990) PCR

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Protocols A Guide to Methods and Applications Academic Press Inc. San Diego, CA, etc). In addition, primer may be designed to introduce restriction sites and so facilitate cloning of the amplified sequence into a vector.

In one embodiment, this invention provides nucleic acids for the recombinant expression of an enediyne (e.g. a C-1027 enediyne or an analogue thereof). Such nucleic acids include isolated gene cluster(s) comprising open reading frames encoding polypeptides sufficient to direct the assembly of the enediyne. In other embodiments of this invention, the C-1027 open reading frames may be unchanged, but the control elements (e.g. promoters, enhancers, etc.) may be modified. In still other embodiments, the nucleic acids may encode selected components (e.g. one or more C-1027 or modified C-1027 open reading frames) and/or may optionally contain other heterologous biosynthetic elements including, but not limited to polyketide synthase (PKS) and/or non-ribosomal polypeptide synthetase (NRPS) modules or enzymatic domains.

Such variations may be introduced by design, for example to modify a known molecule in a specific way, *e.g.* by replacing a single substituent of the enedigne with another, thereby creating a derivative enedigne molecule of predicted structure.

Alternatively, variations can be made randomly, for example by making a library of molecular variants of a known enedigne by systematically or haphazardly replacing one or open reading frames in the biosynthetic pathway. Production of alternative/modified enedigne, and hybrid enedigne PKSs and/or NRPSs and hybrid systems is described below.

Using the information provided herein other approaches to cloning the desired sequences will be apparent to those of skill in the art. For example, the enediyne, and/or optionally PKS and/or NRPS modules or enzymatic domains of interest can be obtained from an organism that expresses such, using recombinant methods, such as by screening cDNA or genomic libraries, derived from cells expressing the gene, or by deriving the gene from a vector known to include the same. The gene can then be isolated and combined with other desired biosynthetic elements using standard techniques. If the gene in question is already present in a suitable expression vector, it can be combined *in situ*, with, e.g., other PKS subunits, as desired. The gene of interest can also be produced synthetically, rather than cloned. The nucleotide sequence can be designed with the appropriate codons for the particular amino acid sequence desired. In general, one will select preferred codons for the intended host in which the sequence will be expressed. The complete sequence can be assembled from overlapping oligonucleotides prepared by standard methods and assembled

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into a complete coding sequence (see, e.g., Edge (1981) Nature 292:756; Nambair et al. (1984) Science 223: 1299; Jay et al. (1984) J. Biol. Chem. 259:6311). In addition, it is noted that custom gene synthesis is commercially available (see, e.g. Operon Technologies, Alameda, CA).

Examples of such techniques and instructions sufficient to direct persons of skill through many cloning exercises are found in Berger and Kimmel (1989) *Guide to Molecular Cloning Techniques, Methods in Enzymology 152* Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.* (1989) *Molecular Cloning - A Laboratory Manual* (2nd ed.) Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor Press, NY; Ausubel (19 1994) *Current Protocols in Molecular Biology*, Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., U.S. Patent 5,017,478; and European Patent No. 0,246,864.

B) Expression of f C-1027 open reading frames.

The choice of expression vector depends on the sequence(s) that are to be expressed. Any transducible cloning vector can be used as a cloning vector for the nucleic acid constructs of this invention. However, where large clusters are to be expressed, it phagemids, cosmids, P1s, YACs, BACs, PACs, HACs or similar cloning vectors be used for cloning the nucleotide sequences into the host cell. Phagemids, cosmids, and BACs, for example, are advantageous vectors due to the ability to insert and stably propagate therein larger fragments of DNA than in M13 phage and lambda phage, respectively. Phagemids which will find use in this method generally include hybrids between plasmids and filamentous phage cloning vehicles. Cosmids which will find use in this method generally include lambda phage-based vectors into which cos sites have been inserted. Recipient pool cloning vectors can be any suitable plasmid. The cloning vectors into which pools of mutants are inserted may be identical or may be constructed to harbor and express different genetic markers (see, *e.g.*, Sambrook *et al.*, *supra*). The utility of employing such vectors having different marker genes may be exploited to facilitate a determination of successful transduction.

In preferred embodiments of this invention, vectors are used to introduce C-1027 biosynthesis genes or gene clusters into host (*e.g. Streptomyces*) cells. Numerous vectors for use in particular host cells are well known to those of skill in the art. For example described in Malpartida and Hopwook, (1984) *Nature*, 309:462-464; Kao *et al.*,

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(1994), Science, 265: 509-512; and Hopwood et al., (1987) Methods Enzymol., 153:116-166 all describe vectors for use in various Streptomyces hosts.

In one preferred embodiment, *Streptomyces* vectors are used that include sequences that allow their introduction and maintenance in *E. coli*. Such *Streptomyces/E. coli* shuttle vectors have been described (*see*, for example, Vara *et al.*, (1989) *J. Bacteriol.*, 171:5872-5881; Guilfoile & Hutchinson (1991) *Proc. Natl. Acad. Sci. USA*, 88: 8553-8557.)

The wildtype and/or modified C-1027 enediyne open reading frame(s) of this invention, can be inserted into one or more expression vectors, using methods known to those of skill in the art. Expression vectors will include control sequences operably linked to the desired open reading frame. Suitable expression systems for use with the present invention include systems that function in eucaryotic and/or prokaryotic host cells. However, as explained above, prokaryotic systems are preferred, and in particular, systems compatible with Streptomyces spp. are of particular interest. Control elements for use in such systems include promoters, optionally containing operator sequences, and ribosome binding sites. Particularly useful promoters include control sequences derived from enediyne, and/or PKS, and/or NRPS gene clusters. Other promoters (e.g. ermE* as illustrated in Example 1) are also suitable. Other bacterial promoters, such as those derived from sugar metabolizing enzymes, such as galactose, lactose (lac) and maltose, will also find use in the present constructs. Additional examples include promoter sequences derived from biosynthetic enzymes such as tryptophan (trp), the beta -lactamase (bla) promoter system, bacteriophage lambda PL, and T5. In addition, synthetic promoters, such as the tac promoter (U.S. Patent 4,551,433), which do not occur in nature also function in bacterial host cells. In Streptomyces, numerous promoters have been described including constitutive promoters, such as ErmE and TcmG (Shen and Hutchinson, (1994) J. Biol. Chem. 269: 30726-30733), as well as controllable promoters such as actI and actIII (Pleper et al., (1995) Nature, vol. 378: 263-266; Pieper et al., (1995) J. Am. Chem. Soc., 117: 11373-11374; and Wiesmann et al., (1995) Chem. & Biol. 2: 583-589).

Other regulatory sequences may also be desirable which allow for regulation of expression of the enediyne open reading frame(s) relative to the growth of the host cell. Regulatory sequences are known to those of skill in the art, and examples include those which cause the expression of a gene to be turned on or off in response to a chemical or physical stimulus, including the presence of a regulatory compound. Other types of regulatory elements may also be present in the vector, for example, enhancer sequences.

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Selectable markers can also be included in the recombinant expression vectors. A variety of markers are known which are useful in selecting for transformed cell lines and generally comprise a gene whose expression confers a selectable phenotype on transformed cells when the cells are grown in an appropriate selective medium. Such markers include, for example, genes that confer antibiotic resistance or sensitivity to the plasmid.

The various enediyne cluster open reading frames, and/or PKS, and/or NRPS clusters or subunits of interest can be cloned into one or more recombinant vectors as individual cassettes, with separate control elements, or under the control of, *e.g.*, a single promoter. The various open reading frames can include flanking restriction sites to allow for the easy deletion and insertion of other open reading frames so that hybrid synthetic pathways can be generated. The design of such unique restriction sites is known to those of skill in the art and can be accomplished using the techniques described above, such as site-directed mutagenesis and PCR.

Methods of cloning and expressing large nucleic acids such as gene clusters, including PKS- or NRPS-encoding gene clusters, in cells including *Streptomyces* are well known to those of skill in the art (*see, e.g.*, Stutzman-Engwall and Hutchinson (1989) *Proc. Natl. Acad. Sci. USA*, 86: 3135-3139; Motamedi and Hutchinson (1987) *Proc. Natl. Acad. Sci. USA*, 84: 4445-4449; Grim *et al.* (1994) *Gene*, 151: 1-10; Kao *et al.* (1994) *Science*, 265: 509-512; and Hopwood *et al.* (1987) *Meth. Enzymol.*, 153: 116-166). In some examples, nucleic acid sequences of well over 100kb have been introduced into cells, including prokaryotic cells, using vector-based methods (*see*, for example, Osoegawa *et al.*, (1998) *Genomics*, 52: 1-8; Woon *et al.*, (1998) *Genomics*, 50: 306-316; Huang *et al.*, (1996) *Nucl. Acids Res.*, 24: 4202-4209). In addition, the cloning and expression of C-1027 enediyne is illustrated in Example 1.

C) Host cells.

The vectors described above can be used to express various protein components of the enediyne, and/or enediyne shunt metabolites, and/or other modified metabolites for subsequent isolation and/or to provide a biological synthesis of one or more desired biomolecules (e.g. C-1027 and/or a C-1027 analogue, etc.). Where one or more proteins of the enediyne biosynthetic gene cluster are expressed (e.g. overexpressed) for subsequent isolation and/or characterization, the proteins are expressed in any prokaryotic or

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eukaryotic cell suitable for protein expression. In one preferred embodiment, the proteins are expressed in *E. coli*.

Host cells for the recombinant production of the subject enediynes, enediyne metabolites, shunt metabolites, etc. can be derived from any organism with the capability of harboring a recombinant enediyne gene cluster and/or subset thereof. Thus, the host cells of the present invention can be derived from either prokaryotic or eucaryotic organisms. Preferred host cells are those of species or strains (e.g. bacterial strains) that naturally express enedignes. Such host cells include, but are not limited to Actinomycetes, Actinoplanetes, and Streptomycetes, Actinomadura, Micromonospra, and the like. Particularly preferred host cells include, but are not limited to Streptomyces globisporus, Streptomyces lividans, Streptomyces coelicolor, Micromonospora echinospora spp. calichenisis, Actinomadura verrucosopora, Micromonospora chersina, Streptomyces carzinostaticus, and Actinomycete L585-6. Other suitable host cells include, but are not limited to S. verticillis S. ambofaciens, S. avermitilis, S. azureus, S. cinnamonensis, S. coelicolor, S. curacoi, S. erythraeus, S. fradiae, S. galilaeus, S. glaucescens, S. hygroscopicus, S. lividans, S. parvulus, S. peucetius, S. rimosus, S. roseofulvus, S. thermotolerans, and S. violaceoruber (see, e.g., Hopwood and Sherman (1990) Ann. Rev. Genet. 24: 37-66; O'Hagan (1991) The Polyketide Metabolites, Ellis Horwood Limited, etc.).

In certain embodiments, a eukaryotic host cell is preferred (e.g. where certain glycosylation patterns are desired). Suitable eukaryotic host cells are well known to those of skill in the art. Such eukaryotic cells include, but are not limited to yeast cells, insect cells, plant cells, fungal cells, and various mammalian cells (e.g. COS, CHO HeLa cells lines and various myeloma cell lines).

D) Recovery of the expression product.

Recovery of the expression product (e.g., enediyne, enediyne analogue, enediyne biosynthetic pathway polypeptide, etc.) is accomplished according to standard methods well known to those of skill in the art. Thus, for example where enediyne biosynthetic gene cluster proteins are to be expressed and isolated, the proteins can be expressed with a convenient tag to facilitate isolation (e.g. a His₆) tag. Other standard protein purification techniques are suitable and well known to those of skill in the art (see,

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e.g., (Quadri et al. (1998) Biochemistry 37: 1585-1595; Nakano et al. (1992) Mol. Gen. Genet. 232: 313-321, etc.).

Similarly where components (e.g. enediyne biosynthetic cluster orfs) are used to synthesize and/or modify various biomolecules (e.g. enediynes, enediyne analogues, shunt metabolites, etc.) the desired product and/or shunt metabolite(s) are isolated according to standard methods well know to those of skill in the art (see, e.g., Carreras and Khosla (1998) Biochemistry 37: 2084-2088, Deutscher (1990) Methods in Enzymology Volume 182: Guide to Protein Purification, M. Deutscher, ed. etc.).

II. Use of C-1027 open reading frames in directed biosynthesis.

Elements (e.g. open reading frames) of the C-1027 biosynthetic gene cluster and/or variants thereof can be used in a wide variety of "directed" biosynthetic processes (i.e. where the process is designed to modify and/or synthesize one or more particular preselected metabolite(s)). Essentially the entire C-1027 gene cluster can be used to synthesize a C-1027 enediyne and/or a C-1027 enediyne analogue. Individual C-1027 cluster open reading frames can be used to perform chemically modifications on particular substrates and/or to synthesize various metabolites. Thus, for example, ORF 6 (C-methyltransferase can be used to methylate a carbon), while ORF 7 (N-methyltransferase) can be used to methylate a nitrogen. ORF 12, and epimerase, can be used to change the conformation of a sugar, and ORF 8 (an amino transferase) can be used to aminate a suitable substrate. Similarly, combinations of C-1027 open reading frames can be used to direct the synthesis of various metabolites (e.g. β-amino acids, deoxysugars, benzoxazolinates, and the like). These examples, are merely illustrative. One of skill in the art, utilizing the information provided here, can perform literally countless chemical modifications and/or syntheses using either "native" enediyne biosynthesis metabolites as the substrate molecule, or other molecules capable of acting as substrates for the particular enzymes in question. Other substrates can be identified by routine screening. Methods of screening enzymes for specific activity against particular substrates are well known to those of skill in the art.

The biosyntheses can be performed *in vivo*, *e.g.* by providing a host cell comprising the desired C-1027 gene cluster open reading frames and/or *in vivo*, *e.g.*, by providing the polypeptides encoded by the C-1027 gene cluster ORFs and the appropriate substrates and/or cofactors.

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A) Synthesis of enedignes and enedigne analogues.

In one embodiment, this invention provides for the synthesis of C-1027 enediynes and/or C-1027 analogues or derivatives. In a preferred embodiment, this is accomplished by providing a cell comprising a C-1027 gene cluster and culturing the cell under conditions whereby the desired enediyne or enediyne analogue is synthesized. The cell can be a cell that does not normally synthesize an enediyne and the entire gene cluster can be transfected into the cell. Alternatively, a cell that typically synthesizes enediynes can be utilized and all or part of the C-1027 gene cluster can be introduced into the cell.

Enediyne derivatives/analogues can be produced by varying the order of, or kind of, gene cluster subunits present in the cell, and/or by changing the host cell (e.g. to a eukaryotic cell that glycosylates the biosynthetic product), and/or by providing altered metabolites (e.g. adding exogenous aglycones to a host that carries a gene cassette of the deoxysugar biosynthesis and glycosylation genes for the production of glycosylated metabolites), etc.

In certain embodiments, the host cell need not be transfected with an entire C-1027 gene cluster. Rather, various components of a C-1027 gene cluster can be altered within a cell already harboring a C-1027 cluster. By varying or adding various biosynthetic open reading frames, C-1027 enedigne variants can be produced.

The use of standard techniques of molecular biology (gene disruption, gene replacement, gene supplement) can be used to modulate and/or otherwise alter enediyne and/or other metabolite (e.g. shunt metabolite) production in an organism that naturally synthesizes an enediyne (e.g. S. globisporus) or an organism that is modified to synthesize an enediyne.

In addition, or alternatively, control sequences that alter the expression of various open reading frames can be introduced that alter the amount and/or timing of enediyne production. Thus, for example, by placing particular C-1027 open reading frames under control of a constitutive promoter (*ermE**) C-1027 production was increased by as much as 4-fold (*see, e.g.* Table 3 and Example 1).

Table 3. Alteration of C-1027 production by engineering the C-1027 biosynthesis gene cluster.

Strain Yield (%)

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WT	100	
WT/pKC1139	100	
WT/ermE*/ORF 2	>150	
WT/ORF 9	>100	
WT/ermE*/ORF 9	<10	
WT/ORF 10, 11	>100	
WT/ermE*/ORF 10, 11	>100	
WT/ ORF 9, 10, 11	>400	

ORF 2: transmembrane eflux protein; ORF 9: CagA apoprotein; ORF 10: TDP-glucose synthase; ORF 11; Hydroxylase/halogenase

Where enediyne analogues are synthesized, it will often prove desirable to assay them for biological activity. Such assays are well know to those of skill in the art. One such assay is illustrated in Example 1. Briefly, this example depicts an assay of antibacterial activity against *M. luteus* as described by Hu *et al.* (1988) *J. Antibiot.* 41: 1575-1579). Other suitable assays for enediyne activity will be known to those of skill in the art.

B) Use of C-1027 open reading frames to synthesize an enediyne core.

The C-1027 open reading frames described herein, or variants thereof, can be used to synthesize an enediyene core, *e.g.*, from a fatty acid precursor. One such synthetic pathway is illustrated in Figure 4. This reaction scheme utilizes ORF 17 (epoxide hydrase), ORF 20 (monooxygenase), ORF 21 (iron-sulfur flavoprotein), ORF 29 (P-450 hydroxylase, ORF 30 (oxidoreductase), ORF 32 (oxidoreductase), ORF 35 (proline oxidase), and ORF 38 (P-450 hydroxylase) to synthesize anenediyne core.

This synthetic pathway, is not considered limiting, but merely illustrative. Using this as a model, one of ordinary skill in the art can design numerous other synthetic schemes to produce enediyne cores and/or core variants.

C) Use of C-1027 open reading frames to synthesize deoxy sugars.

The biosynthesis of various deoxy sugars (e.g., deoxyhexoses) typically share a common key intermediate --4-keto-6-deoxyglucose nucleoside diphosphate or its analogs, whose formation from glucose nucleoside diphosphate is catalyzed by the NGDH enzyme, an NAD⁺-dependent oxidoreductase (Liu and Thorson (1994) *Ann. Rev. Microbiol.* 48: 223-256; Piepersberg (1997) pp. 81-163. In *Biotechnology of antibiotics*, 2nd ed. W. R. Strohl

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(ed). Marcel Dekker, New York.). Similarly, the C-1027 gene cluster includes an NAGDH enzyme which can be exploited to synthesize a variety of deoxy sugars.

One illustrative synthetic pathway is shown in Figure 2. This biosynthetic scheme utilizes ORF 10 (dNDP-glucose synthase), ORF 1 (dNDP-glucose dehydratase), ORF 12 (epimerase), ORF 8 (aminotransferase), ORF 6 (C-methyltransferase), ORF 7 (N-methyltransferase) and ORF 19 (glycosyl transferase).

This synthetic pathway, is not considered limiting, but merely illustrative. Using this as a model, one of ordinary skill in the art can design numerous other synthetic schemes to produce various deoxy sugars.

D) Use of C-1027 open reading frames to synthesize β -amino acids.

In still another embodiment, C-1027 biosynthetic polypeptides can be used in the biosynthesis of β-amino acids. One illustrative synthetic pathway is shown in Figure 3A. This biosynthetic scheme utilizes ORF 4 (hydroxylase), ORF 11 (hydroxylase/halogenase), ORF 24 (aminomutase), ORF 23 (type II NRPS condensation enzyme), ORF 25 (type II NRPS adenylation enzyme), and ORF 26 (type II peptidyl carrier protein).

Again, this synthetic pathway, is not considered limiting, but merely illustrative. Using this as a model, one of ordinary skill in the art can design numerous other synthetic schemes to produce other beta amino acids.

E) Use of C-1027 open reading frames to synthesize benzoxazolinates.

The C-1027 open reading frames can also be used to synthesize a benzoxazolinate. One illustrative synthetic pathway is shown in Figure 3B. This biosynthetic scheme utilizes ORF 15 (anthranilate synthase I, ORF 16 (anthranilate synthase II), ORF 4 (phenol hydroxylase/chlorophenol-4-monooxygenase), ORF 11 (Hydroxylase/Halogenase), ORF 28 (O-methyltransferase), ORF 3 (coenzyme F390 synthetase, ORF 14 (coenzyme F390 synthetase), and ORF 13 (O-acyltransferase). Again, this synthetic pathway, is not considered limiting, but merely illustrative. Using this as a model, one of ordinary skill in the art can design numerous other synthetic schemes to produce other beta amino acids.

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III. Generation of chemical diversity.

In addition to the directed modification and/or biosynthesis of various metabolites as described above, the C-1027 biosynthetic gene cluster open reading frames can be utilized, by themselves or in combination with other biosynthetic subunits (*e.g.* NRPS and/or PKS modules and/or enzymatic domains of other PKS and/or NRPS systems) to produce a wide variety of compounds including, but not limited to various enediyne or enediyne derivatives, various polyketides, polypeptides, polyketide/polypeptide hybrids, various thiazoles, various sugars, various methylated polypeptides/polyketides, and the like.

As with the directed production of various metabolites described above, such compounds can be produced, *in vivo* or *in vitro*, by catalytic biosynthesis, *e.g.*, using large, enediyne cluster units and/or modular PKSs, NRPSs, and hybrid PKS/NRPS systems. In a preferred embodiment large combinatorial libraries of cells harboring various megasynthetases can be produced by the random or directed modification of particular pathways and then selected for the production of a molecule or molecules of interest. It will be appreciated that, in certain embodiments, such libraries of megasynthetases/modified pathways, can be used to generate large, complex combinatorial libraries of compounds which themselves can be screened for a desired activity.

Such combinatorial libraries can be created by the deliberate modification/variation of selected biosynthetic pathways and/or by random/haphazard modification of such pathways.

A) Directed engineering of novel synthetic pathways.

In numerous embodiments of this invention, novel polyketides, polypeptides, and combinations thereof are created by modifying the entediyne gene cluster ORFs and/or known PKSs, and/or NRPSs so as to introduce variations into metabolites synthesized by the enzymes. Such variations may be introduced by design, for example to modify a known molecule in a specific way, *e.g.* by replacing a single monomeric unit within a polymer with another, thereby creating a derivative molecule of predicted structure. Such variations can also be made by adding one or more modules or enzymatic domains to a known PKS or NRPS or enedigne cluster, or by removing one or more module from a known PKS or NRPS.

Using any of these methods, it is possible to introduce PKS domains, NRPS domains, and entediyne domains into a megasynthetase. Mutations can be made to the

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native enediyne, and/or NRPS, and/or PKS subunit sequences and such mutants used in place of the native sequence, so long as the mutants are able to function with other subunits (domains) in the synthetic pathway. Such mutations can be made to the native sequences using conventional techniques such as by preparing synthetic oligonucleotides including the mutations and inserting the mutated sequence into the gene encoding a NRPS and/or PKS subunit using restriction endonuclease digestion. (see, e.g., Kunkel, (1985) Proc. Natl. Acad. Sci. USA 82: 448; Geisselsoder et al. (1987) BioTechniques 5: 786). Alternatively, the mutations can be effected using a mismatched primer (generally 10-20 nucleotides in length) which hybridizes to the native nucleotide sequence (generally cDNA corresponding to the RNA sequence), at a temperature below the melting temperature of the mismatched duplex. The primer can be made specific by keeping primer length and base composition within relatively narrow limits and by keeping the mutant base centrally located (Zoller and Smith (1983) Meth, Enzymol. 100: 468). Primer extension is effected using DNA polymerase, the product cloned and clones containing the mutated DNA, derived by segregation of the primer extended strand, selected. Selection can be accomplished using the mutant primer as a hybridization probe. The technique is also applicable for generating multiple point mutations (see, e.g., Dalbie-McFarland et al. (1982) Proc. Natl. Acad. Sci USA 79:6409). PCR mutagenesis will also find use for effecting the desired mutations.

B) Random modification of enediyne pathways.

In another embodiment, variations can be made randomly, for example by making a library of molecular variants (e.g. of a known enediance) by randomly mutating one or more elements of the subject gene cluster or by randomly replacing one or more open reading frames in a gene cluster with one or more of alternative open reading frames.

The various open reading frames can be combined into a single multi-modular enzyme, thereby dramatically increasing the number of possible combinations obtained using these methods. These combinations can be made using standard recombinant or nucleic acid amplification methods, for example by shuffling nucleic acid sequences encoding various modules or enzymatic domains to create novel arrangements of the sequences, analogous to DNA shuffling methods described in Crameri *et al.* (1998) *Nature* 391: 288-291, and in U.S. Patents 5,605,793 and in 5,837,458. In addition, novel combinations can be made in vitro, for example by combinatorial synthetic methods. Novel molecules or molecule libraries, can be screened for any specific activity using standard methods.

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Random mutagenesis of the nucleotide sequences obtained as described above can be accomplished by several different techniques known in the art, such as by altering sequences within restriction endonuclease sites, inserting an oligonucleotide linker randomly into a plasmid, by irradiation with X-rays or ultraviolet light, by incorporating incorrect nucleotides during in vitro DNA synthesis, by error-prone PCR mutagenesis, by preparing synthetic mutants or by damaging plasmid DNA in vitro with chemicals. Chemical mutagens include, for example, sodium bisulfite, nitrous acid, hydroxylamine, agents which damage or remove bases thereby preventing normal base-pairing such as hydrazine or formic acid, analogues of nucleotide precursors such as nitrosoguanidine, 5-bromouracil, 2-aminopurine, or acridine intercalating agents such as proflavine, acriflavine, quinacrine, and the like. Generally, plasmid DNA or DNA fragments are treated with chemicals, transformed into *E. coli* and propagated as a pool or library of mutant plasmids.

Large populations of random enzyme variants can be constructed in vivo using "recombination-enhanced mutagenesis." This method employs two or more pools of, for example, 10⁶ mutants each of the wild-type encoding nucleotide sequence that are generated using any convenient mutagenesis technique, described more fully above, and then inserted into cloning vectors.

C) Incorporation and/or modification of non-C-1027 cluster elements.

In either the directed or random approaches, nucleic acids encoding novel combinations of gene cluster ORFs are introduced into a cell. In one embodiment, nucleic acids encoding one or more enediyne synthetic cluster ORFS and/or PKS and/or NRPS domains are introduced into a cell so as to replace one or more domains of an endogenous gene cluster within a cell. Endogenous gene replacement can be accomplished using standard methods, such as homologous recombination. Nucleic acids encoding an entire enediyne, enediyne ORF, PKS, NRPS, or combination thereof can also be introduced into a cell so as to enable the cell to produce the novel enzyme, and, consequently, synthesize the novel polymer. In a preferred embodiment, such nucleic acids are introduced into the cell optionally along with a number of additional genes, together called a 'gene cluster,' that influence the expression of the genes, survival of the expressing cells, etc. In a particularly preferred embodiment, such cells do not have any other enediyne and/or PKS- and/or NRPS-encoding genes or gene clusters, thereby allowing the straightforward isolation of the molecule(s) synthesized by the genes introduced into the cell.

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Furthermore, the recombinant vector(s) can include genes from a single enediyne and/or PKS and/or NRPS gene cluster, or may comprise hybrid replacement PKS gene clusters with, *e.g.*, a gene for one cluster replaced by the corresponding gene from another gene cluster. For example, it has been found that ACPs are readily interchangeable among different synthases without an effect on product structure. Furthermore, a given KR can recognize and reduce polyketide chains of different chain lengths. Accordingly, these genes are freely interchangeable in the constructs described herein. Thus, the replacement clusters of the present invention can be derived from any combination of PKS and/or NRPS gene sets that ultimately function to produce an identifiable polyketide.

Examples of hybrid replacement clusters include, but are not limited to, clusters with genes derived from two or more of the act gene cluster, the *whiE* gene cluster, frenolicin (*fren*), granaticin (*gra*), tetracenomycin (*tcm*), 6-methylsalicylic acid (6-msas), oxytetracycline (*otc*), tetracycline (*tet*), erythromycin (*ery*), griseusin (*gris*), nanaomycin, medermycin, daunorubicin, tylosin, carbomycin, spiramycin, avermectin, monensin, nonactin, curamycin, rifamycin and candicidin synthase gene clusters, among others. (For a discussion of various PKSs, *see*, *e.g.*, Hopwood and Sherman (1990) *Ann. Rev. Genet.* 24: 37-66; O'Hagan (1991) The Polyketide Metabolites, Ellis Horwood Limited.

A number of hybrid gene clusters have been constructed, having components derived from the *act*, *fren*, *tcm*, *gris* and *gra* gene clusters (see, e.g., U.S. Patent 5,712,146). Other hybrid gene clusters, as described above, can easily be produced and screened using the disclosure herein, for the production of identifiable polyketides, polypeptides or polyketide/polypeptide hybrids.

Host cells (e.g. Streptomyces) can be transformed with one or more vectors, collectively encoding a functional PKS/NRPS set, or a cocktail comprising a random assortment of enediyne ORFs and/or PKS and/or NRPS genes, modules, active sites, or portions thereof. The vector(s) can include native or hybrid combinations of enediyne ORFs, and/or PKS and/or NRPS subunits or cocktail components, or mutants thereof. As explained above, the gene cluster need not correspond to the complete native gene cluster but need only encode the necessary enediyne ORFs and/or PKS and/or NRPS components to catalyze the production of the desired product(s).

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IV. Variation of starter and/or extender units, and/or host cells.

In addition to varying the nucleic acids comprising the subject gene cluster, variations in the products produced by the gene cluster(s) can be obtained by varying the the host cell, the starter units and/or the extender units. Thus, for example different fatty acids can be utilized in the enediyne synthetic pathway resulting in different enediyne variants. Similarly different intermediate metabolites can be provided (e.g. endogenously produced by the host cell, or produced by an introduced herterologous construct, and/or supplied from an exogenous source (e.g. the culture media)). Similarly, varying the host cell can vary the resulting product(s). For example, a gene cassette carrying the enediyne biosynthesis genes can be introduced into a deoxysugar-synthesizing host for the production of glycosylated enediyne metabolites.

V. Use of C-1027 resistance genes.

The antibiotic C-1027 and metabolites present in C-1027 biosynthesis are highly potent cytotoxins. Accordingly the biosynthesis of C-1027 is facilitated by the presence of one or more antibiotic (e.g. enediyne) resistance genes. Without being bound to a particular theory, it is believed that CagA and SgcB function cooperatively to provide resistance. It is believed that the C-1027 chromophore is first sequestered by binding to the preaproprotein CagA (ORF 9) to form a complex, which is then transported out of the cell by the efflux pump SgcB (ORF 2) and processed by removing the leader peptide to yield the chromoprotein. Other genes that appear to mediate resistance in the C-1027 biosynthesis gene cluster include a transmembrane transport protein (ORF 27), a Na⁺/H⁺ transporter (ORF 0), an ABC transporter (ORF -1, C-terminus), a glycerol phosphate transporter (ORF -2), and a UvrA-like protein (ORF -1, N-terminus) (see, e.g., Table 2).

These ORFs and/or the polypeptides encoded by these ORFs can be utilized alone, or in combination with one or more other C-1027 ORFs to confer resistance to enedigne or enedigne metabolites on a cell. This is useful in a wide variety of contexts. For example, to increase production of enedignes. For example, it is believed that C-1027 resistance could be a limiting factor at the onset of C-1027 production. Provision of an extra copy of the plasmid-born sgcB, and overexpression of sgcB under the control of the constitutive $ermE^*$ promoter resulted in increase of C-1027 production (see example 1).

In a therapeutic context, it is sometimes desirable to confer resistance on certain vulnerable cells. Thus, for example, where an enedigne is used as a

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chemotherapeutic, transfection of vulnerable, but healthy cells (*e.g.* liver cells remote from the tumor site, stem cells, *etc.*) with vector(s) expressing the resistance gene(s) permits administration of the enediyne at a higher dosage with fewer adverse effects to the organism. Such approaches have been taken using the multi-drug resistance gene (MDR1) expressing p-glycoprotein.

In another embodiment vectors are provided containing one or more resistance genes of this invention under control of a constitutive and/or inducible promoter thereby providing a "ready-made" expression system suitable for the expression of an enediyne or enediyne metabolite at high concentration.

It is also noted that the resistance genes are expected to confer resistance to compounds other than enedignes. The resistance genes are expected to confer resistance to essentially any cytotoxic compound that can act as a substrate for the resistance gene(s) of this invention.

VI. Kits.

In still another embodiment, this invention provides kits for practice of the methods described herein. In one preferred embodiment, the kits comprise one or more containers containing nucleic acids encoding one or more of the C-1027 biosynthesis gene cluster open reading frames. Certain kits may comprise vectors encoding the sgc gene cluster orfs and/or cells containing such vectors. The kits may optionally include any reagents and/or apparatus to facilitate practice of the methods described herein. Such reagents include, but are not limited to buffers, labels, labeled antibodies, bioreactors, cells, etc.

In addition, the kits may include instructional materials containing directions (*i.e.*, protocols) for the practice of the methods of this invention. Preferred instructional materials provide protocols utilizing the kit contents for creating or modifying C-1027 gene cluster and/or for synthesizing or modifying a molecule using one or more sgc gene cluster ORFs. While the instructional materials typically comprise written or printed materials they are not limited to such. Any medium capable of storing such instructions and communicating them to an end user is contemplated by this invention. Such media include, but are not limited to electronic storage media (e.g., magnetic discs, tapes, cartridges, chips), optical media (e.g., CD ROM), and the like. Such media may include addresses to internet sites that provide such instructional materials.

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EXAMPLES

The following examples are offered to illustrate, but not to limit the claimed invention.

Example 1

Genes for production of the enedigne antitumor antibiotic C-1027 in Streptomyces globisporus are clustered with the cagA gene that encodes the C-1027 apoprotein

We have been studying the biosynthesis of C-1027 in Streptomyces globisporus C-1027 as a model for the enediyne family of antitumor antibiotics (Thorson et al. (1999) Bioorg. Chem., 27: 172-188). C-1027 consists of a non-peptidic chromophore and an apoprotein, CagA [also called C-1027AG (Otani et al. (1991) Agri. Biol. Chem. 55: 407-417)]. The C-1027 chromophore is extremely unstable in the protein-free state, the structure of which was initially deduced from an inactive but more stable degradation product (Minami et al. (1993) Tetrahedron Lett. 34: 2633-2636) and subsequently confirmed by spectroscopic analysis of the natural product (Yoshida et al. (1993) Tetrahedron Lett. 34: 2637-2640) (Fig. 1). While the absolute stereochemistry of the deoxysugar moiety was established by total synthesis (Iida et al. (1993) Tetrahedron Lett. 34: 4079-4082), the 8S, 9S, 13S and 17R configuration of the C-1027 chromophore were based only on computer modeling (Okuno et al. (1994) J. Med. Chem. 37: 2266-2273). Although no biosynthetic study has been carried out specifically on C-1027, the polyketide origin of the enediyne cores has been implicated by feeding experiments with ¹³C-labeled acetate for the neocarzinostatin chromophore A (Hensens et al. (1989) J. Am. Chem. Soc. 111: 3295-3299), dynemicin (Tokiwa et al. (1992) J. Am. Chem. Soc. 114: 4107-4110), and esperamicin (Lam et al. (1993) J. Am. Chem. Soc. 115: 12340-12345); and deoxysugar biosynthesis has been well characterized in actinomycetes (Liu and Thorson (1994) Ann. Rev. Microbiol. 48: 223-256; Piepersberg (1997) pp. 81-163. In Biotechnology of antibiotics, 2nd ed. W. R. Strohl (ed). Marcel Dekker, New York). Given the structural similarity of C-1027 to the other enediyne cores and to deoxysugars found in other secondary metabolites, we decided to clone either a PKS or a deoxysugar biosynthesis gene as the first step of identifying the C-1027 gene cluster from S. globisporus.

Furthermore, the CagA apoprotein of C-1027 has been isolated, its amino acid sequence has been determined, and the corresponding *cagA* gene has been cloned and sequenced (Otani *et al.* (1991) *Agri. Biol. Chem.* 55: 407-417; Sakata *et al.* (1992) *Biosci.*

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Biotech. Biochem. 56: 1592-1595). Since genes encoding secondary metabolite production in actinomycetes have invariably been found to be clustered in one region of the microbial chromosome (Hopwood (1997) Chem. Rev. 97: 2465-2497), we further reasoned that mapping the cagA gene with either a putative PKS gene, a deoxysugar biosynthesis gene, or both to the same region of the S. globisporus chromosome should be viewed as strong evidence supporting the proposition that the cloned genes constitute the C-1027 biosynthesis gene cluster.

We report here the cloning and sequencing of two genes, sgcA (Streptomyces globisporus C-1027) and sgcB, that encode a dNDP-glucose 4,6-dehydratase (NGDH) and a transmembrane efflux protein, respectively. The sgcA, B locus is indeed clustered with the cagA gene, leading to the localization of a 75-kb gene cluster from S. globisporus. The involvement of the cloned gene cluster in C-1027 biosynthesis was demonstrated by disrupting the sgcA gene to generate C-1027-nonproducing mutants and by complementing the sgcA mutants in vivo to restore C-1027 production. Our results, together with similar effort in the Thorson laboratory on the calicheamicin gene cluster (Thorson et al. (1999) Bioorg. Chem., 27: 172-188), represent the first cloning of a gene cluster for enediyne antitumor antibiotic biosynthesis.

Materials and methods.

Bacterial strains and plasmids.

Escherichia coli DH5α was used as a general host for routine subcloning (Sambrook et al. (1989) Molecular cloning, a laboratory manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). E. coli XL 1-Blue MR (Stratagene, La Jolla, CA) was used as the transduction host for cosmid library construction. E. coli S17-1 was used as the donor host for E. coli-S. globisporus conjugation (Mazodier et al. (1989) J. Bacteriol. 171: 3583-3585). Micrococcus luteus ATCC9431 was used as the testing organism to assay the antibacterial activity of C-1027 (Hu et al. (1988) J. Antibiot. 41: 1575-1579). The pGEM-3zf, -5zf, and -7zf and pGEM-T vectors were from Promega (Madison, WI). S. globisporus strains and other plasmids in this study are listed in Table 3

Table 3. Strains and plasmids.

Strain or	Relevant Characteristics

plasmid	
S. globisp	orus
C-1027	Wild-type (Hu et al. (1988) J. Antibiot. 41: 1575-1579)
AF40	Mutant resulted from acriflavine treatment of S. globisporus C-1027, C-
	1027-nonproducing (Mao et al. (1997) Chinese J. Biotechnol. 13: 195-
	199)
AF44	Mutant resulted from acriflavine treatment of S. globisporus C-1027, C-
	1027-nonproducing (Mao et al., supra)
AF67	Mutant resulted from acriflavine treatment of S. globisporus C-1027, C-
Aro	
CD 1001	1027-nonproducing (Mao et al., supra)
SB1001	sgcA-disrupted mutant resulted from integration of pBS1012 into S.
	globisporus C-1027 Apr ^R , C-1027-nonproducing
SB1002	sgcA-disrupted mutant resulted from integration of pBS1013 into S.
	globisporus C-1027 Apr ^R , C-1027-nonproducing
Plasmids:	
pOJ446	E. coli-Streptomyces shuttle cosmid, Apr ^R (Bierman et al. (1992) Gene, 116: 43-
pOJ260	E. coli vector, non-replicating in Streptomyces, Apr ^R (Bierman et al. supra)
pKC1139	E. coli-Streptomyces shuttle vector, rep ^{TS} , Apr ^R (Bierman et al. supra)
pWHM3	E. coli-Streptomyces shuttle vector, Th ^R (Vara et al. (1989) J. Bacteriol. 171:
	5872-5881)
pWHM79	ermE* promoter in pGEM-3zf (Shen and Hutchinson (1996) Proc. Natl. Acad.
	Sci. USA 93: 6600-6604)
pBS1001	0.75-kb PCR product amplified from S. globisporus with type I PKS primers in pGEM-
	Т
pBS1002	0.55-kb PCR product amplified from S. globisporus with NGDH gene primers in
701000	pGEM-T
pBS1003 pBS1004	0.73-kb PCR product amplified from pBS1005 with <i>cagA</i> primers in pGEM-T
pBS1004 pBS1005	pOJ446 <i>S. globisporus</i> genomic library cosmid pOJ446 <i>S. globisporus</i> genomic library cosmid
pBS1005	pOJ446 S. globisporus genomic library cosmid
pBS1007	3.0-kb BamHI fragment from pBS1005 in pGEM-3zf, sgcA, sgcB
pBS1008	4.0-kb BamHI fragment from pBS1005 in pGEM-3zf, cagA
pBS1009	1.0-kb KpnI truncated fragment of sgcA from pBS1007 in pGEM-3zf
pBS1010	0.75-kb SacII/SphI internal fragment of sgcA from pBS1009 in pGEM-5zf

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pBS1011	0.75-kb SacI/SphI internal fragment of sgcA from pBS1010 in pGEM-3zf
pBS1012	0.75-kb EcoRI/HindIII internal fragment of sgcA from pBS1010 in pOJ260
pBS1013	0.75-kb EcoRI/HindIII internal fragment of sgcA from pBS1010 in pKC1139
pBS1014	2.0-kb EcoRI/SphI fragment from pBS1007 in the SmaI/SphI sites of pWHM79, ermE*,
	sgcA
pBS1015	2.5-kb EcoRI/HindIII fragment from pBS1014 in pWHM3, ermE*, sgcA
pBS1016	Self-ligation of the 5.2-kb KpnI fragment from pBS1007
pBS1017	0.45-kb EcoRI/SacI fragment from pWHM79 in EcoRI/SacI sites of pBS1016, ermE*,
	sgcB
pBS1018	2.5-kb EcoRI/HindIII fragment from pBS1017 in pKC1139, ermE*, sgcB

Biochemicals and chemicals.

Ampicillin, apramycin, nalidixic acid, and thiostrepton were from Sigma (St. Louis, MO). Unless specified otherwise, restriction enzymes and other molecular biology reagents were from standard commercial sources.

Media and culture conditions.

E. coli strains carrying plasmids were grown in Luria-Bertani (LB) medium and were selected with appropriate antibiotics. S. globisporus strains were grown on ISP-4 (Difco Laboratories, Detroit, MI) or R2YE at 28°C for sporulation and in TSB (Hopwood et al. (1985) Genetic manipulation of Streptomyces: a laboratory manual. John Innes Foundation, Norwich, UK) supplemented with 5 mM MgCl₂ and 0.5% glycine at 28°C, 250 rpm for isolation of genomic DNA. For transformation, S. globisporus strains were grown in YEME (Hopwood et al., supra.) for preparation of protoplasts and on R2YE for protoplast regeneration. For conjugation, both the E. coli S17-1 donors and the S. globisporus recipients (upon germination in TSB) were prepared in LB, and donors/recipients were grown on either ISP-4 medium with 0.05% yeast extract and 0.1% tryptone or AS-1 medium (Baltz (1980) Dev. Ind. Microbiol. 21: 43-54; Bierman et al. (1992) Gene 116: 43-69) at 30°C for isolation of exconjugants.

For C-1027 production, *S. globisporus* strains were grown either on R2YE or ISP-4 agar medium at 28°C or in liquid medium by a two-stage fermentation. For liquid culture, the seed inoculum was prepared by inoculating 50 mL medium (consisting of 2% glycerol, 2% dextrin, 1% fish meal, 0.5% peptone, 0.2% (NH₄)₂SO₄, and 0.2% CaCO₃, pH 7.0) with an aliquot of spore suspension, incubating at 28°C, 250 rpm for two days. To a

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fresh 50 mL of the same medium was then added the seed culture (5%), and incubation continued at 28°C, 250 rpm for three to six days (Hu *et al.* (1988) *J. Antibiot.* 41: 1575-1579). The fermentation supernatants were harvested by centrifugation (Eppendorf 5415C, 4°C, 10 min, 14,000 rpm) on day 3, 4 and 5, and assayed for their antibacterial activity against *M. luteus* (Hu *et al.* (1988) *J. Antibiot.*, 41: 1575-1579).

DNA isolation and manipulation.

Plasmid preparation and DNA extraction were carried out by using commercial kits (Qiagen, Santa Clarita, CA). Total *S. globisporus* DNA was isolated according to literature protocols (Hopwood *et al.* (1985) *Genetic manipulation of Streptomyces: a laboratory manual.* John Innes Foundation, Norwich, UK; Rao *et al.* (1987) *Methods Enzymol.* 153: 166-198). Restriction endonuclease digestion and ligation followed standard methods (Sambrook *et al.* (1989) *Molecular cloning, a laboratory manual.* Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). For Southern analysis, digoxigenin labeling of DNA probes, hybridization, and detection were performed according to the protocols provided by the manufacturer (Boehringer Mannheim Biochemicals, Indianapolis, IN).

DNA sequencing.

Automated DNA sequencing was carried out on an ABI Prism 377 DNA Sequencer using the ABI Prism dye terminator cycle sequencing ready reaction kit and Ampli*Taq* DNA polymerase FS (Perkin-Elmer/ABI, Foster City, CA). Sequencing service was provided by either the DBS Automated DNA Sequencing Facility, UC Davis, or Davis Sequencing Inc. (Davis, CA). Data were analyzed by ABI Prism Sequencing 2.1.1 software and the Genetics Computer Group program (Madison, WI).

Polymerase chain reaction (PCR).

Primers were synthesized at the Protein Structure Laboratory, UC Davis.

PCR was carried out on a Gene Amp PCR System 2400 (Perkin-Elmer/ABI) with *Taq*polymerase and buffer from Promega. A typical PCR mixture consisted of 5 ng of *S.*globisporus genomic or plasmid DNA as template, 25 pmoles of each primers, 25 μM dNTP,
5% DMSO, 2 units of *Taq* polymerase, 1 x buffer, with or without 20% glycerol in a final volume of 50 μL. The PCR temperature program was as follows: initial denaturing at 94°C

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for 5 min, 24-36 cycles of 45 sec at 94°C, 1 min at 60°C, 2 min at 72°C, followed by additional 7 min at 72°C.

For type II PKS, the following two pairs of degenerate primers were used—5'-AGC TCC ATC AAG TCS ATG RTC GG-3' (forward, SEQ ID NO:103) / 5'-CC GGT GTT SAC SGC GTA GAA CCA GGC G-3' (reverse, SEQ ID NO:104) and 5'-GAC ACV GCN TGY TCB TCV-3' (forward, SEQ ID NO: 105)/5'-RTG SGC RTT VGT NCC RCT-3' (SEQ ID NO:106) (B, C+G+T; N, A+C+G+T; R, A+G; S, C+G; V, A+C+G; Y, C+T) (reverse) (Seow *et al.* (1997) *J. Bacteriol.,* 179: 7360-7368). No product was amplified under all conditions tested. For type I PKS, the following pair of degenerate primers were used—5'-GCS TCC CGS GAC CTG GGC TTC GAC TC-3' (forward, SEQ ID NO:107) / 5'-AG SGA SGA GCA GGC GGT STC SAC-3' (S, G+C) (reverse, SEQ ID NO:108) (Kakavas *et al.* (1997) *J. Bacteriol.,* 179: 7515-7522). A distinctive product with the predicted size of 0.75 kb was amplified in the presence of 20% glycerol and cloned into pGEM-T according to the protocol provided by the manufacturer (Promega) to yield pBS1001.

For NGDH, the following pair of degenerate primers were used—5'-CS GGS GSS GCS GGS TTC ATC GG-3' (forward, SEQ ID NO:109) / 5'-GG GWR CTG GYR SGG SCC GTA GTT G-3' (R, A+G; S, C+G; W, A+T; Y, C+T) (reverse, SEQ ID NO:110) (Decker, *et al.* (1996) FEMS Lett., 141: 195-201). A distinctive product with the predicted size of 0.55 kb was amplified and cloned into pGEM-T to yield pBS1002.

For *cagA*, the following pair of primers, flanking its coding region, were used—5'-AG GTG GAG GCG CTC ACC GAG-3' (forward, SEQ ID NO:111)/5'-G GGC GTC AGG CCG TAA GAA G-3' (reverse, SEQ ID NO:112) (Sakata *et al.* (1992) *Biosci. Biotechnol. Biochem.*, 56: 159201595). A distinctive product with the predicted size of 0.73 kb was amplified from pBS1005 and cloned into pGEM-T to yield pBS1003.

Genomic library construction and screening.

S. globisporus genomic DNA was partially digested with MboI to yield a smear around 60 kb, as monitored by electrophoresis on a 0.3% agarose gel. This sample was dephosphorylated upon treatment with shrimp alkaline phosphatase and ligated into the E. coli-Streptomyces shuttle vector pOJ446 (Bierman et al. (1992) Gene 116: 43-69) that was prepared by digestion with HpaI, shrimp alkaline phosphatase treatment, and additional digestion with BamHI. The resulting ligation mixture was packaged with the Gigapack II

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XL two-component packaging extract (Stratagene). The package mixture was transduced into *E. coli* XL 1-Blue MR. The transduced cells were spread onto LB plates containing apramycin (100 μg/mL) and incubated at 37°C overnight. The titer of the primary library was approximately 6,000 colony-forming units per μg of DNA. Restriction enzyme analysis of twelve randomly selected cosmids confirmed that the average size of inserts was about 35 to 45 kb (Rao *et al.* (1987) *Meth. Enzymol.*, 153: 166-198).

To screen the genomic library, colonies from five LB plates containing apramycin (100 μg/mL, with approximately 2,000 colonies per plate) were transferred to nylon transfer membranes (Micro Separations, Inc., Westborough, MA) and screened by colony hybridization with the PCR-amplified 0.55-kb NGDH fragment from pBS1002 as a probe. The positive cosmid clones were re-screened by PCR with primers for NGDH and confirmed by Southern hybridization (Sambrook *et al., supra.*). Further restriction enzyme mapping and chromosomal walking of these overlapping cosmids led to the genetic localization of the 75-kb *sgc* gene cluster, as represented by pBS1004, pBS1005, and pBS1006 (Fig. 5A). A 3.0-kb *Bam*HI fragment from pBS1005 that hybridized to the NGDH probe was cloned into the same sites of pGEM-3zf to yield pBS1007. Similarly, a 4.0-kb *Bam*HI fragment from pBS1005 that hybridizes to the PCR-amplified 0.73-kb *cagA* probe from pBS1003 was cloned into the same sites of pGEM-3zf to yield pBS1008 (Fig. 5B).

Generation of sgcA mutants by insert-directed homologous recombination in S. globisporus.

A 1.0-kb *Kpn*I fragment from pBS1007, containing the C-terminal truncated *sgcA*, was subcloned into pGEM-3zf to yield pBS1009. An internal fragment of *sgcA* was moved sequentially as a 0.75-kb *SacII/SphI* fragment from pBS1009 into the same sites of pGEM-5zf to yield pBS1010 and as a 0.75-kb *SacI/SphI* fragment from pBS1010 into the same sites of pGEM-3zf to yield pBS1011. The latter plasmid was digested with *Eco*RI and *HindIII*, and the resulting 0.75-kb *Eco*RI/*HindIII* fragment was cloned into the same sites of pOJ260 and pKC1139 (Bierman *et al.* (1992) Gene, 116: 43-69 to yield pBS1012 and pBS1013, respectively.

Introduction of pBS1012 and pBS1013 into *S. globisporus* was carried out by either polyethyleneglycol (PEG)-mediated protoplast transformation (Hopwood *et al.* (1985) *Genetic manipulation of Streptomyces: a laboratory manual.* John Innes Foundation, Norwich, UK) or *E. coli-S. globisporus* conjugation (Bierman *et al.* (1992) *Gene* 116: 43-69;

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Matsushima and Baltz (1996) Microbiology 142: 261-267; Matsushima et al. (1994) Gene 146: 39-45), methods for both of which were developed recently in our laboratory. In brief, for transformation, pBS1012 and pBS1013 were propagated in E. coli ET12567 (MacNeil et al. (1992) Gene 111: 61-68), and the resulting double strand plasmid DNA was denatured by alkaline treatment (Ho and Chater (1997) J. Bacteriol. 179: 122-127). The latter DNA (5 μL) and 200 μL of 25% PEG 1000 in P buffer (Hopwood et al. supra) were sequentially added to 50 µL of S. globisporus protoplasts (10⁹) in P buffer. The resulting suspension was mixed immediately and spread on R2YE plates. After incubation at 28°C for 16 to 20 hrs. the plates were overlaid with soft R2YE (0.7% agar) containing apramycin (100 µg/mL, final concentration); incubation continued until colonies appeared (in 5 to 7 days). For conjugation, E. coli S17-1(pBS1012) or E. coli S17-1 (pBS1013) was grown to an OD600 of 0.3 to 0.4. Cells from a 20-mL culture were pelleted by centrifugation, washed in LB, and resuspended in 2 mL of LB as the E. coli donors. S. globisporus spores (10³ to 10⁹) were washed, resuspended in TSB, and incubated at 50°C for 10 min to activate germination. After additional incubation at 37°C for 2 to 5 hrs, the spores were pelleted and resuspended in LB as the S. globisporus recipients. The donors (100 µL) and recipients (100 µL) were mixed and spread equally onto two modified ISP-4 or AS-1 plates supplemented freshly with 10 mM MgCl₂ (see Media and culture conditions). The plates were incubated at 28°C for 16 to 22 hrs. After removal of most of the E. coli S17-1 donors by washing the surface with sterile water, the plates were overlaid with 3 mL of soft LB (0.7% agar) containing nalidixic acid (50 µg/mL, final concentration) and apramycin (100 µg/mL, final concentration) and incubated at 28°C until exconjugants appeared (in approximately 5 days).

Unlike pBS1012, which is a *Streptomyces* non-replicating plasmid, pBS1013 bears a temperature-sensitive *Streptomyces* replication origin (Bierman *et al.* (1992) *Gene* 116: 43-69; Muth *et al.* (1989) *Mol. Gen. Genet.* 219: 341-348) that is unable to replicate at temperatures above 34°C (Table 3), while the *S. globisporus* wild-type strain grows normally up to 37°C. Thus, spores of *S. globisporus* (pBS1013), from either the transformants or the exconjugants, were spread onto R2YE plates containing apramycin (100 μg/mL). The plates were incubated directly at 37°C, and mutants, resulting from single crossover homologous recombination between pBS1013 and the *S. globisporus* chromosome, were readily obtained in 7 to 10 days. Alternatively, the plates were first incubated at 28°C for 2 days until pinpoint-size colonies became visible and then shifted to 37°C to continue incubation.

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Mutants resulting from single crossover homologous recombination grew out of the original pinpoint-size colonies as easily distinguishable sectors in 7 to 10 days.

Construction of the sgcA and sgcB expression plasmids.

pBS1007 was digested with *Eco*RI, and made blunt-ended by treatment with the Klenow fragment of DNA polymerase I. Upon additional digestion with *Sph*I, the resulting 2.0-kb blunt-ended *Sph*I fragment containing the intact *sgcA* gene was cloned into the *SmaI/Sph*I sites of pWHM79 (Shen *et al.* (1996) *Proc. Natl. Acad. Sci., USA*, 93: 6600-6604) to yield pBS1014. The latter was digested with *Eco*RI and *Hind*III, and the resulting 2.5-kb *Eco*RI/*Hind*III fragment was cloned into the same sites of pWHM3 (Vara *et al.* (1989) *J. Bacteriol.* 171: 5872-5881) to yield pBS1015, in which the expression of *sgcA* is under the control of the *ermE** promoter (Bibb *et al.* (1994) *Mol. Microbiol.* 14: 533-545).

Alternatively, pBS1007 was digested with *Kpn*I, removing most of the *sgcA* gene, and the 5.2-kb *Kpn*I fragment was recovered and self-ligated to yield pBS1016. The *ermE** promoter was subcloned from pWHM79 (Shen *et al.* (1996) *Proc. Natl. Acad. Sci., USA,* 93: 6600-6604) as a 0.45-kb *EcoRI/SacI* fragment and cloned into the same sites of pBS1016 to yield pBS1017. The latter was digested with *EcoRI* and *HindIII*, and the resulting 2.5-kb *EcoRI/HindIII* fragment was cloned into the same sites of pKC1139 to yield pBS1018, in which the expression of *sgcB* is under the control of the *ermE** promoter.

Determination of C-1027 production.

The production of C-1027 was detected by assaying its antibacterial activity against M. luteus (Hu et~al. (1988) J. Antibiot. 41: 1575-1579). From liquid culture, fermentation supernant (180 μ L) was added to stainless steel cylinders placed on LB plates pre-seeded with overnight M. luteus culture (0.01% vol/vol). From solid culture, a small square block (0.5 x 0.5 x 0.5 cm³) of agar from either R2YE or ISP-4 medium was directly placed on M. luteus-seeded LB plates. The plates were incubated at 37°C for 24 hrs, and C-1027 production was estimated by measuring the size of inhibition zones.

Nucleotide sequence accession number.

The nucleotide sequence reported here has been deposited in the GenBank database with the accession number AF201913.

Results.

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No polyketide synthase gene was amplified by PCR from S. globisporus.

On the assumption that the C-1027 enediyne core is of polyketide origin, the PCR approach was adopted to screen *S. globisporus* for any putative PKS genes, although it is far from certain *a priori* if the biosynthesis of the enediyne core invokes a PKS and, if so, whether the enediyne PKS will exhibit a type I or type II structural organization. PCR methods for cloning either type I or type II PKS genes have been developed, and these methods have proven to be very effective in cloning PKS genes from various polyketide-producing actinomycetes (Kakavas *et al.* (1997) *J. Bacteriol.* 179: 7515-7522; Seow *et al.* (1997) *J. Bacteriol.* 179: 7360-7368). While no distinctive product was amplified under all conditions examined with both pairs of primers designed for type II PKS, a single product with the expected size of 0.75 kb was readily amplified by PCR from *S. globisporus* with primers designed for type I PKS, which was subsequently cloned (pBS1001). Intriguingly, sequence analysis of six randomly selected pBS1001 clones yielded an identical product—indicative of a specific PCR amplification—the deduced amino acid sequence of which, however, showed no homology to known PKSs (data not shown), excluding the possibility of using PKS as a probe to identify the *sgc* biosynthesis gene cluster.

Cloning of a putative NGDH gene by PCR from S. globisporus.

The biosynthesis of various deoxyhexoses share a common key intermediate—4-keto-6-deoxyglucose nucleoside diphosphate or its analogs—whose formation from glucose nucleoside diphosphate is catalyzed by the NGDH enzyme, an NAD⁺-dependent oxidoreductase (Liu and Thorson (1994) *Ann. Rev. Microbiol.* 48: 223-256; Piepersberg (1997) pp. 81-163. In *Biotechnology of antibiotics*, 2nd ed. W. R. Strohl (ed). Marcel Dekker, New York). The PCR method was adopted to clone the putative NGDH gene from *S. globisporus* with primers designed according to the homologous regions of various NGDH enzymes from actinomycetes (Decker *et al.* (1996) *FEMS Lett.* 141: 195-201), resulting in the amplification of a single product with the expected size of 0.55 kb (pBS1002). Sequence analysis of pBS1002 confirmed its identity as a part of a putative NGDH gene.

To clone the complete NGDH gene, an *S. globisporus* genomic library, constructed in the *E. coli-Streptomyces* shuttle vector pOJ446 (Bierman *et al.* (1992) *Gene*

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116: 43-69; Rao *et al.* (1987) *Methods Enzymol.* 153: 166-198), was analyzed by Southern hybridization with the PCR-amplified 0.55-kb fragment from pBS1002 as a probe. Of the 10,000 colonies screened, 36 positive colonies were identified, 9 of which were confirmed by PCR to harbor the DGDH gene. Restriction enzyme mapping showed that all of them contained a single 3.0-kb *Bam*HI fragment hybridizing to the NGDH probe. Additional chromosomal walking from this locus eventually led to the localization of the 75-kb *sgc* gene cluster, covered by 18 overlapping cosmids as represented by pBS1004, pBS1005, and pBS1006 (Fig. 5A). The 3.0-kb *Bam*HI fragment was subcloned (pBS1007) (Fig. 5B), and its nucleotide (nt) sequence was determined.

Analysis of the DNA sequences of the sgcA and sgcB genes.

Two complete open reading frames (ORFs) (sgcA and sgcB) were identified within the 3.0-kb BamHI fragment of pBS1007, the 3,035-nt sequence of which is shown in Figure 6. The sgcA gene most likely begins with an ATG at nt 101, preceded by a probable ribosome biding site (RBS), GGAGG, and ends with a TGA stop codon at nt 1099. SgcA should therefore encode a 332-amino acid protein with a molecular weight of 36,341 and an isoelectric point of 6.01. A Gapped-BLAST search showed that the deduced sgcA gene product is highly homologous to various putative and known NGDH enzymes from antibiotic-producing actinomycetes, including Gdh from the erythromycin biosynthesis gene cluster in Saccharopolyspora erythraea (64% identity and 70% similarity) (Linton et al. (1995) Gene 153: 33-40), MtmE from the mithramycin biosynthesis gene cluster in Streptomyces argillaceus (64% identity and 68% similarity) (Lombo et al. (1997) J. Bacteriol. 179: 3354-3357), and TylA2 from the tylosin biosynthesis gene cluster in Streptomyces fradiae (62% identity and 68% similarity) (Merson-Davies and Cundliffe (1994) Mol. Microbiol. 13: 349-355) (Fig. 7). A conserved sequence of 14 amino acid residues close to the N-termini can be easily identified in these proteins, which has been described as a βαβ fold with an NAD⁺-binding motif, GxGxxG, (Fig. 7, boxed), consistent with their biochemical role in deoxyhexose biosynthesis (Liu and Thorson (1994) Ann. Rev. Microbiol. 48: 223-256; Piepersberg (1997) pp. 81-163. In Biotechnology of antibiotics, 2nd ed. W. R. Strohl (ed). Marcel Dekker, New York). The function of Gdh and MtmE as TDPglucose 4,6-dehydratases, requiring NAD⁺ as a cofactor, has been confirmed by an enzyme assay following expression of the gdh (Linton et al. (1995) Gene 153: 33-40) and mtmE gene (Lombo et al. (1997) J. Bacteriol. 179: 3354-3357) in E. coli, respectively, and by

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purification of the Gdh protein from *Sacc. erythraea* (Vara *et al.* (1989) *J. Bacteriol.* 171: 5872-5881). From these data, it is reasonable to suggest that *sgcA* encodes the NGDH enzyme required for the biosynthesis of the 4,6-dideoxy-4-dimethylamino-5-methylrhamnose moiety of the C-1027 chromophore.

Transcribed in the same direction as sgcA, the sgcB gene is located 43 nt downstream of sgcA. It should begin with a GTG at nt 1143, preceded by a probable RBS, AGGAG, and end with a TGA at nt 2708 (Fig. 6). Correspondingly, sgcB should therefore encode a 521-amino acid protein with a molecular weight of 52,952 and an isoelectric point of 4.64. Database comparison of the deduced sgcB product revealed that SgcB is closely related to a family of membrane efflux pumps, such as LfrA from Mycobacterium smegmatis (43% identity and 50% similarity, protein accession number AAC43550) (Takiff et al. (1996) Proc. Natl. Acad. Sci. USA 93: 362-366), OrfA from Streptomyces cinnamomeus (42% identity and 47% similarity, protein accession number AAB71209) (Sommer et al. (1997) Appl. Environ. Microbiol. 63: 3553-3560), and RifP from the rifamycin biosynthesis gene cluster in Amycolatopsis mediterranei (35% identity and 44% similarity, protein accession number AAC01725) Augus et al. (1998) Chem. Biol. 5: 69-79). These proteins are membrane-localized transporters involved in the transport of antibiotics (conferring resistance), sugars, and other substances. While direct evidence is lacking for RifP conferring rifamycin resistance in A. mediterranei by transporting it out of the cells (August et al. (1998) Chem. Biol., 5: 68-79), it has been proven that LfrA employs the transmembrane proton gradient in an antiporter mode to drive the efflux of intracellular antibiotics, resulting in fluoroquinolone resistance in M. smegmatis (Takiff et al. (1996) Proc. Natl. Acad. Sci. USA 93: 362-366). On the basis of the high degree of amino acid sequence conservation, an equivalent role could be proposed for SgcB, conferring resistance by exporting C-1027 from S. globisporus.

The cagA gene is clustered with the sgcA and sgcB locus.

To determine if *cagA* is clustered with the *sgcA* and *sgcB* locus, PCR primers were designed according to the flanking regions of *cagA* (Sakata *et al.* (1992) *Biosci. Biotech. Biochem.* 56: 1592-1595). A single product with the predicted size of 0.73 kb was indeed amplified from several of the overlapping cosmids (which cover the 75-kb *sgc* cluster), including pBS1004 and pBS1005, the identity of which as *cagA* was confirmed by sequencing. Restriction enzyme mapping and Southern hybridization analysis localized

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cagA to a single 4.0-kb BamHI fragment that is approximately 14 kb upstream of the sgcA,B locus (Fig. 5B). The 4.0-kb BamHI fragment was subcloned (pBS1008), and its nt sequence was determined, revealing the cagA gene along with two additional ORFs (data not shown) (Fig. 5). As reported earlier, cagA encodes a 142-amino acid protein that is processed by cleavage of a 32-amino acid lead peptide to yield the mature CagA apoprotein (Sakata et al. (1992) Biosci. Biotech. Biochem. 56: 1592-1595).

Disruption of the sgcA gene in S. globisporus.

To examine if the cloned sgc cluster encodes C-1027 biosynthesis, sgcA was insertionally disrupted by a single crossover homologous recombination event to generate C-1027-nonproducing mutant strains (Fig. 8A). Two plasmids were used—pBS1012 (a pOJ260 derivative) and pBS1013 (a pKC1139 derivative), each of which contain a 0.75-kb internal fragment from sgcA (Table 3). After introduction of pBS1012 into S. globisporus either by PEG-mediated protoplast transformation or E. coli-S. globisporus conjugation, transformants or exconjugants that were resistant to apramycin were isolated in all cases. Since pBS1012 is derived from the Streptomyces non-replicating plasmid of pOJ260, these isolates must have resulted from integration of pBS1012 into the S. globisporus chromosome by homologous recombination. Plasmid pBS1013 was similarly introduced into S. globisporus. However, since pBS1013 is derived from pKC1139 that carries the temperature-sensitive Streptomyces replication origin from pSG5 and can replicate normally at 28°C (Muth et al. (1989) Mol. Gen. Genet. 219: 341-348), these isolates were subjected to incubation at the non-permissive temperature of 37°C to eliminate free plasmids from the host cells. As expected, normal growth stopped except for the recombinants that continue to grow at 37°C, indicative of integration of pBS1013 into S. globisporus by homologous recombination. The apramycin-resistant S. globisporus SB1001 and S. globisporus SB1002 strains were chosen as representatives of mutant strains with disrupted sgcA gene resulted from integration of pBS1012 and pBS1013, respectively.

To confirm that targeted sgcA disruption has occurred by a single crossover homologous recombination event, Southern analysis of the DNA from the mutant strains was performed as exemplified for S. globisporus SB1001 with either pOJ260 or the 0.75-kb SacII/KpnI internal fragment of sgcA from pBS1010 as a probe. As shown in Fig. 8B, a distinctive band of the predicted size of 6.3 kb was detected with the pOJ260 vector as a probe in all mutant strains (lanes 2, 3, and 4); this band was absent from the wild-type strain

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(lane 1). Complementarily, when using the 0.75-kb SacII/KpnI internal fragment of sgcA as a probe (Fig. 8C), the 3.0-kb band in the wild-type strain (lane 1) was split into two fragments with the size of 6.3 kb and 1.0 kb in the mutant strains (lanes 2, 3, and 4), as would be expected for disruption of sgcA by a single crossover homologous recombination event.

S. globisporus SB1001 and S. globisporus SB1002 are C-1027-nonproducing mutants.

No apparent difference in growth characteristics and morphologies between the wild-type *S. globisporus* and mutant *S. globisporus* SB1001 and *S. globisporus* SB1002 strains was observed. While C-1027 production in the wild-type *S. globisporus* strain could be detected on day 3, peaked on day 5, and continued for a few more days, as judged by assaying the antibacterial activity of the culture supernant against *M. luteus* (Hu *et al.* (1988) *J. Antibiot.* 41: 1575-1579), C-1027 production is completely abolished in the *sgcA* mutant strains *S. globisporus* SB1001 and *S. globisporus* SB1002 (Fig. 9A). The latter phenotype was identical to that of the AF40, AF44, and AF67 mutants, C-1027-nonproducing *S. globisporus* strains that have been characterized previously (Fig. 9A and 9C) (Mao, *et al.* (1997) *Chinese J. Biotechnol.* 13: 195-199).

In vivo complementation of S. globisporus SB1001.

The ability of the wild-type sgcA gene to complement the disrupted sgcA gene was tested in the S. globisporus SB1001 strain. The construction of pBS1015, in which the expression of sgcA is under the control of the constitutive $ermE^*$ promoter, was described in Materials and Methods. Both the pBS1015 construct and the pWHM3 vector as a control were introduced by transformation into the S. globisporus SB1001 mutant strains. Culture supernants from each transformant were bioassayed against M. luteus for C-1027 production. pBS1015 restored C-1027 production to S. globisporus SB1001 to the wild-type level; no C-1027 production was detected in the control in which pWHM3 was introduced into S. globisporus BS1001 (Fig. 9B and 9C). A significant reduction of C-1027 production was observed when S. globisporus SB1001(pBS1015) was cultured under identical conditions but without thiostrepton (Fig. 9B vs. 6C), indicative that pBS1015 may be unstable in S. globisporus SB1001 in the absence of antibiotic selection pressure.

Expression of sgcB in S. globisporus.

The effect of *sgcB* on C-1027 production was tested in the wild-type *S. globisporus* strain. The construction of pBS1018, in which the expression of *sgcB* is under the control of the constitutive *ermE** promoter, was described in Materials and Methods. pBS1018 and the pKC1139 vector as a control were each introduced by conjugation into *S. globisporus*. Culture supernatants from each exconjugant were harvested on days 3, 4, and 5, and assayed for C-1027 production by determining the antibacterial activity against *M. luteus*. While no apparent difference for C-1027 production was observed between the *S. globisporus* and *S. globisporus* (pKC1139) strains, a significant increase in C-1027 production (150±25%) was evident in the early stage of *S. globisporus* (pBS1018) fermentation (Fig. 9D, day 3). However, such effect on C-1027 production leveled off as the fermentation proceeded and became insignificant when the culture reached the late stationary phase of fermentation (Fig. 9D, day 4 and 5).

Discussion.

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Our inability to clone the putative enedigne PKS gene by PCR, with degenerate primers designed according to the highly conserved amino acid sequences of either type I or type II PKSs, or by DNA hybridization, with homologous type I or type II PKS as probes (data not shown), was unexpected, since feeding experiments by incorporation of [1-13C]- and [1,2-13C]acetate into the enediyne cores of esperamicin (Lam et al. (1993) J. Am. Chem. Soc. 115: 12340-12345), dynemicin (Tokiwa et al. (1992) J. Am. Chem. Soc. 114: 4107-4110), and neocarzinostatin (Hensens et al. (1989) J. Am. Chem. Soc. 111: 3295-3299) supported their polyketide origin. Although the enediyne cores are structurally distinct from either the reduced or aromatic polyketides, the biosynthesis of which is well characterized by type I or type II PKS, respectively, it could be imagined that an enediyne PKS catalyzes the biosynthesis of a polyunsaturated linear heptaketide intermediate that is subsequently cyclized into the enediyne core structure (Hu et al. (1994) Mol. Microbiol. 14: 163-172; Spaink et al. (1991) Nature 354: 125-130; Thorson et al. (1999) Bioorg. Chem., 27: 172-188). Alternatively, Hensens and co-workers proposed a fatty acid origin for the enediyne core that was also consistent with the isotope labeling results. These authors suggested oleate as a precursor that is shortened by loss of carbons from both ends and is desaturated via the oleate-crepenynate pathway to furnish the enediyne core (Hensens et al. (1989) J. Am. Chem. Soc. 111: 3295-3299). The latter pathway

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resembles polyacetylene biosynthesis in higher plants and fungi and requires an acetylene forming enzyme—a plant gene encoding such an enzyme was identified recently (Lee *et al.* (1998) *Science* 280: 915-918). Our DNA sequence analysis of approximately 60 kb of the *sgc* gene cluster, fails to reveal any gene that resembles PKS.

Although little is known about the resistance mechanism for the enediyne antibiotics in general, the apoproteins of the chromoprotein type of enediynes could be viewed as resistance elements that confer self-resistance to the producing organisms by drug sequestration (Thorson *et al.* (1999) *Bioorg. Chem.*, 27: 172-188). Such a resistance mechanism is in fact well established in antibiotic-producing actinomycetes, for example, BlmA, the bleomycin-binding protein from *Streptomyces verticillus* (Shen *et al.* (1999) *Bioorg. Chem.* 27: 155-171). Given the fact that antibiotic production genes have invariably been found to be clustered in one region of the microbial chromosome, consisting of structural, resistance, and regulatory genes, we adopted a strategy to clone the *sgc* gene cluster by mapping a putative C-1027 structural gene to the previously cloned *cagA* gene, considered as a resistance gene that encodes the C-1027 apoprotein.

We chose NGDH as the putative C-1027 structural gene on the basis of the 4,6-dideoxy-4-dimethylamino-5-methylrhamnose moiety of the C-1027 chromophore. It has been well established that all deoxyhexoses could be derived from the common intermediate of 4-keto-6-deoxyglucose nucleoside diphosphate, the biosynthesis of which from glucose nucleoside diphosphate is catalyzed by an NGDH enzyme. We cloned the NGDH gene from *S. globisporus* by PCR and used it as a probe to screen an *S. globisporus* genomic library, resulting in the isolation of the 75-kb *sgc* gene cluster. DNA sequence analysis of a 3.0-kb *Bam*HI fragment of the *sgc* cluster confirmed the presence of the NGDH protein, encoded by *sgcA*, along with *sgcB* that encodes a transmembrane efflux protein (Fig. 6). The *cagA* gene indeed resides approximately 14 kb upstream of *sgcA* (Fig. 5); DNA sequence analysis of a 4.0-kb *Bam*HI fragment confirmed the identity of *cagA* along with two additional ORFs (data not shown). These results underline once again the effectiveness of cloning natural product biosynthesis gene clusters by exploiting the clustering phenomenon between resistance and structural genes.

The involvement of the cloned gene cluster in C-1027 biosynthesis was demonstrated by disrupting the sgcA gene to generate S. globisporus mutants, the ability of which to produce C-1027 was completely abolished (Fig. 9A), and by complementing the sgcA mutants in vivo upon expression of sgcA in trans to restore C-1027 production (Fig. 9B)

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and 6C). These data unambiguously establish that sgcA is essential for C-1027 production, and thus support the conclusion that the cloned gene cluster encodes C-1027 biosynthesis. It should be pointed out that, although the sgcA mutants S. globisporus SB1001 and S. globisporus SB1002 were characterized as C-1027-nonproducing on the basis of the antibacterial assay alone (Fig. 9A), this phenotype was identical to that of the controls of the AF40, AF44, and AF67 mutants (Fig. 9A and 9C). The latter strains were isolated previously upon randomly mutagenizing the wild-type S. globisporus strain with acriflavine and confirmed to be C-1027-nonproducing by both the antibacterial bioassay and an antitumor spermatogonial assay (Mao, et al. (1997) Chinese J. Biotechnol. 13: 195-199), providing strong support to the current study. Gene disruption and complementation in S. globisporus were made possible by the recently developed genetic system that allowed us to introduce plasmid DNA into S. globisporus via either PEG-mediated protoplast transformation (Hopwood et al. (1985) Genetic manipulation of Streptomyces: a laboratory manual. John Innes Foundation, Norwich, UK) or E. coli-S. globisporus conjugation (Bierman et al. (1992) Gene 116: 43-69; Matsushima and Baltz (1996) Microbiology 142: 261-267; Matsushima et al. (1994) Gene 146: 39-45) for analyzing the sgc biosynthesis gene cluster in vivo. Given the difficulties encountered with calicheamicin biosynthesis in Micromonospora echinospora, into which all attempts to introduce plasmid DNA have failed (Thorson et al. (1999) Bioorg. Chem., 27: 172-188), the latter results underscore the importance of selecting C-1027 as a model system for enedigne biosynthesis so that many of the genetic tools developed in Streptomyces species can now be directly applied to the study of enediyne biosynthesis.

Finally, the function of *sgcB* was probed by examining C-1027 production, following expression of the gene in the wild-type *S. globisporus* strain. Database comparison of the deduced amino acid sequence clearly suggested SgcB as a transmembrane efflux protein, conferring resistance by exporting C-1027 out of the cell. Hence, in addition to CagA, SgcB could be viewed as the second resistance element identified for C-1027 biosynthesis. Multiple resistance genes have been identified in numerous antibiotic biosynthesis gene clusters (Hopwood (1997) *Chem. Rev.* 97: 2465-2497). It could be imagined that CagA and SgcB function cooperatively to provide resistance—the C-1027 chromophore is first sequestered by binding to the preaproprotein CagA to form a complex, which is then transported out of the cell by the efflux pump SgcB and processed by removing the leader peptide to yield the chromoprotein, although we do not have any experimental

data to substantiate this speculation. Since it is known that yields for antibiotic production could be profoundly altered by the introduction of extra copies of regulatory, resistance, or structural genes into wild-type organisms (Hutchinson (1994) *Bio/Technology* 12: 375-380), we tested the effect of overexpressing *sgcB* in *S. globisporus* on C-1027 production. While no apparent adverse effect on C-1027 production was observed upon introduction of the pKC1139 vector into *S. globisporus* (data not shown), a significant increase in C-1027 production (150±25%) was observed in the early stage of *S. globisporus* (pBS1017) fermentation (Fig. 9D, day 3), supporting the predicted function for SgcB in C-1027 biosynthesis. We propose that C-1027 resistance could be a limiting factor at the onset of C-1027 production, which is circumvented by the extra copy of the plasmid-born *sgcB*, and overexpression of *sgcB* under the control of the constitutive *ermE** promoter results in increase of C-1027 production. However, as the *S. globisporus* (pBS1017) fermentation proceeds to its stationary phase, C-1027 resistance is no longer a limiting factor for overall C-1027 production, and the effect of extra copy of SgcB on C-1027 production consequently became insignificant (Fig. 9D, day 5).

In conclusion, genetic analysis of enediyne biosynthesis has heretofore met with little success in spite of considerable effort (Thorson et al. (1999) Bioorg. Chem., 27: 172-188). The localization of the sgc gene cluster and characterization of the sgcA and sgcB genes have now provided an excellent basis for genetic and biochemical investigations and/or modification of C-1027 biosynthesis, and gene disruption and overexpression in S. globisporus clearly demonstrated the potential to construct enediyne-overproducing strains and to produce novel enediynes that may have enhanced potency as novel anticancer drugs using combinatorial biosynthesis and targeted mutagenesis. We envisage that the results from C-1027 biosynthesis should facilitate the cloning and characterization of biosynthesis gene clusters of other enediyne antibiotics in Streptomyces as well as in other actinomycetes, and could have a great impact on the overall field of combinatorial biosynthesis.

It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference in their entirety for all purposes.

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CLAIMS

What is claimed is:

- 1. An isolated nucleic acid comprising a nucleic acid selected from the group consisting of
- a nucleic acid encoding any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA);
 - a nucleic acid encoding a polypeptide encoded by any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA); and
 - a nucleic acid amplified by polymerase chain reaction (PCR) using primer pairs that amplify any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA).
 - 2. The isolated nucleic acid of claim 1, wherein said nucleic comprises a nucleic acid encoding at least two open reading frames (ORFs) selected from the group consisting of ORF-1 through ORF 42, excluding ORF 9 (cagA).
 - 3. The isolated nucleic acid of claim 1, wherein said nucleic comprises a nucleic acid encoding at least three open reading frames (ORFs) selected from the group consisting of ORF-1 through ORF 42, excluding ORF 9 (cagA).
 - 4. An isolated nucleic acid comprising a nucleic acid that specifically hybridizes under stringent conditions to an open reading frame (ORF) of the C-1027 biosynthesis gene cluster, excluding ORF 9 (cagA), and can substitute for the ORF to which it specifically hybridizes to direct the synthesis of an enediyne.
 - 5. The isolated nucleic acid of claim 4, wherein said isolated nucleic acid comprises a nucleic acid that specifically hybridizes under stringent conditions to a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 10, ORF 11, ORF 12, ORF 13, and ORF 14.
 - 6. The isolated nucleic acid of claim 4, wherein said isolated nucleic acid comprises a nucleic acid that specifically hybridizes under stringent conditions to a nucleic

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acid selected from the group consisting of ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42.

- The isolated nucleic acid of claim 5, wherein said isolated nucleic acid comprises a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 10, ORF 11, ORF 12, ORF 13, and ORF 14.
- 8. The isolated nucleic acid of claim 6, wherein said isolated nucleic acid comprises a nucleic acid selected from the group consisting of ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42.
 - 9. The isolated nucleic acid of claim 4, wherein said nucleic acid comprises a nucleic acid that is a single nucleotide polymorphism (SNP) of a nucleic acid selected from the group consisting of ORF -7, ORF -6, ORF -5, ORF -4, ORF -3, ORF -2, ORF -1, ORF 0, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 81, ORF 1, ORF 2, ORF 3, ORF 4, ORF 5, ORF 6, ORF 7, ORF 8, ORF 10, ORF 11, ORF 12, ORF 13, ORF 14, ORF 15, ORF 16, ORF 17, ORF 18, ORF 19, ORF 20, ORF 21, ORF 22, ORF 23, ORF 24, ORF 25, ORF 26, ORF 27, ORF 28, ORF 29, ORF 30, ORF 31, ORF 32, ORF 33, ORF 34, ORF 35, ORF 36, ORF 37, ORF 38, ORF 39, ORF 40, ORF 41, and ORF 42.
 - 10. An isolated gene cluster comprising open reading frames encoding polypeptides sufficient to direct the assembly of a C-1027 enedigne or a C-1027 enedigne analogue.
- The gene cluster of claim 10, wherein said gene cluster is present in a bacterium.
 - 12. The gene cluster of claim 11, wherein said gene cluster is present in a bacterium selected from the group consisting of *Actinomycetes*, *Actinoplanetes*, *Actinomadura*, *Micromonospora*, and *Streptomycetes*.

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- 13. The gene cluster of claim 11, wherein said gene cluster is present in a bacterium selected from the group consisting *Streptomyces globisporus*, *Streptomyces lividans*, *Streptomyces coelicolor*, *Micromonospora echinospora* spp. calichenisis, *Actinomadura verrucosopora*, *Micromonospora chersina*, *Streptomyces carzinostaticus*, and *Actinomycete* L585-6.
- 14. The gene cluster of claim 13, wherein one or more open reading frames is operatively linked to a heterologous promoter.
- 15. An isolated polypeptide comprising a catalytic domain encoded by a nucleic acid of a C-1027 gene cluster wherein said nucleic acid comprises a nucleic acid selected from the group consisting of

a nucleic acid encoding any of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA); and

a nucleic acid amplified by polymerase chain reaction (PCR) using any one of the primer pairs identified in Tables I and II that specifically amplify one or more of (ORFs) -7 through 42, excluding ORF 9 (cagA).

- 16. The polypeptide of claim 15, wherein said polypeptide is encoded by at least two open reading frames selected from the group consisting of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA).
- 17. The polypeptide of claim 15, wherein said polypeptide is encoded by at least three open reading frames selected from the group consisting of C-1027 open reading frames (ORFs) -7 through 42, excluding ORF 9 (cagA).
 - 18. An expression vector comprising a nucleic acid of any one of claims 1 through 9.
 - 19. A host cell transformed with an expression vector of claim 18.
- 25. The host cell of claim 19, wherein said cell is transformed with an exogenous nucleic acid comprising a gene cluster encoding polypeptides sufficient to direct the assembly of a C-1027 enediyne or a C-1027 enediyne analogue.
 - 21. The host cell of claim 19, wherein said host cell is a bacterium.

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- 22. The host cell of claim 21, wherein said bacterium is selected from the group consisting of Actinomycetes, *Actinoplanetes, Actinomadura, Micromonospora*, and *Streptomycetes*.
- 23. The host cell of claim 21, wherein said bacterium is selected from the group consisting of *Streptomyces globisporus*, *Streptomyces lividans*, *Streptomyces coelicolor*, *Micromonospora echinospora* spp. *calichenisis*, *Actinomadura verrucosopora*, *Micromonospora chersina*, *Streptomyces carzinostaticus*, and *Actinomycete* L585-6.
 - 24. A method of chemically modifying a biological molecule, said method comprising contacting a biological molecule that is a substrate for a polypeptide encoded by a C-1027 biosynthesis gene cluster open reading frame, with a polypeptide encoded by a C-1027 biosynthesis gene cluster open reading frame whereby said polypeptide chemically modifies said biological molecule.
 - 25. The method of claim 24, wherein said polypeptide is an enzyme selected from the group consisting of a hydroxylase, a homocysteine synthase, a dNDP-glucose dehydrogenase, a citrate carrier protein, a C-methyl transferase, an N-methyl transferase, an aminotransferase, a CagA apoprotein, an NDP-glucose synthase, an epimerase, an acyl transferase, a coenzyme F390 synthase, and epoxidase hydrolase, an anthranilate synthase, a glycosyl transferase, a monooxygenase, a type II condensation protein, an aminomutase, a type II adenylation protein, an O-methyl transferase, a P-450 hydroxylase, an oxidoreductase, and a proline oxidase.
 - 26. The method of claim 24, wherein said method comprising contacting said biological molecule with at least two different polypeptides encoded by C-1027 biosynthesis gene cluster open reading frames.
- 27. The method of claim 24, wherein said method comprising contacting said biological molecule with at least three different polypeptides encoded by C-1027 biosynthesis gene cluster open reading frames.
 - 28. The method of claim 24, wherein said contacting is in a host cell.
 - 29. The method of claim 28, wherein said host cell is a bacterium.

- 30. The method of claim 24, wherein said contacting ex vivo.
- 31. The method of claim 28, wherein said biological molecule is an endogenous metabolite produced by said host cell.
- 32. The method of claim 28, wherein said biological molecule is an exogenous supplied metabolite.
 - 33. The method of claim 28, wherein said host cell is a eukaryotic cell.
 - 34. The method of claim 33, wherein said eukaryotic cell is selected from the group consisting of a mammalian cell, a yeast cell, a plant cell, a fungal cell, and an insect cell.
 - 35. The method of claim 28, wherein said host cell synthesizes sugars and glycosylates the biological molecule.
 - 36. The method of claim 35, wherein said host cell synthesizes deoxysugars.
- The method of claim 24, wherein said method further comprises contacting said biological molecule with a polyketide synthase or a non-ribosomal polypeptide synthetase.
 - 38. The method of claim of claim 24, wherein said contacting is in a bacterial cell.
 - 39. The method of claim of claim 24, wherein said contacting is ex vivo.
- 20 40. The method of claim 24, wherein said method comprises contacting said biological molecule with at substantially all of the polypeptides encoded by C-1027 biosynthesis gene cluster open reading frames and said method produces an enediyne or enediyne analogue.
- 41. The method of claim 24, wherein said biological molecule is a fatty
 25 acid and said biological molecule is contacted with a C-1027 orf polyeptide selected from the

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group consisting of an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase.

- 42. The method of claim 41, wherein said biological molecule is a fatty acid and said biological molecule is contacted with a plurality of C-1027 orf polypeptides comprising an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase.
- 43. The method of claim 42, wherein said biological molecule is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38.
- 10 44. The method of claim 41, wherein said biological molecule is contacted with polypeptides encoded by ORF 15, ORF 16, ORF 28, ORF 3, ORF 14, and ORF 13.
 - 45. The method of claim 44 wherein said biological molecule is also contacted with polypeptides encoded by ORF 4 and ORF 3.
 - 46. The method of claim 24, wherein said method comprises contacting a sugar with one or more C-1027 open reading frame polypeptides selected from the group consisting of a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase.
 - 47. The method of claim 46, wherein said method comprises contacting a dNDP-glucose with a plurality of C-1027 open reading frame polypeptides comprising a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase.
 - 48. The method of claim 24, wherein said method comprises contacting an amino acid with one or one or more C-1027 open reading frame polypeptides selected from the group consisting of a hydroxylase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein.
 - 49. The method of claim 48, wherein said method comprises contacting an amino acid with a plurality of C-1027 open reading frame polypeptides comprising a

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hydroxylase, a halogenase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein.

- 50. The method of claim 48, wherein said amino acid is a tyrosine.
- 51. A method of synthesizing a chromaprotein type enediyne core, said method comprising contacting a fatty acid with one or more C-1027 orf polypeptides selected from the group consisting of an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase.
 - 52. The method of claim 51, wherein said fatty acid is contacted with a plurality of C-1027 orf polypeptides comprising an epoxide hydrase, a monooxygenase, an iron-sulfer flavoprotein, a p-450 hydroxylase, an oxidoreductase, and a proline oxidase.
 - 53. The method of claim 52, wherein said fatty acid is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38.
 - 54. A method of synthesizing a deoxysugar, said method comprising contacting a sugar with one or more C-1027 open reading frame polypeptides selected from the group consisting of a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase.
- 55. The method of claim 54, wherein said method comprises contacting a dNDP-glucose with a plurality of C-1027 open reading frame polypeptides comprising a dNDP-glucose synthase, a dNDP glucose dehydratase, an epimerase, an aminotransferase, a C-methyltransferase, an N-methyltransferase, and a glycosyl transferase.
 - 56. The method of claim 55, wherein said dNDP-glucose is contacted with polypeptides encoded by ORF17, ORF20, ORF21, ORF29, ORF30, ORF32, ORF35, and ORF38.
 - 57. A method of synthesizing a beta amino acid, said method comprising contacting an amino acid with one or one or more C-1027 open reading frame polypeptides selected from the group consisting of a hydroxylase, an aminomutase, a type II NRPS

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condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein.

- 58. The method of claim 57, wherein said method comprises contacting an amino acid with a plurality of C-1027 open reading frame polypeptides comprising a hydroxylase, a halogenase, an aminomutase, a type II NRPS condensation enzyme, a type II NRPS adenylation enzyme, and a type II peptidyl carrier protein.
 - 59. The method of claim wherein said amino acid is contacted with polypeptides encoded by ORF 4, ORF11, ORF24, ORF23, ORF25, and ORF26.
 - 60. The method of claim 57, wherein said amino acid is a tyrosine.
- 10 61. A method of synthesizing an enediyne or an enediyne analogue said method comprising:

culturing a cell comprising a recombinantly modified C-1027 gene cluster under conditions whereby said cell expresses said enediyne or enediyne analogue; and

recovering said enediyne or enediyne analogue.

- 62. The method of claim 61, wherein said gene cluster is present in a bacterium.
- 63. The gene cluster of claim 62, wherein said gene cluster is present in a bacterium selected from the group consisting of Actinomycetes, *Actinoplanetes*, *Actinoplanetes*, *Actinomadura*, *Micromonospora*, and *Streptomycetes*.
- 64. The gene cluster of claim 62, wherein said gene cluster is present in a bacterium selected from the group consisting *Streptomyces globisporus*, *Streptomyces lividans*, *Streptomyces coelicolor*, *Micromonospora echinospora* spp. calichenisis, *Actinomadura verrucosopora*, *Micromonospora chersina*, *Streptomyces carzinostaticus*, and *Actinomycete* L585-6.
- 65. The method of claim 61, wherein said gene cluster is present in a eukaryotic cell.

- 66. The method of claim 65, wherein said eukaryotic cell is selected from the group consisting of a mammalian cell, a yeast cell, a plant cell, a fungal cell, and an insect cell.
- 67. The method of claim 61, wherein said host cell synthesizes sugars and glycosylates said enediyne or enediyne analogue.
 - 68. The method of claim 67, wherein said host cell synthesizes deoxysugars.
 - 69. A method of making a cell resistant to an enediyne or an enediyne metabolite, said method comprising expressing in said cell one or more isolated C-1027 open reading frame nucleic acids that encode a protein selected from the group consisting of a CagA apoprotein, a SgcB transmembrane efflux protein, a transmembrane transport protein, a Na+/H+ transporter, an ABC transport, a glycerol phosphate transporter, and a UvrA-like protein.
- 70. The method of claim 69, wherein said isolated C-1027 open reading frame nucleic acids are selected from the group consisting of ORF 9, ORF2, ORF 27, ORF 0, ORF 1 c-terminus, ORF 2, and ORF 1 N-terminus.
 - 71. The method of claim 69, wherein said cell is a bacterial cell.

GENE CLUSTER FOR PRODUCTION OF THE ENEDIYNE ANTITUMOR ANTIBIOTIC C-1027

ABSTRACT OF THE DISCLOSURE

This invention provides nucleic acid sequences and characterization of the gene cluster responsible for the biosynthesis of the enediyne C-1027 (produced by *Streptomyces globisporus*). Methods are provided for the biosynthesis of enediynes, enediyne analogs and other biological molecules.

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Fig. 1

ORF10: dNDP-glucose synthase, 355 aa ORF1: dNDP-glucose dehydratase, 332 aa

ORF12: epimerase, 192 aa ORF8: aminotransferase, 410 aa

ORF6: C-methyltransferase, 423 aa ORF7: N-methyltransferase, 244 aa

ORF19: glycosyl transferase, 459 aa

Fig. 2

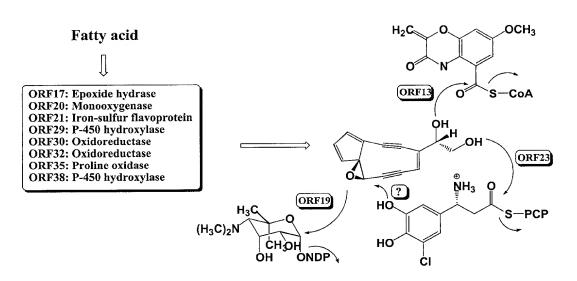
Fig. 3A

ORF4: Hydroxylase, 527 aa
ORF11: Hydroxylase/halogenase, 492/494 aa
ORF24: Aminomutase, 539 aa
ORF26: Type II NRPS adenylation enzyme, 459 aa
ORF26: Type II peptidyl carrier protein, 93 aa

ORF15 ORF4 ORF11
$$H_2N$$
 OCH3 H_2N OCH3

ORF15: Anthranilate synthase I, 493 aa ORF16: Anthranilate synthase II, 220 aa ORF28: O-methyltransferase, 350 aa
ORF3: Coenzyme F390 synthetase, 463 aa ORF14: Coenzyme F390 synthetase, 484 aa ORF13: O-acyltransferase, 378 aa

Fig. 3B



ORF13: O-acyltransferase, ORF19:Glycosyl transferase, ORF23: Type II NRPS condensation enzyme

Fig. 4

	RADHI	
Н	GGGAAGACCG	1.00
101	ATGAGGATGCTGGTGACGGGCGGGGTTTCATCGGCTCGCAGTTCGTGCGGGCCACACTGCACGGCGAGCTGCCGGGTTCCGAGGACGCCCGGGTGA M R M L V T G G A G F I G S Q F V R A T L H G E L P G S E D A R V T SACTI	200
201	GCTGACGTACTCCGGCAATCCGGCCAACCTCACC	300
301	V L D K L T Y S G N P A N L T S V A A H P K Y T F V Q G D T V D P G GCGTCGTCGACGAGGTGGTCGCCGCCACGGCTATCGTCGTCGTCGTCGTCGTCGTCGTCGACGTCGCTCGACGCCGCCACGCCACGGTTCGTCGTCGTCGTCGTCGTCGTCGTCGTCGTCGTTCGTCG	400
401	R V V D E V V A G H D V I V H F A A E S H V D R S I D T A T R F V ACGACCAACGTGCTCGGGACCCAGGGGTCGGGGTCGGGGTCGGGGTCGAAGGGGTCTAAGGGGTCGAA	500
501	T T N V L G T Q T L L E A A L R H G V G R F V H V S T D E V Y G S I TORCCTCGCTCGGCTCCCTACGCGGCGTCGAGGCGGGCTCGATGGACCTGATGGCGCTCGCCTGGCACG	009
601	A S G S W T E D T P L A P N V P Y A A S K A G S D L M A L A W H R CACCEGGGGCTTGGACACCACCAACAACTACGGTCCCTACCACACAACTACCATACCATACCATACCAGAAAGGTGATCCCGCTCTTCGTCACCAACAACTCTC	700
701	T R G L D V V V T R C T N N Y G P Y Q Y P E K V I P L F V T N I L GACGGCTTGCGGGGCCTTGCGGGGCCATCCAGAGGCGCCACCACCGGGCCATCCAGATGGTCATGAGTTCCG	800
. (DGLRVPLYGDGAHRRDWLHVSDHCRAIDMVNNSG	006
30 T	R A G E V Y H I G G G T E L S N E E L T G L L T A C G T D W S C))
901	CGTGGACCGGGTGGCCGACCGCCAGGGGGCACGACCGCCGCTACTCGCTCG	1000
001	GAGGACGGCCTGGCCGCGACGGTGAAGTGGTACCACGAGAACCGTTCGTGGTGGCAGCCGCTGAAGGAAG	1100
	EDGLAATVKWYHENRSWWQPLKEAAGLLDAVG*	
10	GGCAGCCACCGCTAGGAACCCCCAGGAAAGGAGCCACCTCC GTG ACAGCAGTCAAGGAGCCGACGTCCCGCGCAGGAGGGGGGGG	1200
201	TCGTCCTCTCCTTGCCCACGATGCTGTTGATGCTGGACATCAACGTCCTCATGCTGGCCTTGCCGCAGTTGAG	1300
	V L S L P T M L L M L D I N V L M L A L P Q L S E D L G A S S T Q	0071
301	ACAGCTGTGGATCACCGACATCTGCGGGCTTCCTGGTGACCATGGGGCACCTTGGCGGACCGGGCTGGCGGCGCGCGC)
401	GGGGGCGCGCCGTCTTCGCGGTCGTCCTCGTCGTCGCCGC	1500
	G G A A V F A V V S V V A A F S D S A A M L V V S R A V L G V A G A G A A M L V V S R A V L G V A G A	1600
TOC		
601	ဗ	1700
	G V A L G P A V G G L V L A A F W W G S V F L I A V P V M L L V V	

Fig. 6 cont'd.

303	TCTCCCCCTTACGGCACCTGGCCCTCGCTGGATCC	3001
	BamHI	
300	CTGGCGGGAACGACCACCAGAACATCACCTCCATGGCGGACTACGCGGCGCGCGGGTGCCTCTCCTGCGCAAGGCCGAACCTCCTCGCCGGAAGCCGCG	2901
290(2801
280	S L A L A V CCACATGAGCGCACTTCC	2701
270(2601
))
260		2501
250(GCACCGTCGTGTACAGCG	2401
240(GAGCCGCCCCCGGAG	2301
230(CCAGGTGGACACCGGCCTCCTCCTCATCGCGGCCACCACCTGATCTACTTCGGCGCCTCACGGTGGGGCCGATCACGGCGCGCGATCATG C 47	2201
220(2101
210(GGGACCGGACTGATGGTCGCCTGTACCTCCAGACGATCGCCGGTCACTCCCCGTTGGCCGCGGGCTGTGGCTGATCCCGGCCTGCAGCTGCTCGTCG G T G L M V A L Y L Q T I A G H S P L A A G L W L L I P A C M L V V	2001
	RRLADPMLDLGLFADRTLRAGLTVSLVNAVIMG Sphi	
2000		1901
190(TCTACGGACTGAAGGAGCI	1801
1800		1701

Fig. 7

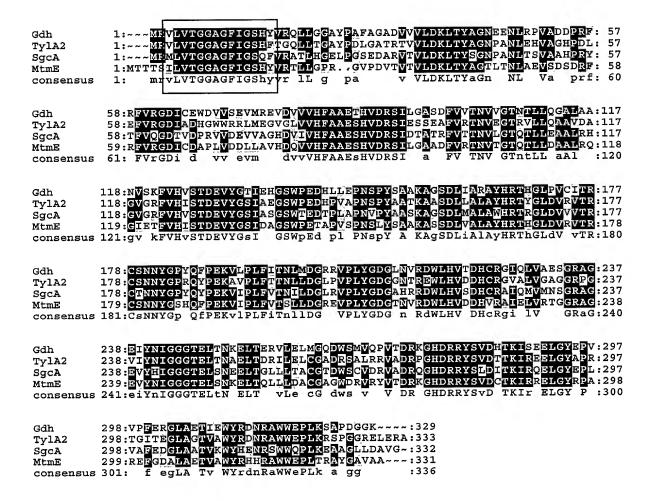
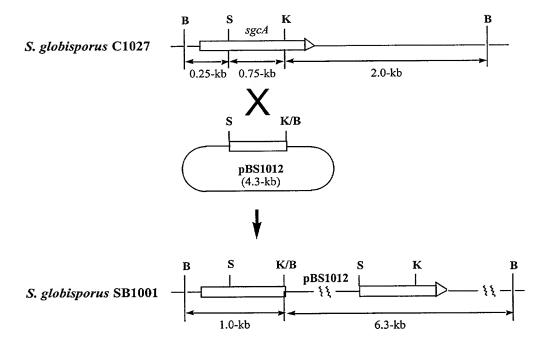
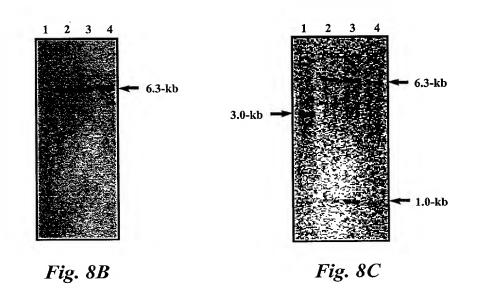


Fig. 8A





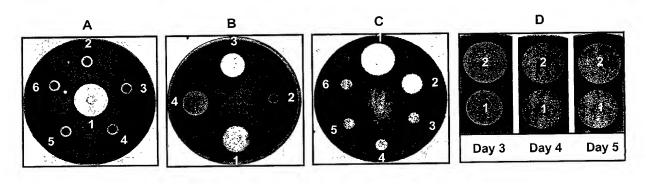


Fig. 9A Fig. 9B Fig. 9C Fig. 9D

PATENT APPLICATION DECLARATION

(Attorney's Docket No.: 2500.125US2)

Each of the Applicants named below hereby declares as follows:

1. My residence, post of	fice address and country of citizenship given below
are true and correct.	
2. I believe I am the orig	ginal, first and joint inventor of the subject matter
which is claimed and for which a patent is	sought in the patent application entitled "GENE
CLUSTER FOR PRODUCTION OF THE EN	NEDIYNE ANTITUMOR ANTIBIOTIC C-1027,'
Serial No, filed January	5, 2000, and I have reviewed and understand the
contents of the specification, including its cla	aims.
3. I acknowledge my dut	ry to disclose to the Office all information known to
me to be material to patentability of this appli	ication, in accordance with 37 C.F.R. Section 1.56
which is defined on the attached page.	
I further declare that all statem	nents made herein of my own knowledge are true and
that all statements made on information and	belief are believed to be true; and further that these
statements were made with the knowledge th	hat willful false statements and the like so made are
	under Section 1001 of Title 18 of the United States
Code, and that such willful false statements r	nay jeopardize the validity of the application or any
patent issuing thereon.	
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	Ben Shen
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	Wen Liu
Residence and	Institute of Medicinal Biotechnology
Post Office Address:	Tiantan, Beijing, 100005, China (Citizenship: Peoples Republic of China)

Date:		
	-	Steven D. Christenson
	Residence and	1079 Monarch Lane
	Post Office Address:	Davis, California 95616
		(Citizenship: United States)
_		
Date:		Coast Standage
		Scott Standage
		63 Tudor Road
	Post Office Address:	Bornet, Herts, EN5 5NW, United Kingdom

(Citizenship: United Kingdom)

Section 1.56 Duty to Disclose Information Material to Patentability.

- A patent by its very nature is affected with a public interest. The public interest is (a) best served, and the most effective patent examination occurs when, at the time an application is being examined, the Office is aware of and evaluates the teachings of all information material to patentability. Each individual associated with the filing and prosecution of a patent application has a duty of candor and good faith in dealing with the Office, which includes a duty to disclose to the Office all information known to that individual to be material to patentability as defined in this section. The duty to disclose information exists with respect to each pending claim until the claim is cancelled or withdrawn from consideration, or the application becomes abandoned. Information material to the patentability of a claim that is cancelled or withdrawn from consideration need not be submitted if the information is not material to the patentability of any claim remaining under consideration in the application. There is no duty to submit information which is not material to the patentability of any existing claim. The duty to disclose all information known to be material to patentability is deemed to be satisfied if all information known to be material to patentability of any claim issued in a patent was cited by the Office or submitted to the Office in the manner prescribed by §§ 1.97(b)-(d) and 1.98. However, no patent will be granted on an application in connection with which fraud on the Office was practiced or attempted or the duty of disclosure was violated through bad faith or intentional misconduct. The Office encourages applicants to carefully examine:
 - (1) prior art cited in search reports of a foreign patent office in a counterpart application, and
 - (2) the closest information over which individuals associated with the filing or prosecution of a patent application believe any pending claim patentably defines, to make sure that any material information contained therein is disclosed to the Office.
- (b) Under this section, information is material to patentability when it is not cumulative to information already of record or being made of record in the application, and
 - (1) It establishes, by itself or in combination with other information, a prima facie case of unpatentability of a claim; or
 - (2) It refutes, or is inconsistent with, a position the applicant takes in:
 - (i) Opposing an argument of unpatentability relied on by the Office, or
 - (ii) Asserting an argument of patentability.

A prima facie case of unpatentability is established when the information compels a conclusion that a claim is unpatentable under the preponderance of evidence, burden-of-proof standard, giving each term in the claim its broadest reasonable construction consistent with the specification, and before any consideration is given to evidence which may be submitted in an attempt to establish a contrary conclusion of patentability.

- (c) Individuals associated with the filing or prosecution of a patent application within the meaning of this section are:
 - (1) Each inventor named in the application;
 - (2) Each attorney or agent who prepares or prosecutes the application; and
 - (3) Every other person who is substantively involved in the preparation or prosecution of the application and who is associated with the inventor, with the assignee or with anyone to whom there is an obligation to assign the application.
- (d) Individuals other than the attorney, agent or inventor may comply with this section by disclosing information to the attorney, agent, or inventor.

GENE CLUSTER FOR PRODUCTION OF THE ENEDIYNE ANTITUMOR ANTIBIOTICC-1027

SEQUENCE LISTING

SEQ ID No. 1. C-1027 gene cluster DNA sequence from 1 to 42,000, ORF-(-7) to ORF-26

	-																				CGG	60
7-*	1	CAG	CTC	BAGA	TCI	rcci L	rage	GCC	CAC	CGCC	CTCA S	TCC	CCC	TAA	3CC7	GC.	rtcc	CTCC	CCA	CGG	+ GCC G	-
	61	CGA	CGC	ССТС	CGC	GCG?	AAGO	GCC	GTT	CCI	TGA	GT7	rcg2	AGG(CCGC	TG	GCGI	AGGI	CGA	CGT	'GGT +	120
.7	01	GCT	'GC	GA(CGC	CGC:		CCGC	CAZ	AGGZ	AACI	CA?	AGC:	rcco	GGC	CAC	CGCI	rcci	GCI		CCA	-
																					'ACG	100
- 7	121	GGC	GCI		rcc:	ΓAG			\GC(CCC		CCG	GT	CCC	GCG.	rcg(GGZ	AGC	CAGI		TGC Y	-
	101																				CCT	240
- 7	181	CCC	CAC'	rcco	GG(GAC.	rgco	CAG:	rgg <i>i</i>	AGC:	rtco	FTC	GCC2	AGC	ACC	CTG	GCC	CGC	AGC1	CGC L	CGGA	-
	241	CCC	CCG	rcc	GCT'	rccz	ACAZ	AGGZ	ACG	ACG	CCGC	GA(CAG	GAC'	rcc	CGT	GCG	GCC	rcgz	ACCA	GTC	300
- 7	241	GGC	GC/		CGA	AGG'	rgt:	rcc:	rgc:	rgc	GGC(CCT	GTC	CTG	AGG	3CA	CGC	CGG	\GC]		CAG	-
	301				-+-			+-		- -					-+-	-		+			TCC	360
-7				AGC: D			AGG2 D													rtg <i>i</i> F	AAGG K	-
	261	ACC	JAA:	GCC(CAG	CGC	CGG	GGC	CAG'	rcg	CGC	CGGʻ	TCG	GCC'	TCC'	rgg	TTG	GCC	CAG:	rtg <i>i</i>	ATGA	420
- 7	361	TG	CTT S	CGG	GTC	GCG	GCC(CCG	GTC.	AGC	GCG	GCC.	AGC	CGG.	AGG.	ACC.	AAC	CGG	GTC/	AACT N	TACT	-
	421	AG.	rcg.	AGC	ACG	TCC	TCG	CGG	AAC	ACC	GAC	ATC	CTG	CCG	GCC'	TGG.	ATA'	TTG.	AAG	ACGT	rggt +	480
-7	421	TCZ F	AGC' D	TCGʻ	TGC.	AGG.	AGC E	GCC'	TTG'	TGG	CTG' S	ГAG	GAC	GGC	CGG. A	ACC	TAT.	AAC	rrc: F	rgc?	ACCA	-
	481																				CGCT	540
-7	101	GG	GTC	CCC.	AAC	GGC	AGT	GCC.	ACT.	ATC	CGC'	TGC	GGC	CGG	CTC	GCC	ATc	CGC	CGC	GCG(R	GCGA	-
	541																				GCCG	600
-7	311	GG'	rcc	TCC	TGC	TGA		TCG	CCA	GAA	GAG	CGC	TTT	ACT	TCG	TCC	GCA	TAG		CAG	CGGC	_
	CO1																				ATGA +	660
-7 - <	ρU1	AC	GGA	CGG	TCC	GGG	CGG	GGA	TGC	TGG	TCG	TGG	GAC	CCC	:GCG	CGT	'GGG	CAG	TAC		TACT	-

	661	TCGGAGGGGGCGACTGAGTCCCGCGCGCGCGCGCGAGGGCAGCCACAGGAGCGACTGA	720
	721	GGAAGTTCCCTGACCTGGCGTCAACTCCACTGATCCGTAAGGGGATCGCGGGAGTGGATA	780
	781	CGGGTCAGGTCGTGCACGATCGTGGCACCAGACAGATCACCACGTCGATAGGCACTCGTG+ GCCCAGTCCAGCACGTGCTAGCACCGTGGTCTGTCTAGTGGTGCAGCTATCCGTGAGCAC	840
		AGCCGCGCCCGGGGCTCGACGGGGCGCCACCGGCAGGGGCCGCCGTGATCAGCCGG TCGGCGCGGGGCCCCGAGCTGCCCCGCCGGCGCGCGCACTAGTCGGCC	900
-6-*	901	AGCCTGTCCGGGGGCGTGCGGGGGCGTCAGCTGTCGATGTCGGGAACGCCAGGGACG TCGGACAGGCCCCGCACGCACGCCCCGCAGTCGACAGCTACAGCCCTTGCGGTCCCTGC * S D I D P V G P V	960 -
		TCGATCTCGGTGCGGGCGTAGTGGTTGAAGTAGTTGGTGTAGAGGTTCACGGCCACGTGG+ AGCTAGAGCCACGCCCGCATCACCAACTTCATCAACCACATCTCCAAGTGCCGGTGCACC	1020
-6		D I E T R A Y H N F Y N T Y L N V A V H ACGAAGACCTCGGCGAGCTCGGTGTCCGTCCATCCCTGTGCCACGGCCGCGTTCCACGAG TGCTTCTGGAGCCGCTCGAGCCACAGGCAGGTAGGGACACGGTGCCGGCGCAAGGTGCTC	1080
-6	1081	V F V E A L E T D T W G Q A V A A N W S GCGTCAGACGCCTCGCCCACTTCGCCGGCGATCTCCCTGGCCACCTGGACCAGTGCTTCG	1140
-6		CGCAGTCTGCGGAGCGGGTGAAGCGGCCGCTAGAGGGACCGGTGGACCTGGTCACGAAGC A D S A E G V E G A I E R A V Q V L A E AGCTTCACGTCGCCGGGGCGTCCCCCGGCGAATCGCCACGGTCTCCTCCAGCGTGAAA	-
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-6	1261	CCCACGGCGAGGGCGATCGCCTCGCGTGTGCGGGCGTCGAACGTTCCATGTTCGGCGACG+ GGGTGCCGCTCCCGCTAGCGGAGCGCACACGCCCGCAGCTTGCAAGGTACAAGCCGCTGC G V A L A I A E R T R A D F T G H E A V	
-6	1321	GCTCCGGTGATCGCGGCGTAGGTTTCCAGGACCACGGGGGAATGGGCCATTCCCCCGTGG+ CGAGGCCACTAGCGCCGCATCCAAAGGTCCTGGTGCCCCCTTACCCGGTAAGGGGGCACC A G T I A A Y T E L V V P S H A M G G H	
-6	1381	ATGTTGAGCACTCGCCCGAACCGCTTCTCCAGTCGGCGCAGGATGTCTCCGCCGGCTGCG TACAACTCGTGAGCGGGCTTGGCGAAGAGGTCAGCCGCGTCCTACAGAGGCGGCCGACGC I N L V R G F R K E L R R L I D G G A A	1440
-0	1441	GGTGCGGTGTCGATGGTGTGGACGGGAATCCGCGGCATGGGAATGCCTCTCCTCGTAGTG CCACGCCACAGCTACCACCACCAGCCCTTAGGCGCCCTTACGGAGAGGAGCATCAC	
-6	<	PATDITHVPIRPM	-

		ATGGGA	+			+-			+				+			- + -			+	1560
		TACCCT																		
		GCCCTC																		1620
	1301	CGGGAG																		
		GCACCA																		1600
	1621	CGTGGT								TCT	'CCG	GCI	CGI	'GGA	\GG!	\GC(CGGI	CCC	CGCC	1680
-5-*										*	L	G	L	V	E	Е	A	L	A	-
	1681	TGCCCC																		1740
- 5		ACGGGG	CTTC	TGCC	CGG?	AGC1	'AGA	ACC	GCT	TCC	:GGT	CCA	AGCG	CAC	CAC	CAC	CTCC	CACA		_
-5																		_		
		CGGCGA	4			+-			+				· + - -			-+-			+	1800
-5		GCCGCI D A		CCTC			C C							Q Q	E E	v V	_	IACA	Y Y	-
		GGGCGG																		
	1801	CCCGCC																		1860
-5		R A	A I	L	I	R	D	R	V	Q	E	P	T	E	V	V	P	D	I	-
	1061	GGTCGG																		1920
_	1001	CCAGCC	AGTO	GGGC	TCC	TTC?	rgce	3CCC	CGCC	GTC	CCC	CCGI	rcc <i>i</i>	ACCI	AGT(GCC.	rgc	CACC	BAGT	1,520
-5		P D																	S	_
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	1921					+-			+			- - -	-+			+			+	1980
-5	1921	CCTGGG	CGAC		AGG	+- CGA/	AGCO	GGCC	+ CGGI	CAA	GCI	CT	- + \TCI	TCI	AAC	+ · GGG(CGGZ	AACI	rcga	1980 -
-5	1921	CCTGGG	R E	CCCC P	AGG(D	CGAZ A	AGCC E	G G	GGT A	CAA L	GCI E	rcta L	Y Y	TC# F	AAC N	GGG(CGG7 A	AACT K	rcga L	1980
-5		CCTGGG L V GGAAGA	CGAC R E	GGGC	AGG D AGC	+- CGAZ A AGTI	AGCC E	GGCG	GGT A A	CAA L CGA	GCT E	rctz L	ATCT Y	TTC# F	AACO N	+ GGG(G GTG(CGG/ A GAG	AACT K	rcga L rggt	-
-5 -5		CCTGGG L V	R F	GGGC	AGGC D CAGC STCG	CGAZ A AGTI	AGCO E TCGO	GCGT	CGGT A CAGT	CAA L CGA	ATGT	CTI L CGI	ATCT Y AGG(TTCI F CTGT	AACO	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGZ A GAGT	AACT K rcct	rggt	-
	1981	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG	GCGAG R F AGCTT CCGAF L F	GCCCC F GGGC ACCCC F GGCAG	AGGC D AGC TCG TCG L	CGAAAAGTT	AGCC AGCC AGCC E	GGCG GGCG GCGGA	CGGT A CAGT CAGT Y Y	CGA	AGCI E ATGI ACI I	CGA CGA AGCT	ATCT Y AGGGG -+ FCCG L	TTC/ F CTGT SAC/ S	AACO N TGCO ACGO H	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A GAG: CTCA S	AACT K rcct AGGZ D	rcga L rggt + Acca Q	- 2040 -
-5		CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGGGC	GCGAC R F AGCTT CGAA L F CCGGC	GCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGGG D AGC TCG TCG L GTG CCAC	CGAZAGTTA	AGCO AGCO E AGCO E	GGG	ATCA	CGAACCAACCAACCAACCAACCAACCAACCAACCAACCA	ATGT EACA	CAAA	AGGO	TTCA F CTGT SACA S TCGC	AACO N TGCO ACGO H	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A GAGC CTCA S AAGC	AACI K ICCI AGGA D CGCC	reget L reget + ACCA Q CCCA +	-
	1981	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGGCC D G	GCGAC R I AGCTT CCGAA L I CCGGC GGCCC G I	GCCCC FCGGGC CCCCC CP GGCAG F CCGTC	AGGG D AGC TCG TCG L GGTG CCAC	CGAZ A AGTT +- TCAZ L TGTZ ACAT	AGCO E TCGO AGCO E ACGO V	GGCZ A CCGZ A CCGZ GGCZ	CGGT A CAGT ATCA Y ATGO I	CGAA L CCGA CGGG CGGG R	ATGT EACA I	CAA	AGG AGG L TCCT AGG E	TTCA F CTGT GACA S CCGC	AACC N TGCC ACGC H GCGCC A	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A GAGC CTCA S AAGC TTCC	AACT K FCCT AGGZ D CGCC R	rGGA L rGGT + ACCA Q CCCA + GGGT G	- 2040 -
-5	1981	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGGCC D G	GCGAG R I	GCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGGC D CAGCA CTCG L CCACA T	CGAZ A AGTT TCAZ L TGTZ ACAT H	AGCO E AGCO E ACGO V ATGA	GGCZ A CCGZ A CCGZ G	ATCA Y ATGG I TCGI	CCAA L CCGA AGCT D CCGGG R	AGGA	CCTA CCGA AGCC D GTTTT T ACCGG	AGG	F F F F F F F F F F F F F F F F F F F	AACGO H CCCCC	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA GAGC CTCA S AAGC TTCC	AACT K ICCT AGGZ D CGCC R	rggt + ACCA Q CCCA + GGGT G	- 2040 - 2100 -
-5	1981	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGGCC D G	GCGAC R F AGCTT CCGAL L F CCGGC G F CCGGC G F AGCAL	GCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGGO AGGO AGGO AGGO AGGO AGGO AGGO AGGO	CGAZ A AGTT CAZ TCAZ ACAT ACAT ACAT CCGCT	AGCCC E FCGCC E ACGCCC V ATGA	GGGCCGAACCGAACCGAACCGAACCGAACCGAACCGAA	CGGTA ACGGTA	CCGAACCCCAACCCCAACCCCAACCCCAACCCCAACCCCAACCCC	AGGA	CCAAAAAACGGGCCCCCCCCCCCCCCCCCCCCCCCCCC	-+ Y AGGGG -+ FCCC L FCCCGG E	F CTGT	PGCCGCCGCCGCCGCCGCCGCCGCCGCCGCCGCCGCCGCC	+: GGGGG GTGG CACC T CTGACC S GGGG S GGGG CCCC	CGGA A GAGC CTCA S AAGC F TCGA	K FCCT AGGA D CGCC R AGCT	rggt + ACCA Q CCCA + GGT G	- 2040 - 2100 - 2160
-5 -5	1981	CCTGGG L V GGAAGA CCTTCT Q F CGCCGC D G GGACTT CCTGAA L V	GCGAC R I	GCCCC GCCCC GCCCCCCCCCCCCCCCCCCCCCCCCC	AGGC D AGCZ T CCACZ T SAGGC CTCC	AGTTCAACA: H GCGC: A	AGCCC E FCGCC AAGCCC E ACGCC V V ATGA	GGCGA GCGA A CCGA GGCT GGCT TTCA	CGGTA A FAGTA Y ATGO I FCGTA AGCO D	CCGAACCT D CCGGGC R CCGAACCT D	AGGA	CCTA L CCGA AGCT D TTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	Y AGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	F F F F F F F F F F F F F F F F F F F	TGCC N H GCGCC A CCGCC A	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A SAGG CTCA S AAGG F TCGA D	R FCCT AAGGA D CGCC R AGCT L	rggt + ACCA Q CCCA + GGT G rtga + AACT K	- 2040 - 2100 - 2160
-5 -5	1981 2041 2101	CCTGGC L V GGAAGA CCTTCT Q F CGCCGC D G GGACTT CCTGAA L V GGGACA	GCGAC R I	GCCCC GCCC GCCCC GCCC GCCCC GCCC GCCCC GCCCC GCCCC GCCCC GCCCC GCCCC GCCCC GCCCC GCCC GCCCC GCCCC GCCCC GCCC GCCCC GCCC GC	AGGG D CAGCL CAGC	AGTTCAACA: H GCGC: A GTGC: A GTGC: A GTGC: A	AGCC E FCGC E ACGCC V ATGA ATGA ATGA ATGA ATGA ATGA ATGA AT	GGCCGAAAGCCGGAAAAGCCGGAAAAGCCGAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCCGAAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCGAAAAGCCCAAAAAGCCCAAAAGCCCAAAAAGCCCAAAAAA	CGGT A CAGGT Y ATCA Y ATGC I CGGT I CGGT A CGGT A A A A A A A A A A A A A A A A A A	CGAAL COGA COGA COGA COGA COGA COGA COGA COGA	AGGZ	CCTA L CCGA AGCT D TTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	-+ Y Y AGGGG FCCCC L FCCCC E CCGGG+ GGCCC G	TTCA F CTGT SACA S CCGG E	TGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A SAGG CTCA S AAGG F TCGA D	PAGGO	FGGT+ ACCA Q CCCA+ FGGT G FTGA+ AACT K CAGC	- 2040 - 2100 - 2160
-5 -5	1981 2041 2101	CCTGGC L V GGAAGA CCTTCT Q F CGCCGC GCGGCC D G GGACTT CCTGAA L V GGGACZ	GCGAC R I AGCTT CGAA L I CCGGC G I CGGCC G I AGCAC AGCAC E I AGCCC CGGCC CGCC CCC CGCC CCC CGCC CCC CGCC CCC C	GCCCC GCCC GCCCC GCCC GC	AGGG D CAGCL CAGC	AGTTCAACA: H GCGC: A GTGCCACCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGCC E FCGC E ACGCC V ATGA ATGA ATGA ATGA ATGA ATGA TTTCA	GGCCGA A CCCGA G G G G G TTTCA F	TAGCA TA	CGAA L CCGAA AGCT D CGGGG R CCGA D	AGCTACACACACACACACACACACACACACACACACACAC	CCAAAA V	-+ Y AGGGG-+ FCCCC L FCCCC E CCGGG-+ GGGCC G TGTC AAGACACACACACACACACACACACACACACACACACA	TTCA F CTGT SACA S CCGG G G GCCGGGCCGGGCCGGGCCGGGCC	FGCGCA A CTGC SACCES	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA A SAGG CTCA S AAGG F TCGA D TCCA	PAGGO	rggt L rggt + ACCA Q CCCA + gggt G rtga + K CAGC	- 2040 - 2100 - 2160
-5 -5	1981 2041 2101 2161	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGCCC D G GGACTT CCTGAA L V GGGACA CCCTGT L S CTCGGA	CCGGC G I CGGCA AGCA AGCA AGCA AGCA AGCA AGCA AG	GCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	EAGCO T CCACC	AGTTCAM AGTTTCAM L TTGTM ACA: H GCGM A GTGM T GCGC T GCCC T	AGCCC E ACGCCC V ATGA ATGA ATGA TTTCA TTCGCCC	GGCCGA ACCCGA FITCA FITCGA D		CCGAACCO R CCGGGC R CCGGC R CC	AGGZ	CGC	-+ Y AGGGG FCCC L FCCCT ECGGG E TGTC ACACAC H AGGG	TTCZ F F CTGT S S FCGG G CCGG G CCGGG A	ACGC A CCCC S G G G G G G G G G G G G G G G G	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGA GAGG AAAGG F TCGA AAGG TCCA CGGGA CGGGGA CGGGGGGGA CGGGGGGA CGGGGGGA CGGGGGGA CGGGGGGA CGGGGGGGA CGGGGGGA CGGGGGGGG	AGGCTCCCL	rcga L rggt + Acca Q ccca + gggt G rtga + Aact K cagc +	- 2040 - 2100 - 2160 - 2220
-5 -5	1981 2041 2101 2161	CCTGGG L V GGAAGA CCTTCT Q F CGCCGC D G GGACTT CCTGAA L V GGGACA CCCTGT L S CTCGGAA CCCTGT CCCTCT CCCTC	GCGAC R F AGCTT CCGAA L F CCGGC G F CCGTT AGCCA E 1 AGCCA L F AGCCA AGCCA L F AGCCA L F AGCCA AG	GCCCC GCCCC GCCCCCCCCCCCCCCCCCCCCCCCCC	AGGG D AGCACACACACACACACACACACACACACACACACACA	AGTTCAACACACACACACACACACACACACACACACACAC	AGCC E FCGC E ACGC V ATGA TTCC F TCGC	GGCGGAAGCCGAAGCCGAAGCCAAGCCAAGCCAAGCCA	TAGO AGCT CGGT	CGAA L CGGA CGGGG CGGGG R CGGGG R CGGGG R CGGGG R CGGGGG R CGGGGGG R CGGGGGGGG	ACCA ACCA	CCTA L CCGA D TT T CAAAA T V ACGG V CCGCG	-+ Y AGGG -+ FCCC L FCCC E CCGG G FGTC -+ H AGGZ H AGGZ H AGGZ TCCC	TTCA F CTGT SACA S CCGG G G GCCGGG A AACC	PGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGGAA SAGGTTCGA TCGA TCGA TCGA CGGGGA CGGGGCC	AGGC AGGGC AGGCC A	rGGA L rGGT+ ACCA Q CCCA+ EGGT G rTGA+ AACT K CAGC+ GTCG C	- 2040 - 2100 - 2160 - 2220
-5 -5	1981 2041 2101 2161	CCTGGG L V GGAAGA CCTTCT Q F CGCCGG GCGCCC D G GGACTT CCTGAA L V GGGACA CCCTGT L S CTCGGA	GCGAC R F AGCTT CGAF L F CCGGC G F CCGTT AGCAF E F AGCAF	GCCCC GGGCA GGCAC CCGTC CCGTC CCGTC CCGTC CCGGCC CCGGCC CCGGCC CCGCC CCCC CCCC CCCC CCCC CCCC CCCC CCCC	AGGGETGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	AGTTTCAAACACACACACACACACACACACACACACACAC	AGCC E ACGC V ATGA AAGT TACT TTCG AAGCC E AAGCC	GGCCGA ACCGA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA AAGCCA	TAGT Y ATGC I FCGT AGCT L AGCT AGCT AGCT AGCT AGCT AGCT AGCT AGCT	CCGAACCTACCAACCAAC	AGGA	CGC, R	-+ Y AGGGG ICCT L ICCT AGGAG E CCGGG G H AGGAC ACAC H AGGAC L L	TTCZ F F CTGT S S ICGGC CCGGC G G CCGGC A AAC' F	TGCC CCCC GGGG GGGG TGCC ACG	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CGGGA GAGG TTCC TCCA AGGC TCCA CGGGA CGGGA CGGGA CGGGCC R	AGGC AGGGC AGGCC A	FGGA L FGGT+ ACCA Q CCCA+ FGGT G FTGA+ K CAGC CAGC+ FTCG C TAGC Y	- 2040 - 2100 - 2160 - 2220

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-5		GGAGCTACCCTCACCGCCCATCTCCTCCGACTCCCGCCTCCCACGCTACTGGCGGACGA G E I P T A P Y L L S L A S P A I V A Q -	-
		TCAGGGGGCGTCCGTGAGCTGCCGTGCGGCGCGCAGATAGGTTTCGGCCCGCACCTGGT	2400
-5		AGTCCCCGGCAGGCACTCGACGGCACGCCGCGCGTCTATCCAAAGCCGGGCGTGGACCA K L P R D T L Q R A A R L Y T E A R V Q -	-
	2401	AGCGGAAGGGCCCTTGGGTGATGCTGGGGAGCTGCCGGGTGTGCCCGTCTGCGAAGGGGA	2460
-5		TCGCCTTCCCGGGAACCCACTACGACCCCTCGACGGCCCACACGGGCAGACGCTTCCCCT Y R F P G Q T I S P L Q R T H G D A F P	_
	2461	TGACAGCGCCGTCGGGCGAGAGGGTGTCGAGGCCGGTCACGGGGTAGGTGGCGAAGCTCG	2520
-5		ACTGTCGCGGCAGCCCGCTCTCCCACAGCTCCGGCCAGTGCCCCATCCACCGCTTCGAGC I V A G D P S L T D L G T V P Y T A F S	-
	2521	GCTTGGACTGTTCACCGTCCACGAGGACGGGGCTGCCGACTCGTTCCAGTCGTGTCAGGG	2580
-5		CGAACCTGACAAGTGGCAGGTGCTCCTGCCCCGACGGCTGAGCAAGGTCAGCACAGTCCC PKSQEGDVLVPSGVRELRTL	-
	2581	TGTCCGCGACGGCCTGTTCCTGCTGTTTGGCCAGGTCCGTGGCGTCCAGGGTTCCCTGGG	2640
-5		ACAGGCGCTGCCGGACAAGGACGACAAACCGGTCCAGGCACCGCAGGTCCCAAGGGACCC T D A V A Q E Q Q K A L D T A D L T G Q	-
	2641	CATGCGCGGCAAGGCGTGCAGGAGTGTCGCGGAGCGCGGAAGGCTGCCGATCGGCTCAG	2700
-5		GTACGCGCCGTTCCCGCACGTCCTCACAGCGCCTCCGCGCCTTCCGACGGCTAGCCGAGTC A H A A L A H L L T A S R P L S G I P E	-
	2701	TGGCGATGGTCATGGCCGAAGAGTAGGGAAGAGGCTGGGTTTCGAACCACCGCAAAGCTT	2760
-5-	<	ACCGCTACCAGTACCGGCTTCTCATCCCTTCTCCGACCCAAAGCTTGGTGGCGTTTCGAA T A I T M -	
	2761	TGATTGCCGCTTTTTCAGGGGAAGTTGATGCGAAGTCGCCGAGCGGCGGAACGTGCTGAT	2820
		ACTAACGGCGAAAAAGTCCCCTTCAACTACGCTTCAGCGGCTCGCCGCCTTGCACGACTA GTATGGGGGGCGGGAGGAGCCTGCGGGGTTCTAGGAGCCGGTCGCGGCCACGGTGGAGGA	
		CATACCCCCGCCCTCCTCGGACGCCCCAAGATCCTCGGCCAGCGCCGGTGCCACCTCCT * S G T A A V T S S	
-4-	•	GGTGCCCAGCTGGGAGCGGGGGTCTTTTCGCCGACGCGGTTGGGCTCGATGGTGCGGGG	
-4	2881	CCACGGGTCGACCCTCGCCCCCAGAAAAGCGGCTGCGCCAACCCGAGCTACCACGCCCC T G L Q S R P T K E G V R N P E I T R P	
-4		GTCGACGGCCTCTCCGGGGGCACCTTGCCGGTAGACGCCTTCGGGGTCGGAGTCCCGGTC	
-4	2941	CAGCTGCCGGAGAGGCCCCGTGGAACGGCCATCTGCGGAAGCCCCAGCCTCAGGGCCAG D V A E G P A G Q R Y V G E P D S D R D	
-		ATGGGGGAGCAGAAGAAGACCCGGCGCCGGTACAGACCGCTGTCCGGGTCCGCTTCGGC	
-4	3001	TACCCCCTCGTCCTTCTTCTGGGCCGCCATGTCTGGCGACAGGCCCAGGCGAAGCCG	
-		GTCGGCCCGAGTTCGATGTAGCCGATCATGCGGCCGTCGCGGGCGTAGCGCGCTTGTT	
-4	3061	CAGCCGGGGCTCAAGCTACATCGGCTAGTACGCCGGCAGCGCCCGCATCGCGCCGAACAA D A G L E I Y G I M R G D R A Y R P K N	

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		CTTGCGCCGGGGGTCTTGTCCAGGGCCTGGCGGACGTAGTCGAGTCCCTCGGGATCTTC	0.0
	3121	GAACGCGGCCCCCAGAACAGGTCCCGGACCGCCTGCATCAGCTCAGGAGCCCTAGAAG	30
-4		KRRPTKDLAQRVYDLGEPDE-	
	3181	GAGCCACACGACCTTCGCCTCGTGAACGAGATCGCTGTCGGTCAGTAGCGAGCTCATGGC	40
	3101	CTCGGTGTGCTGGAAGCGGAGCACTTGCTCTAGCGACAGCCAGTCATCGCTCGAGTACCG	
-4-	<	L W V V K A E H V L D S D T L L S S M -	
		GGCGACCTCTCCTTCGTCGGCGTGCACCGGGTGGGGAAGCGGTGCCTGCGTGATGTGTGT	
	3241		00
		CCGCTGGAGAGGAAGCAGCCGCACGCCCCTTCGCCACGGACGCACTACACACA	
		TCGTCTGCGGCGGTGGCCGAGTGGTGCGGACCGCCCGTGGTGCCGGTTCTCGGCCAAA	
	3301		60
		AGCAGACGCCGCCACCCGGCGTCACCACGCCTGGCGGGCACCACGGCCAAGAGCCGGTTT	
		GCACGGGCAGGTACGTCCTGGGGCACTCACATCGTAGATGGGGTCCGCTTCCGCAGGGCA	
	3361		20
		CGTGCCCGTCCATGCAGGACCCCGTGAGTGTAGCATCTACCCCAGGCGAAGGCGTCCCGT	
		GTGCCTCCGGTCGGAGGACGTTCATTCGTCGGCTGCCAGAGCGAGGTTGGGGTAGAACTT	
	3421	+ 34	80
-3-1	*	CACGGAGGCCAGCCTCCTGCAAGTAAGCAGCCGACGGTCTCGCTCCAACCCCATCTTGAA * E D A A L A L N P Y F K -	
•			
	2401	CCGGCCGTTGGATTTGATCATGTCGGCAGGTGAGGCGAGGCCCACTTCCTGGCGGACCCG	40
	3481	GGCCGCCACCTAAACTAGTACAGCCGTCCACTCCGCTCCGGGTGAAGGACCGCCTGGGC	40
-3		R G N S K I M D A P S A L G V E Q R V R -	
		GGTGGCGAAGGCACGGGCGGTCCCGGGGCGGATGCCTTCACTGTGTGCGCACCAGGTGCT	
	3541	+ 36	00
		CCACCGCTTCCGTGCCCGCCAGGGCCCCGCCTACGGAAGTGACACACGCGTGGTCCACGA	
-3		TAFARATGPRIGESHACWTS-	
		GTAGGACGTGTAGAGAAGGCCCTGTTCGACGCGTAGCTCGCTGTTCTCGGGGTCGTGGAG	
	3601		60
-3		CATCCTGCACATCTCTCCGGGACAAGCTGCGCATCGAGCGCACAAGAGCCCCAGCACCTC Y S T Y L L G Q E V R L E S N E P D H L -	
_			
	2661	GCAGCACTCGGCGAGGAAGCGGCCGATGTGGTCCTCGGTGTTCGCGTATGCGCTGGTGGC	20
	3001	CGTCGTGAGCCGCTCCTTCGCCGGCTACACCAGGAGCCACAAGCGCATACGCGACCACCG	20
-3		C C E A L F R G I H D E T N A Y A S T A-	
		GATGCGGACCCGGTCGGGGCCGGCGAGTGTCGCGGGTGGCGAGGTAGCGGCGGCCCC	
	3721		80
		CTACGCCTGGGCCAGCCCGGCCGCTCACACAGCGCCCACCGCTCCATCGCCGCCCGGGG	
-3		IRVRDPGALTDRTALYRRAG-	
		TTCGGTGAGCCAGTGCAGGATCCCGGGGCCCTCGTCCTGGACGAGTTCGACAGCCAGGTT	
	3781	AAGCCACTCGGTCACGTCCTAGGGCCCCGGGAGCAGGACCTGCTCAAGCTGTCGGTCCAA	40
-3		E T L W H L I G P G E D Q V L E V A L N -	
	2011	GTCGATCTTGCGTTCGTCGGGGACGATCCGTTCGAAGGGCAGGAGGCGGATGCGGCCCA	100
	3041	CAGCTAGAACGCAAGCCCCTGCTAGGCAAGCTTCCCGTCCTCCGCCTACGCCGCGT	-
-3		DIKREDPVIREFPLLRIRRW-	
		GAAGGCGAAGCCGCCGGTGGAGACCTCGGGGCGGTGGTTGCCCAGCAGCCACAGCTTGTG	
	3901		60
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-3		F	A	F	G	G	T	s	v	E	P	R	H	N	G	L	\mathbf{L}	M	\mathbf{L}	K	Н	-
	3961		- -	+-				+ -			-+			4			- -	+			-+	4020
-3		GCAC(CCA(P																	GGC(-
	4021	GGTCZ		+-				+			-+-			+				+			-+	4080
-3		T	L L	L										N						C		-
	4081	GATG		+-				+			-+-			+				+			-+	4140
- 3		I	L	R	R	G	Н	L	E	Т	L	E	Т	s	Н	E	s	Y	A	G	R	-
-3	4141	CAGG'	TAC	+ : I'CC'	 TTT(GGG(CCG	+ CCC	 CGA	CGC	- + - AGC	 CGC	 ATC	+	 GGC'	 TCT	TAG.	+ ACC		TAG'	-+	
-3		GTCG	AGG	AGA	ACG(GAT:	TTG:	CCG'	TTC'	TTT	ccc'	rgg	CCG	TGG.	AGA.	AAG	GGC.	~ AGC	ACC	TGC	GC	
-3	4201	CAGC'	TCC'	rct'	TGC	CTA	AAC	GGC.	AAG.	AAA	GGG:	ACC	GGC	ACC	TCT	TTC	CCG	TCG	TGG		CG	
-3	4261	GGGC	TGC	+ AGT	 GGC	CAC	 TAC	+ CTC	 ATC	 GGC'	- + - TCT	 TCC	TCC	ACC	TCC	TTC		+ TAG		AGG	-+	
-3	4321	CCCT GGGA	TCG(GCG' + CGC.	TCA AGT	CTG	CCG. GGC	AAG + TTC	GTG CAC	TCT	TCG. -+- AGC	AGG TCC	AAA TTT	.CGG + 'GCC	TGC ACG	CAG GTC	CGG	GGG + CCC	CAC	.ccc	-+	4380
	4381	GTCC	TGG	GGG +	GAG	GCG	CTG	GTG +	GCG		-+-	-		+				+			-+	4440
-3		D CATA	Q CGG	P CCG	S TTG	A CGG	S AGG	$_{ m TCG}$	A ACC		s ccg		F .GGG		R CAC	T AGG	P GCG		P I GGG		R 'CC	-
-3	4441	GTAT	GCC	+ GGC	AAC	 GCC	 TCC	+ AGC	 TGG	TGA	-+- GGC	 AGT	CCC	+	GTG	TCC	CGC	+ ATC	CCC		-+ .GG	
-3	4501	CAGC	TCC	+ CAC	AGC	CCT	AGC	+ TCC	 CTC	TCC	-+- AGC	CCI	 CTC	+ CGG	AAA	.CGG	ACC	+ CAC	TCC		-+ CG	
	4561	CTTC		+				+		- 	-+-			+	. – – –			+			-+	4620
-3			M	G	Т	Т	S	L	Т	R	R	K	H	Н	H	L	E	R	D	Т	F	-
-3	4621	GTCG	 GGC	+ GCC	 CCI	'AGC	GAC	+	CCC	TAG	-+-	AGG	CGC	 DAT	AGA		CCGT	+	GTO		·-+ CCG	
	4681	AGCT		+				+			-+-			+		· ·		+			-+	4740

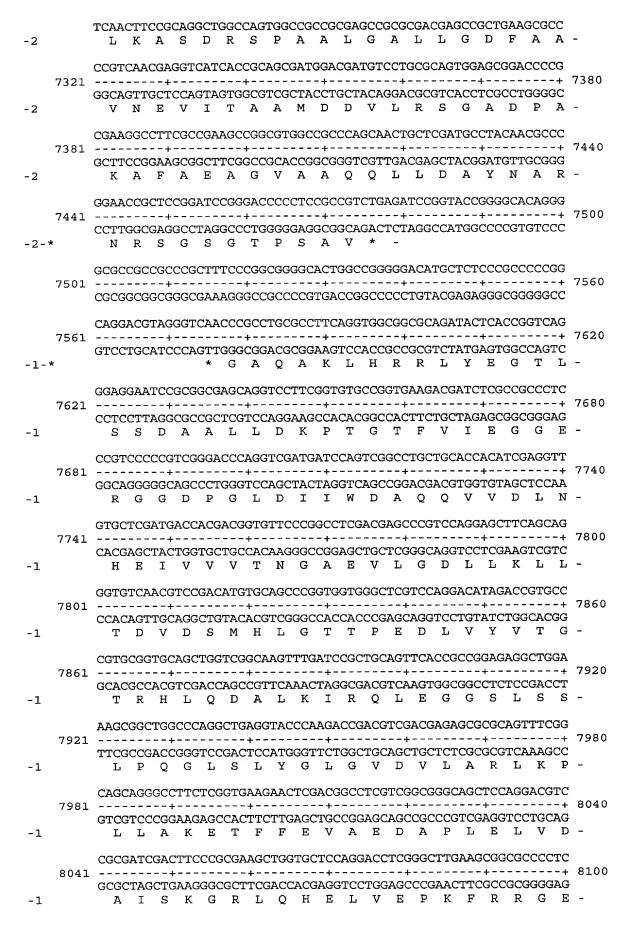
-3		A	K	E	G	G	A	R	K	M	R	Y	G	D	W	S	Y	W	G	L	G	-
	4741	CTCC																				4800
-3	4/41	GAGG'	TGC	ACG(GCC7	rtg <i>i</i>	ACCZ	AGT(GCC.	ATC'	TCT	GCC	TGC	TTC	TCG.	AAC	CGC.	AAC	GGC		\G	
	4801	GGTC	·	+-			- - -	+ -			-+-			+				+	- 		-+	4860
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	4861		- ·	+				+			-+-	-		+				+			-+	4920
-3		TCGC A												CAC T						TAGA M	_	-
	4921	CTGA		+				+			-+-			+				+			-+	4980
-3-4	<	GACT Q												CGC A						CAG	GΑ	
	4981	TCGA	GAT	GGA(GCG(GTC	3GG(CGG' +	TCC	CCG	CTG	CGG	GAA	CGG	CAT	GAA	TGA	TCT +	TCC	CGG'	TG -+	5040
		AGCT	CTA	CCT	CGC	CAG	CCC	GCC.	AGG	GGC	GAC	GCC	CTT	GCC	GTA	CTT	ACT	'AGA	AGG	GCC	AC	
	5041	CGGA GCCT		+				+			-+-			+				+			-+	5100
		CCGG	CCG.	ACG(GGG	GGA.	AGC.	AGG	GGC	:CGG	CAA	.CCG	GGT	GGC	:GGG	GCG	GCG	TGA	\GCG	AGG	GC	
	5101	GGCC																				5160
	5161	ACGA	GCG	GCC	CGG'	TAC	GGG	GGG +	AAG	GGC	TCG	TCI	CTC	CGT	GGG	GCG	GCA	+	TGT	GGT	CC -+	5220
		TGCT																				
	5221	TCGT AGCA		+				+			-+-		. – – -	+	. -			+			-+	5280
		GTCA	AGC	ATC	${f TTT}$	CGT	GAC	ACT	CGG	CGA	'GGG	ACI	GAZ	\GGG	ACI	GTC	CTTI	CGC	TAAE	'GAG	TG	E240
	5281	CAGT	TCG	+ TAG	AAA	GCA	CTG	+ TGA	.GCC	GCI	CCC	TG	ACT	rccc	TGA	CAC	SAAA	AGC(CTTA	CTC	AC	5340
	5341	TAGG		+				+			-+-			+				-+			-+	5400
		ATCC																				
	5401	CCAG		+				+			-+-				+ -			-+-			-+	5460
	F 4 6 1	TCGG	TGA	.GGG	GCC	CCG	GTG	GAG	GGF	ACTO	AGC	GT	CTG:	TATO	GAC	GCGI	ATA	AGA(GGG"	CTG	AA 	5520
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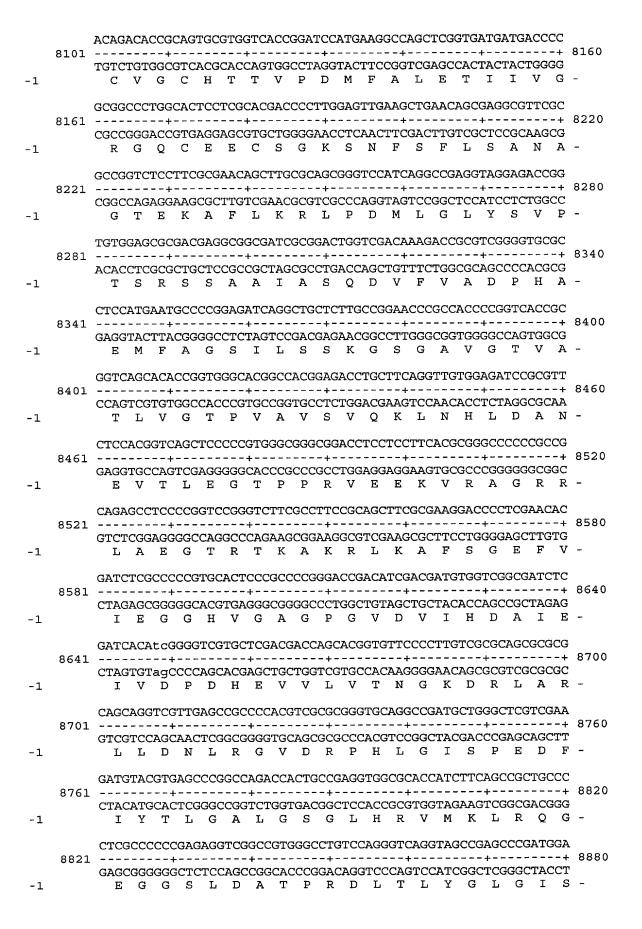
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	5701	GCGGAAGTTGAGTCCCTTCAGTCCCTTTTCGTGGGGTCGTATCCCCTCTGACTGCGTTGA++++++	5760
	5761	CCGTCGCCGCTCCGCGCAGGACCGAAGAGGGACCAAGTCCCTGCGCGGGGCGGCGACG	5820
	5821	GTAATCGTGCAGTGCCCCCCCCCGTTTCCCACAGCGAGTCGTCGCTCCCCTGTGAGGC	5880
	5881	CGGAGAGGGTCCTAGAACCCCTCAGGGGCCGTTCTGTGGCCCTCTGGGCCTCCTCGGC	5940
-2-:		CATTTACCCCATGGGGGCGCTTGGGGGGCGTCAGGAGGGCTTGTGAGGGCTCTGCCGGGAA GTAAATGGGGTACCCCGGGAACCCCCGCAGTCCTCCCGAACACTCCCGAGACGGCCCTT M R A L P G S	
-2	6001	GTGGCGGATTGCGCATGGCAGGAGATGCCCCGACAGCGGCCGGGAATCGACGATGTCCCC CACCGCCTAACGCGTACCGTCCTCTACGGGGCTGTCGCCGGCCCTTAGCTGCTACAGGGG G G L R M A G D A P T A A G N R R C P P	
-2	6061	CGACCCCTATCCAGCGTCCGCTGATCCTCAGGAGGCAGACCTTGCAGGCTCCAGAAGCGA GCTGGGGATAGGTCGCAGGCGACTAGGAGTCCTCCGTCTGGAACGTCCGAGGTCTTCGCT T P I Q R P L I L R R Q T L Q A P E A K	
-2	6121	AGAACGGCCGGTCCCCGGAGCAGCCGCAGGAAGAGCGGATCGTCCTGGACGTATGGCTGG TCTTGCCGGCCAGGGGCCTCGTCGGCGTCCTTCTCGCCTAGCAGGACCTGCATACCGACC N G R S P E Q P Q E E R I V L D V W L A	
-2	6181	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
-2	6241	CGGCGGAGTTCGAGCGCCCCCCCCCCGATACCGGGTCGACATCAACGGCCACGACTTCT+ GCCGCCTCAAGCTCGCGCGGGTGGGGGCTATGGCCCAGCTGTAGTTGCCGGTGCTGAAGA A E F E R A H P R Y R V D I N G H D F W	
-2	6301	GGACCATCCCCGAGAAGGTGGCGCGCGCCACCGCGGAGGGCAGGCCTCCGCACATAGCGG+ CCTGGTAGGGGCTCTTCCACCGCGCGCGCGCGCGCGCTCCCGTCCGGAGGCGTGTATCGCC T I P E K V A R A T A E G R P P H I A G	
-2	6361	GCTACTACGCCACCGACAGCCAGTTGGCGCGGGACGCGCGCAGGCCCGACGGGAAGCCGG CGATGATGCGGTGGCTGTCGGTCAACCGCGCCCCTGCGCGCGTCCGGGCTGCCCTTCGGCC Y Y A T D S Q L A R D A R P D G K P V	
-2	6421	TCTTCACCTCGGTGGAGGCCGCGTTGGCCGGCCGGACGGA	
		TGGTGGAGGACCTCGACCCCGTGGTGCGCGACTCCTACTCGTTCGGGGGGGG	

		ACCACCTCCTGGAGCTGGGGCACCACGCGCTGAGGATGAGCAAGCCCCCGCTCAACCACA	
-2		V E D L D P V V R D S Y S F G G E L V S -	-
		CGCTGCCGCTCACGGTCACCACCATGCTCTGCTACGCCAACTCCTCCCTC	6600
-2		GCGACGGCGAGTGCCAGTGGTGCTACGAGACGATGCGGTTGAGGAGGAGGAGCGCGCGC	-
	6601	CCGGTGTTCCGGAGTTGCCCCGTACCTGGGATGAGGTCGAAGCAGCCTGCCAGGCGGTGG	6660
-2	9901	GGCCACAAGGCCTCAACGGGGCATGGACCCTACTCCAGCTTCGTCGGACGGTCCGCCACC G V P E L P R T W D E V E A A C Q A V A	
		CCAGCGTCGACGGGGGGCCCGGTCACGGAATCACCTGGGCCAACGACGGCTGGGTTTTCC	 .
-2	6661	GGTCGCAGCTGCCCCCGGGCCAGTGCCTTAGTGGACCCGGTTGCTGCCGACCCAAAAGG S V D G G P G H G I T W A N D G W V F Q	
	6721	AGCAGGCCGTCGCCCTTCAGAACGGGGTGCTGACCGATCAGGACAACGGCCGCTCCGGCT	6780
-2		TCGTCCGGCAGCGGAAGTCTTGCCCCACGACTGGCTAGTCCTGTTGCCGGCGAGGCCGAQAVALQNGVLTDQDNGRSGCS	
	6781	CCGCCACGACGGTGGACGTCACATCGGACGAGATGCTGGACTGGGTCCGCTGGTGGACGC	6840
-2		GGCGGTGCTGCCACCTGCAGTGTAGCCTGCTCTACGACCTGACCCAGGCGACCACCTGCG A T T V D V T S D E M L D W V R W W T H	
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-2		TGGAGGTACTCGCGCCGGTAATGGAGATGTGCCCGCCCGGGAGCCTGACCCCGCCCCGCA L H E R G H Y L Y T G G P S D W G G A F	-
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-2	0301	AGCTCCGAAAGCAGGTCGTCTTCCAGCGTAAGTGGAAGCTGAGCAGGTTCCGGCGGGCCC E A F V Q Q K V A F T F D S S K A A R E	
	6961	AACTCATCCAGGCCGGTGCACAGGCCGGTTTCGAGGTCGCGGTGTTCCCGTTGCCCAGGA	7020
-2		TTGAGTAGGTCCGGCCACAGGCCCACAAGGCCCACAAGGGCAACGGGTCCT L I Q A G A Q A G F E V A V F P L P R N	-
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	7081	GACTCGACGAGACCACGCAGGACGGGCTGCTCGCTCTCACCCAGTACCTGATCAGCCCGG	7140
-2		CTGAGCTGCTCTGGTGCGTCCTGCCCGACGAGCGAGAGTGGGTCATGGACTAGTCGGGCC L D E T T Q D G L L A L T Q Y L I S P A	-
	7141	CCAACGCCGCGGACTGGCACCGCACCAACGGTTTCGTACCGGTGACCGGCGCGCGGGG	7200
-2		GGTTGCGGCGCCTGACCGTGGCGTGGTTGCCAAAGCATGGCCACTGGCCGCGCCCCC N A A D W H R T N G F V P V T G A A G E	-
	7201	AACTGCTGGAAGCGACAGGCTGGTTCGACCGCCGGCCGCAGCAACGGGTGGCCGGGAGC	7260
-2	,201	TTGACGACCTTCGCTGTCCGACCAAGCTGGCGGCCGGCGCGTCGTTGCCCACCGGCCCCTCG L L E A T G W F D R R P Q Q R V A G E Q	
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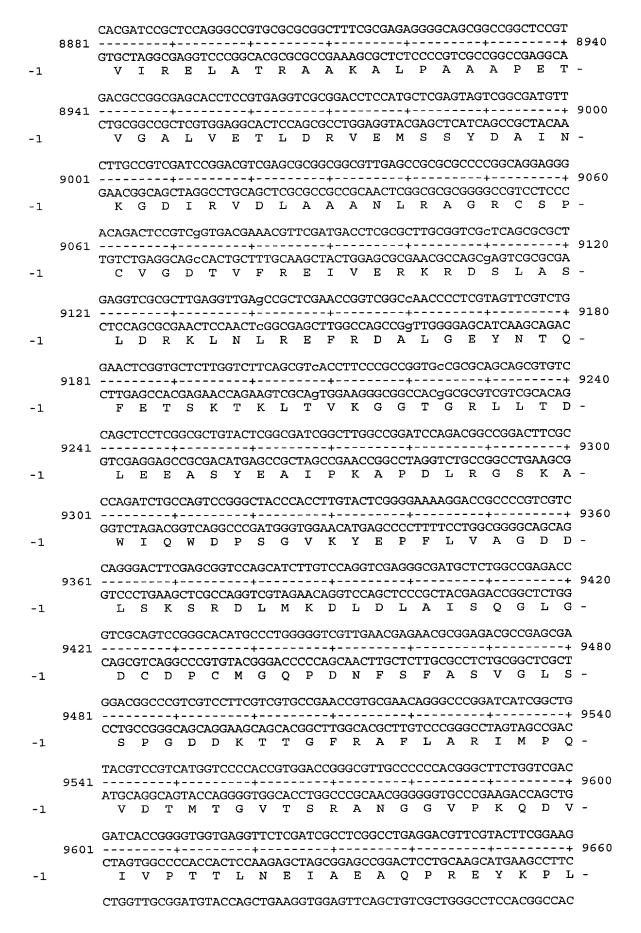


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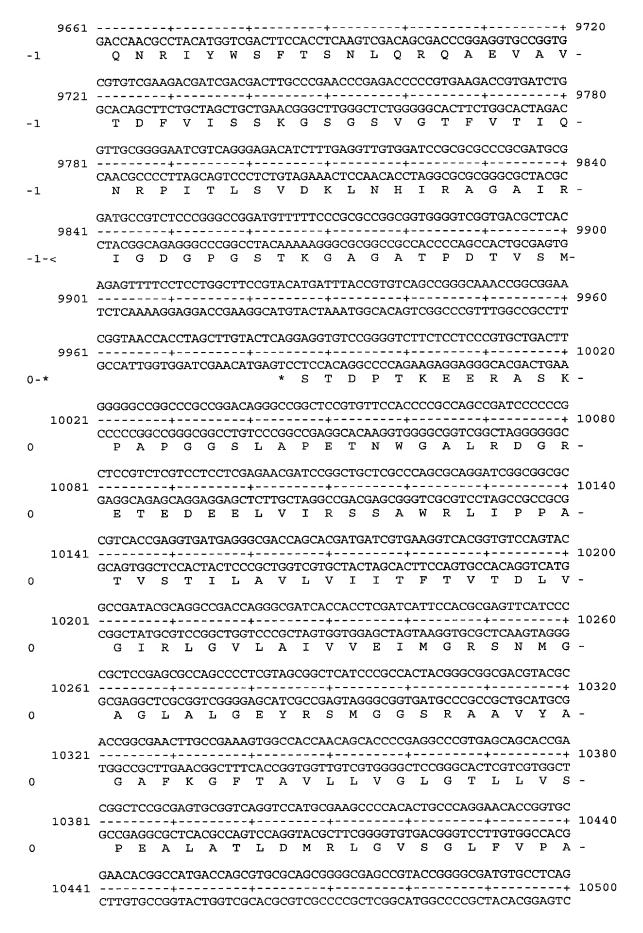


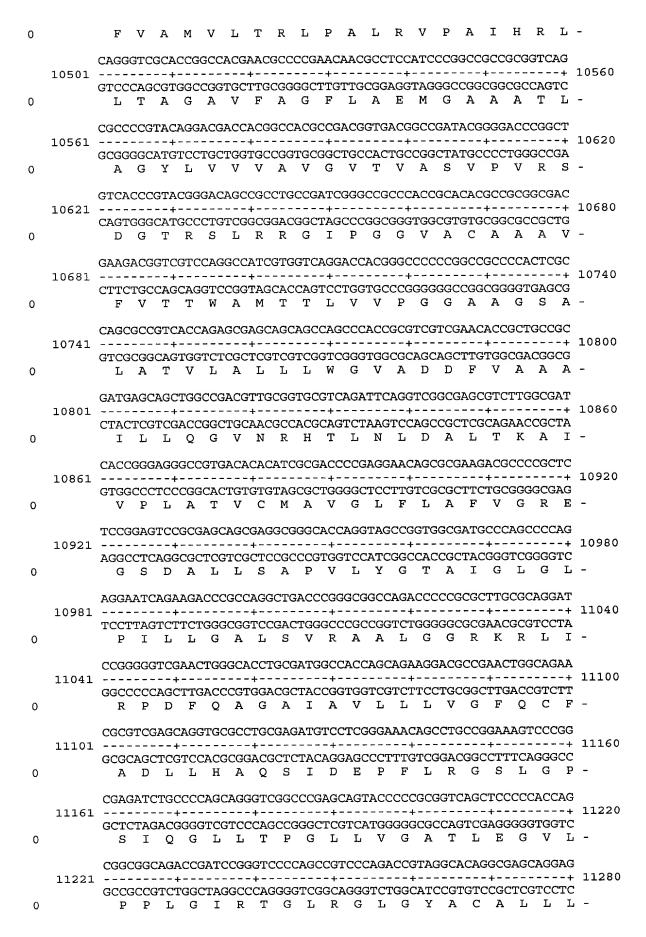
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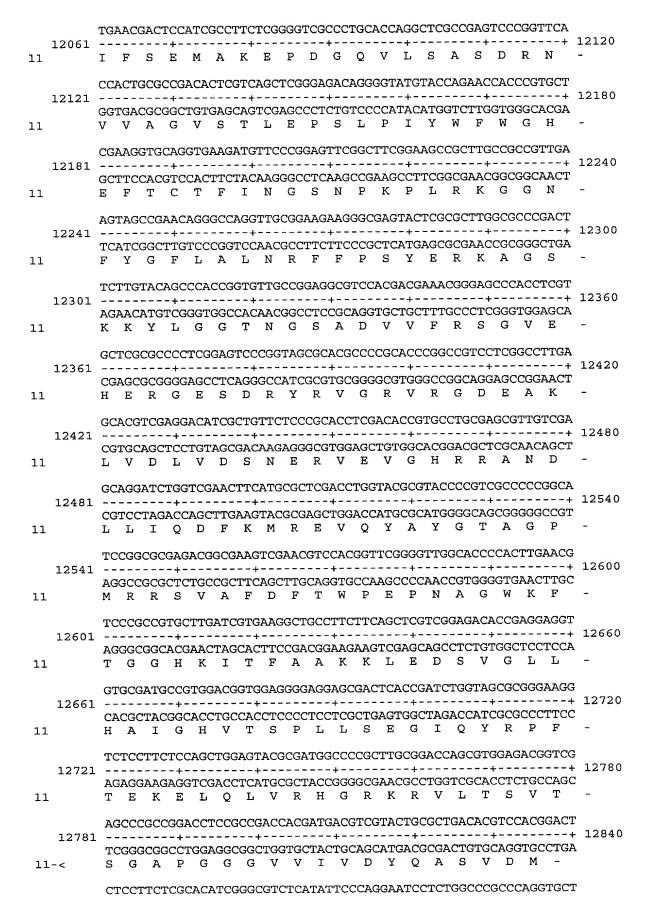




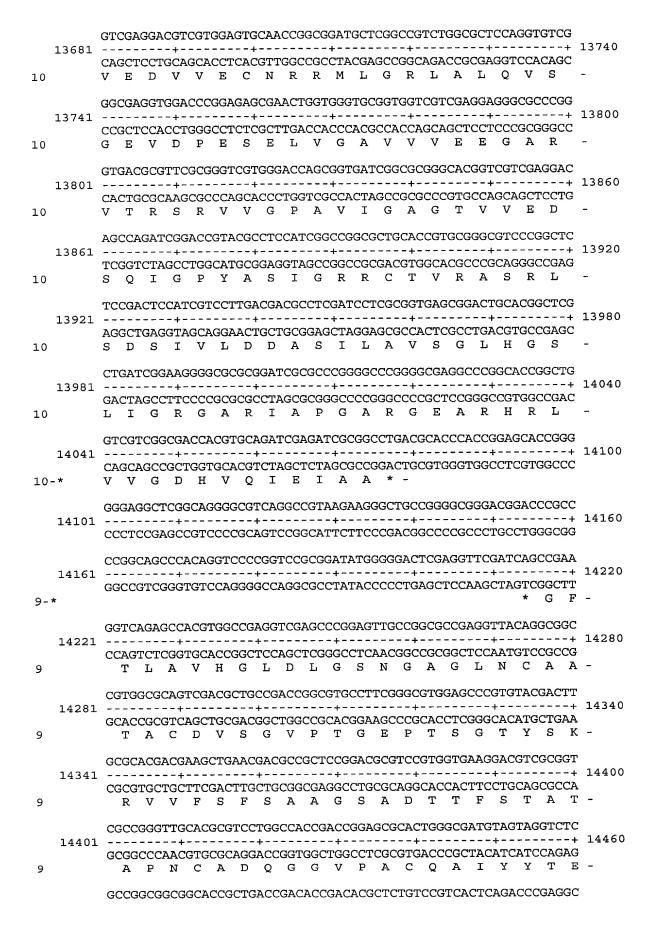
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11	11461	CGTCA Q L	.CGC	-+ CGC(BAG	ACG!	+ AGG'	TAC	rcc	GCC	+ GGG'	 TGGʻ	 TGG.	-+- AAA	.GGG	ccc	+ TGC	CCGC		+ ACGC	11520
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11		CGGGC P G	D	A	D	G	A	A	Т	Н	Т	M	G	Α	M	Ε	D	S	A	Е	-
11	11581	AGAAC TCTTG S F	GCG	- + - · ACG(GAC	CGG(+ 3CC	 CGG	 CAC	 AGC	+ CGC'	 TTG	 AGC	-+- AGC	CTC	CTC	CTGC	 3GG(CGGC	TAGT	11640
11	11641	GTTCG CAAGC L E	TGC	- + - · TTC(CTG	ACG	+ TCC	 AGC	 CTC.	 AGG	+ CGC	 CAC	 AAC	- + - CTC	TAC	AAG	GC(CCG(GACC	GTCT	11700
11	11701	AATAG	GAG CTC	TCC' -+-	rcc agg	GAA' CTT	TGG + ACC	TGC 	ATG TAC	TCG AGC	TAG + ATC	AAG TTC	CCG GGC	ACC	AGC	AA(CTC(GTA(+	GAAC	CGGC CGCCG	11760 -
11	11761	GCATO		- + - TCG	 GCC	 ATC	+ GCC	 !CGG	AGC	 TTG	+ AGG	AGC	TTG	- + -	GAC	CCA	GAG(+ CAG	CTG	+	11820
11		CGTCC GCAGC G D	TCC	-+- GTC	 CTC	 AAC	+ TCG	CTC	 GCG	 CGA	+ .CGG	TCG	TCA	- + -	CGA	CAT	CCA	+ CCG	CTC	CACGT	11880
11		GGGGG	CCTC	-+- CTC	 TTG	 TGG	+	AGC	TGC	TTC	+ GTG	 CGC	CGI	-+- 'AG(: 3GG(- CTG	GTC	+ CCG	GTA	CGGGC	11940
11		GCGCG CGCGG P A	 GGTC	-+- TTG:	 AAG	CAC	+ AAC	GAC	ATG	CTG	+ GTC	AGC	 BAAC	- + ·	CTG	 3GC	 CTC	+ GAG	CGG	+ CATCC	12000
11	12001	CCGG	GAGC	-+- CAG	TGC	GCC	 CAC	CCGG	 BAGC	CTC	+ CTCC		AGG	- + : 3CG(CTA	 GTC	 GCC	+ CGT	CCG	+ GCGCT	12060

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		GAGGAAGAGCGTGTAGCCCGCAGAGTATAAGGGTCCTTAGGAGACCGGGCGGG	12900
		GCCGCATCTTCGGTATTGCGAAGTCGTGGGCATTCTGCGAGAAGCATGAACCGCGTGGCC++ CGGCGTAGAAGCCATAACGCTTCAGCACCCGTAAGACGCTCTTCGTACTTGGCGCACCGG	12960
10-		CGGTCTACAGTGGCGTGGAATTTCAGTGATTGCGCTGAAGGGCGGCACACGATGAAGGCA+ GCCAGATGTCACCGCACCTTAAAGTCACTAACGCGACTTCCCGCCGTGTGCTACTTCCGT M K A	13020
10		CTTGTACTGTCGGGTGGTTCGGGGACCCGCCTGCGCCCGATCAGTTACGCCATGCCGAAG GAACATGACAGCCCACCAAGCCCCTGGGCGGACGCGGGCTAGTCAATGCGGTACGGCTTC L V L S G G S G T R L R P I S Y A M P K	13080
10	13081	CAGCTCGTTCCGATCGCCGGGAAGCCAGTCCTTGAATATGTTCTGGATAATATCCGGAAC++ GTCGAGCAAGGCTAGCGGCCCTTCGGTCAGGAACTTATACAAGACCTATTATAGGCCTTG Q L V P I A G K P V L E Y V L D N I R N	13140
10	13141	CTCGATATCAAAGAGGTCGCCATTGTCGTCGGTGACTGGGCTCAGGAAATTATTGAGGCA+ GAGCTATAGTTTCTCCAGCGGTAACAGCAGCCACTGACCCGAGTCCTTTAATAACTCCGT L D I K E V A I V V G D W A Q E I I E A	
10	13201	ATGGGTGACGGCAGCCGTTTCGGTCTGCGCCTCACCTACATACGCCAGGAGCAACCTCTG+ TACCCACTGCCGTCGGCAAAGCCAGACGCGGAGTGGATGTATGCGGTCCTCGTTGGAGAC M G D G S R F G L R L T Y I R Q E Q P L	13260 -
10	13261	GGCATCGCGCACTGCGTGAAACTGGCCCGAGACTTCCTCGACGAGGACGACTTCGTCCTC+ CCGTAGCGCGTGACGCACTTTGACCGGGCTCTGAAGGAGCTGCTCCTGCTGAAGCAGGAG G I A H C V K L A R D F L D E D D F V L	13320 -
10	13321	TACCTAGGCGACATCATGCTGGACGGAGACCTGTCCGCGCAGGCGGGGCACTTCCTCCAC ATGGATCCGCTGTAGTACGACCTGCCTCTGGACAGGCGCGTCCGCCCCGTGAAGGAGGTG Y L G D I M L D G D L S A Q A G H F L H	13380
10	13381	ACCCGCCCGCGCGCGGATCGTCGTGCGCCAGGTGCCCGACCCCCGGGCCTTCGGGGTG TGGGCGGGGCG	
10	13441	ATCGAGCTGGACGGCGAAGGGCGTGTGCTGCGCCTGGTCGAGAAACCCCGTGAACCGCGC TAGCTCGACCTGCCGCTTCCCGCACACGACGCGGACCAGCTCTTTGGGGCACTTGGCGCG I E L D G E G R V L R L V E K P R E P R	13500
10	13501	AGCGACCTCGCGGCGGTCGGCGTGTACTTCTTCACCGCGGACGTGCACCGCGCCGTCGAC TCGCTGGAGCGCCGCCAGCCGCACATGAAGAAGTGGCGCCTGCACGTGGCGCGCAGCTGCACCTG S D L A A V G V Y F F T A D V H R A V D	13560
10	13561	GCGATTAGCCCGAGCCGACGGGGGGGAGCTGGAAATCACCGACGCCATCCAGTGGCTGCTG+ CGCTAATCGGGCTCGGCTGCCCCGCTCGACCTTTAGTGGCTGCGGTAGGTCACCGACGAC A I S P S R R G E L E I T D A I Q W L L	13620
10		GAGCAGGGCCTGCCGGTCGAGGCCGGCCGCTACACGGACTACTGGAAGGACACCGGCCGG	



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		GGGACTGACGGAGAAGGCGGCGCGGCGAAGGCGACGGACTGTGCGGCGGCCGGC	
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		CCCTGACTGCCTCTTCCGCCCGCGCCGCTTCCGCTGCCTGACACGCCGCCGCCGGTCCGG	
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	14581	CTACCTACGCTGCCGGTGCTGCGCCTTCGTGCCCCCCTGTACACTGCATTGCT	11010
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		CATGCGTAGGCTCCGATTCGAGGAGGGGGGTTGATCACTCCATGAAAGGATCACCTCGCCG	1 4 7 0 0
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		GACGGCCGCCTGCATCTCCCTCTGTGCTCTCGTGGATTTCCGGCACGGCACTCCCGTCGA	14760
	14701	CTGCCGGCGGACGTAGAGGGAGACACGAGAGCACCTAAAGGCCGTGCCGTGAGGGCAGCT	14/60
8		PRGGADGET SEHIEPVASGD	-
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		CGGCCGCCGCAGAATGCGGCAGACCCCCCGCACCTCCTCCGGCCCCACCGCCGTACCGG	
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8		VAAREIRCVGRVEEICV	
		TGGGCAGCGACACCCGCTCGGTGAGCGCCTCCACCTTCGGGAGCGGATCGGGCGCGT	
	14821		14880
_		ACCCGTCGCTGTCGTGGGCGAGCCACTCGCGGAGGTGGAAGCCCTCGCCTAGCCCGCGCA	_
8		T P L S L V R E T L A E V K P L P D P A	-
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	14881		14940
		CCGCGCGCTCCAGCCTGGCCATCCCGAGCGTCGACACCGTCGGCCCCGACTTCATCCGCG	
8		H R A L D S R Y P E C S H C G P S F Y A	-
		GGGCCAGGACGTTGTGCCGTTGGAGCACCGCCTGGAGTTCGTCGCGGTGCAGCCCGGCGC	
	14941		15000
		CCCGGTCCTGCAACACGGCAACCTCGTGGCGGACCTCAAGCAGCGCCACGTCGGGCCGCG	
8		RALVNHRQLVAQLEDRHLGA	-
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	15001	CCTGCCGCAGGTGGAGCTACTGCTGCATGACCGTCAAGCTGTCGAGCAAGCCTAGGACGC	
8		R V A D V E I V V Y Q C N S L E N P D Q	-
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	12001	CCGCCTGGGCCTGCGGCCCGTCAGGCAGCTCCATGACGAGCATGTCTGCCATCAACGCGG	
8		PRVRVGPLGDLYQEYLRYNR	-
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	15121	CCAACTAGCGCCACTTCACTAGCCGCCTGAGGTCCCTCCACTCCGGGTACCGGCGCGACT	12100
8		R N I A T F H D A S E L S T L G M A A S	-
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	15181	AGAGCACGTAGGCGCGCTGGCAAGGCGAGGGCCACTAGAGTACGCGCCGCAACTCGGGGA	15240
8		I E H M R A V T G S G T I E H A A N L G	_
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	15721	I G	L A	K GTC	A AGC	A TCG	V GCC	Q ACA	I CGG	G GCC +	T TCG.	T AAC' 	A TCC: -+-	CGG.	ACC	AGC	A GGG	V SCCG	C TCAT	15780
0	15721	I G GCCTGA CGGACT	L A CCTGG	K EGTC CCAG	A AGC TCG	A TCG(+ AGC	V GCC CGG	Q ACA TGI	I .CGG 	G GCC + CGG	T TCG. AGC	T AAC' TTG.	A TCC(-+- AGG(CGG. GCC	ACC 	AGC	A GGG CCCC	V GCCG GGC	C TCAT + CAGTA	15780
8	15721	I G	L A CCTGG	K EGTC CCAG	A AGC TCG	A TCG(+ AGC	V GCC CGG	Q ACA TGI	I .CGG 	G GCC + CGG	T TCG. AGC	T AAC' TTG.	A TCC(-+- AGG(CGG. GCC	ACC 	AGC	A GGG CCCC	V GCCG GGC	C TCAT + CAGTA	15780
8	15721	GCCTGA CGGACT H R	L A CCTGC CGGACC V Q	K EGTC CCAG T	A AGC TCG L	A TCG(+ AGC) E	V GCC CGG A	Q ACA TGT V	I CGG GCC R	G GCC + CGG A	T TCG. AGC E	T AAC' TTG. F	A TCC -+- AGG E	CGG. GCC R	ACC TGG V	AGC TCC L	A GGGG CCCC	V GGGG G	C TCAT + CAGTA D	- 15780 -
8		GCCTGA CGGACT H R	L A CCTGC GGACC V Q ACCAC	K GGTC CCAG T CAGG	AGC TCG. L	A TCG(AGC) E TCC.	V GCC CGG A	Q ACA TGT V	I CGG CGCC R	G GCC + CGG A TCG +	T TCG. AGC E ATC	T AAC' TTG. F CGT	A TCC -+- AGG E TCC	CGG. GCC R ATC.	ACC TGG V AAA	AGG	A GGGG CCCC P STCC	V GGGC G GGGC	C TCAT CAGTA D EGAGC	15780 - 15840
8		GCCTGA CGGACT H R TGGTGA	L A CCTGC CGGACC V Q ACCAC	K GGTC CCAG T CAGG	A AGC TCG L CGC	A TCG AGC E TCC AGG	V GCC CGG A AGC TCG	Q ACA TGT V	I CGGG R CCCG	G GCC + CGG A TCG +	T TCG. AGC E ATC 	T AAC TTG. F CGT GCA	A TCC AGG E TCC -+- AGG	CGG. GCC R ATC. TAG	ACC TGG V AAA	AGC	A GGGG P STCG	V GCCG G G G G G G G G G G G G G	C STCAT CAGTA D SGAGC C+ CCTCG	15780 - 15840
8		GCCTGA CGGACT H R	L A CCTGC CGGACC V Q ACCAC	K GGTC CCAG T CAGG	A AGC TCG L CGC	A TCG AGC E TCC AGG	V GCC CGG A AGC TCG	Q ACA TGT V	I CGGG R CCCG	G GCC + CGG A TCG +	T TCG. AGC E ATC 	T AAC TTG. F CGT GCA	A TCC AGG E TCC -+- AGG	CGG. GCC R ATC. TAG	ACC TGG V AAA	AGC	A GGGG P STCG	V GCCG G G G G G G G G G G G G G	C STCAT CAGTA D SGAGC C+ CCTCG	15780 - 15840
		GCCTGA CGGACT H R TGGTGA ACCACT N T	L A CCTGG CGGACG V Q ACCAG TGGGT F W	K GGTC CCAG T CAGG CAGG	A AGC TCG L SCGC GCG R	TCGG + AGC E TCC + AGG	V GCC CGG A AGC TCG	Q ACA TGT V SGCC A	I CGG R R CCG CGGG	G GCC + CGG A TCG + AGC	TCG. AGC E ATC TAG	T AAC TTG. F CGT GCA R	A TCC AGG E TCC AGG E TCC E	CGG. GCC R ATC TAG M	ACC TGG V AAA TTT	AGG L L CGG R	A GGGG P GTCG F CAGG	V GCCG GGC GGCG GCGG R	C GTCAT CAGTA D GGAGC CCTCG S	15780 - 15840 -
	15781	GCCTGA CGGACT H R TGGTGA ACCACT N T	L A CCTGC CGGACC V Q ACCAC TGGTC F W	K GGTC CCAG T CAGG GTCC L GCGT	A AGC TCG L CGC R	TCG(+ AGC) TCC+ AGG' E	V GCC CGG A AGC TCG TCG	Q ACA TGT V EGCO A CAGO	I CGG R CCCG CCCG G	G GCC + CGG A TCG + AGC D	TCG AGC' E ATC TAG I CTG	T AAC TTG F CGT GCA R AAG	TCC -+- AGG E TCC -+- AGG TAG	CGG. GCC R ATC. TAG M	ACC TGG V AAA TTT L	CAGO L CGO R	A GGGG P GTCG CAGG	V GCCG G G GCGG CGCC R	C TCAT CAGTA D GGAGC CCTCG S TCCA	15780 - 15840 -
	15781	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT	L A CCTGC CGGACC V Q ACCAC TGGTC F W	K GGTC CCAG T CAGG GTCC L GCGT	A AGC TCG C CGC R CCCC	TCGGE E TCC. AGGE E AGGE E ACG	V GCC CGG A AGC TCG L	Q ACA TGT V CGCC A	I CGGG R CCCG CCCG G	G GCC + CGG A TCG + AGC D	T TCG. AGC E ATC TAG I CTG	T AAC TTG F CGT GCA R AAG	TCC -+- AGG E TCC -+- AGG E TAG	CGG. GCC R ATC TAG M	ACC TGG V AAA TTT L	CAGO L CGO R GGGT	A GGGG P STCG CAGG D	V GCCG G GCGG R GGAG	C GTCAT CAGTA D GGAGC CCTCG S GTCCA	15780 - 15840 -
8	15781	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA	L A CCTGC CGGACC V Q ACCAC F W CTCGGC	K GGTC CCAG T CAGG GTCC L GCGT	A AGC TCG CGC R CCCC	TCGGE E TCC+ AGGE E E E E E E E E E E E E E E E E E E	V GCC A AGC TCG L TGC ACG	Q PACA TGT V PGCC A PAGC	I CGGG R CCCG G G CGGT	G GCC + CGG A TCG + AGC TCG	T TCG. AGC E ATC TAG I CTG	T AAC TTG F CGT GCA R AAG TTC	TCC TCC E TCC AGG E TAG TAG	CGG. GCC ATC. TAG M GGC CCG	ACC TGG V AAA TTTT L	CAGO L CGO R CGCO	A GGGG P GTCG CAGG	V GCCG G GCGC R GGAG	C GTCAT CAGTA D GGAGC CCTCG S GTCCA CAGGT	15780 - 15840 -
	15781	GCCTGA GCCTGA H R TGGTGA ACCACT N T CCACGT GGTGCA G V	L A CCTGC CGGACC V Q ACCAC TGGTC F W TCGGC N P	K GGTC CAGG T CAGG L GTCC L GCGT CGCA	A AGC TCG. CGCC R CGCG CGCG CGCG	TCG+ AGC+ AGC+ AGG+ CC+	V GCC CGG A AGC TCG L TGC ACG H	Q ACA TGT V CGCC A CAGC TAGC L	I CGGGC R CCCGG GGGC GGGC F	G GCC + CCGG A TCG + AGC D TCG + AGC E	TCG. AGC' E ATC TAG I CTG GAC S	T AAC' F CGT GCA R AAG TTC F	TCC: -+- AGGG E TCC: -+- AGGG E TAG -+- ATC -+- ATC	CGG. GCC R ATC. TAG M GGC	ACC TGG V AAA TTTT L GGTG	L L CGCC R R GGGCT P	A GGGGG P GGTCG CAGG D FAGGG ATCG L	V GCCGGCGGCGCCR R GGGACCCCCCCCS	C GTCAT CAGTA D GGAGC CCTCG S GTCCA CAGGT D	15780 - 15840 - 15900
8	15781 15841	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA GACGCA	L A CCTGC GGACC V Q ACCAC TGGTC F W TCGGC N P	K GGTC CCAG T CAGG GTCC L GCGT CGCA	A AGC TCG. L GCGC R CCCC GGGG GGGG GGGGGG	A TCG+ AGC E TCC+ AGG E ACG+ CTCC	V GCC CGG A AGC TCG TCG L TGC H ACG	Q ACA TGT V GGCG A ACA GGCG L GGGG GGGG GGGGG GGGGGGGGGG	I CGGGC R CCCGG GGGC GGGCA P CTGCCA	G GCC + CGG A TCG + AGC D TCG AGC E CGCA	T TCG. AGC' E ATC TAG I CTG GAC S CGC	T AACC TTG. F CGT GCA R AAG TTTC F	TCCC -+- AGGG E TCCC -+- AGG E TAG -+- ATC Y	CGG. GCC R ATC. TAG M GGC P AGA	ACC TGG V AAA TTTT L GTG GCAC	L L L L CGCC R R GGGT P	A GGGGG P FFTCC CCCC P FFTCC CAGC D FFAGC D FFAGC L GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	V GCCGG G GCGGC R GGGAG CCCTC S	C GTCAT CAGTA D GGAGC GTCG S GTCCA CTCG TTCGG	15780 - 15840 - 15900
8	15781 15841	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA	L A CCTGC CGGACC V Q ACCAC F W CTCGGC AGCCC N P	K GGTC CCAG T CAGG L GGGT A GGGGT A R GGCGCA R	A AGCC TCG. L CCGCC R CCCCC GGGG G	A TCG+ AGC E TCC+ AGG E ACG+ TGC V CTC	V GCC CGG A AGC TCG L TGC ACG H	Q ACA TGT V CGCC A CAGCC L GCCC GCCC CGCC CGCCC CGCC CGCCC CGCCC CGCCC CGCCC CGCC CGCCC CGCCC CGCCC CGCCC CGCC CGCCC CGCC CGCCC	I CGGG R CCCGG GGGCCA GGGCCA P GGGCCA FTGCCA GGGCCA FTGCCA GGGCCA	G GCC + CGGG A TCG + AGC D TCG E CGCA	T TCG. AGC' E ATC TAG I CTG GAC S CGC	T AACC TTG. F CGT GCA R AAG TTC F	TCCC -+ AGGG E TCC+ AGGG E TAG -+- ATC Y CGA	CGG. GCC R ATC. TAG M GGC CCG P	ACC TGG V AAA TTTT L GGTG T GGGG	LACGOOR R	A GGGGG P CCCCC P TAGC D TAGC L GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	V GCCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	C GTCAT CAGTA D GGAGC S GTCCA STCCA CTCGGTCG CTGGGT CTGGGT	15780 - 15840 - 15900 -
8	15781 15841	GCCTGA GCCACTI H R TGGTGA ACCACTI N T CCACGTI GGTGCA G V GACGCA CTGCGCACTI CTGCCACTI CTGCC	L A CCTGC CGGACC V Q ACCAC F W CTCGGC N P ACCGGC	K GGTC CCAG T CAGG L GGGCA R GGCCA R CGGCA CGGGCA CGGGCA CGGGGCA CGGGGCA CGGGGCA CGGGGCA CGGGGCA CGGGGCA	A AGCC TCG. L CCGCC R CCCCC G G G G G G G G G G G G G	TCG+ AGC+ AGG+ AGG+ CC+ CGAGG+	V GCC GGG A AGC TCG L TGC H ATG	Q ACA TGT V CGCC A CAGCC L GCCCC CGGCC	I ACGG R ACCCG R ACCCG G G G C G G C C G G C C C C C C C	G GCC + CGGG A TCG CAGC CAGC CAGC CAGC CAGC CAG	T TCG AGC' E ATC TAG I CTG S CGC S	T AACC TTG. F CGT GCA R AAG TTC F CGGA	TCCC -+ AGGG E TCCC -+ AGGG E TAGG -+- ATC Y CGA -+-	CGG. GCC R ATC. TAG M GGC P AGA	ACC TGG V AAA TTTT L GGGG	ZAGGC L L L L L L L L L L L L L L L L L L	A GGGGG P CCCCC P TAGC L GGGGGGGGGGC CCCCCCCCCCCCCCCCCCCCCC	V GCCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	C GTCAT CAGTA D GGAGC S GTCCA STCCA CAGGT D TTGGG	15780 - 15840 - 15900 -
8 8	15781 15841	GCCTGA GCCACGT N T CCACGT GGTGCA G V GACGCA CTGCGCA	L A CCTGC CGGACC V Q ACCAC F W TCGGCC N P ACCGGCC P G	K GGTC CCAG T CAGG L GGGCA R GGCGCA R CGGCA R	A AGCC TCG. L CGCC R CCCC GGCG G GCGG G CGGCG R	TCGG+ AGCCE TCCCE+ AGGGE ACGGEV TGCC V CCTCCE+ CGAGGS	V GCC CGG A AGC TCG L TGC H ATG TTAC C	Q ACA TGT V CGCC A ACA CTCC L CGGC R	I ACGG R ACCCG G G G G G G G G G C G G G C G G G C G G G G C G	G GCC + CGGG A TCG CAGC CAGC CAGC CAGC CAGC CAG	T TCG AGC' E ATC TAGG I CTG GAC S CGC	T AAC' TTG. F CGT GCA R AAG TTC F CGA	TCCC -+ AGGG E TCCC -+ AGGG E TAGG -+- ATC Y CGA -+- CGCT E	CGG. GCC R ATC. TAG M GGC CCG P AGA	ACC TGG V AAA TTTT L GGTG T GGGG	ZAGGC	A GGGGG P FFTCC CAGC D FAGC L GGGGG A	V GCCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	C GTCAT CAGTA D GGAGC S GTCCA STCCA CAGGT D TTGGG CACCC G	15780 - 15840 - 15900 -
8 8 a b	15781 15841	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA CTGCGC D A T H	L A CCTGC CGGACC V Q ACCAC F W AGCCC N P ACCGGC P G R	K GGTC CCAG T CAGG L GGGCA R GGCGCA R GGCGCA R GGCGCA R A A A	A AGCC TCG. L CCGCC R CCCCC GCG G AGGG G AGGG R AGGG A A A A A	TCG+ AGC+ AGG+ AGG+ TGC+ TGC+ TGC+ TGC+	V GCC CGG A AGC TCGG L TGC H ACG H TTACG C TACG C T T T T T T T T T T T T T T T T T T	Q ACA TTGT V CGCCC A A CGCCC CGCC R A A A A A A A A A A A A A A A A A A	I ACGG R ACCCG G G G G G C G G C G G C C G G C	G GCC + CGGG A TCG CAGC CAGC CAGC CAGC CAGC CAG	T TCG AGC' E ATC TAGG I CTG GAC S CGC GCG A	T AAC' F CGT GCA R AAG TTCC F CGA CCGA CCGA CCGCT CCCCT CCCT CCCCT CCCT CCCCT CCCT CC	TCCC -+ AGGG E TCCC -+ AGGG E TAGG -+- ATCC Y CGA -+- CGCT E ' K	CGG. GCC R ATC. TAGG M GGC P AGA TCT E R	ACC TGG V AAA TTTT L CGTG T AGG T AGG T CCC A	ZAGG	A GGGGG P TAGG L GGGGG A GGGGG A GGGGG A	V GCCGG G GGCGGCC R GGGAC CCCTC S CCTGT V L 1	C GTCAT CAGTA D GGAGC S GTCCA CAGGT D TTGGG AACCC G L G	15780 - 15840 - 15900 - 15960
8 8	15781 15841 15901	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA CTGCGT D A T H	L A CCTGC C+ CGGACC V Q ACCAC F W TCGGCC N P ACCGGC P G R T G	K GGTC CCAG T CAGG L GGCGT R GCGCA R GCCGCA R	A AGC' TCG. L GCGC' R CCCC GGGG G GCGG GCGGC A A A	A TCG+ AGC+ AGC+ AGG+ CCTC+ CGAG CGAG CGAG CGAG	V GCC GGG A AGC TCGG L TGG H ATG C TACG H ATG	Q ACA TTGT V CGCC A A A CGCC A CGGC A A CGGCC A A A CGGCC A A A A	I CGGG R CCCGG GGGCA P CACCA A V F CCCGG CCGGT CCCA A CCCCA CCCA CCCA CC	G GCC + CGG A TCG TCG CAGC CGCA CGCA CGCA CGCA	T TCG. AGC' E ATC TAG CTG GAC S CGC GCG A ' P R	T AAC' F CGT GCA R AAG TTC F CGGA C TTC F CGGA R C C C C C C C C C C C C C C C C C C	TCC: -+- AGG: E TCC: -+- AGG: E TAGG: -+- ATC Y CCGA -+- CCGA CCGA R	CGG. GCC R ATC. TAG M GGC P AGA TCT TCT E R	ACC TGG V AAA TTTT L GGGG T AGGGC A C G	EAGG L LCGCCR R CCCCCC P CCCCCCCCCCCCCCCCCCCCCCC	A GGGGG P GTCC CCCCC P GTCC CAGC D GGGGG L GGGGG A GGGG GGGG GGGGG GGGGG GGGGG GGGGG	V GCCGG GGCG R GGGAG SCCTGT SCCTGT V CCCTGT	C TCAT TAGTA D GGAGC S TTCCA S TTGGG TTGGG TTGGG G L G W A	15780 - 15840 - 15900 - 15960
8 8 a b	15781 15841 15901	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA CTGCGT D A T H	L A CCTGC C+ CGGACC V Q ACCAC F W TCGGCC N P ACCGGC P G R T G	K GGTC CCAG T CAGG L GGCGT R GCGCA R GCCGCA R	A AGC' TCG. L GCGC' R CCCC GGGG G GCGG GCGGC A A A	A TCG+ AGC+ AGC+ AGG-E ACG+ CCTC+ CGAG-S+ LL+	V GCC GGG A AGC TCG L TGC H ATG C C C M M M M M M M M M M M M M M M M	Q ACA V CGCC A CGGC A CGGCC CGGC CGGC CCGGC R P	I CGGG R CCCGG GCCA GGCCA P CACCA A V F C C C C C C C C C C C C	G GCC + CGG A TCG TCG CAGC CGCA CGCA CGCA CGCA	T TCG. AGC' E ATC TAG CTG GAC S CGC GCG A TGC	T AAC' F CGT GCA R AAG F CCGA CCGA CCGA CCGA CCGA CCGA CCG	TCC: -+- AGG: E TCC: -+- AGG: E TAGG: -+- ATC Y CCGA -+- CCGCT E K R	CGG. GCC R ATC. TAG M GGC P AGA TCT E R R	ACC TGG V AAA TTTT L GGGG T CCCC A C I	EAGG L LCGCCR R CCCCR P CCCCCC GGCCCCR R CCCCCR P	A GGGGG P GTCC CCCCC P GGGGC L GGGGC A GGGGC A GGGGC GGGGC A GGGGC GGGC A GGGC A GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGCC GGCC	V GCCGG GGCC R GGGAC CCCTC S CCTGT V C C C	C TCAT TAGTA D GGAGC S TTCCA S TTGGG TTGGG TTGGG G L G W A	15780 - 15840 - 15900 - 15960
8 8 abc	15781 15841 15901	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA T H R T H R	L A CCTGC CGGACC V Q ACCAC F W AGCCC N P AGCCC P G CGGCC P G R T G	K GGTC CCAG T CAGG L GGGCA R GCGGCA R GCGGCA R	A AGC TCG. L CGCG R CCCG R CGCG R CGGG A A P	A TCG+ AGC+ AGG+ AGG+ TGC+ CGAGS L+	V GCC CGG A AGC TCG L TGC H ATG TACG H ATG TACG M M *	Q ACA V CGCC A CGGC A CGGCC CGGC CGGC CCGGC R P	I CGGG R CCCGG GCCA GGCCA P CACCA A V F C C C C C C C C C C C C	G GCC + CGG A TCG TCG CAGC CGCA CGCA CGCA CGCA	T TCG. AGC' E ATC TAG CTG GAC S CGC GCG A TGC	T AAC' F CGT GCA R AAG F CCGA CCGA CCGA CCGA CCGA CCGA CCG	TCC: -+- AGG: E TCC: -+- AGG: E TAGG: -+- ATC Y CCGA -+- CCGCT E K R	CGG. GCC R ATC. TAG M GGC P AGA TCT E R R	ACC TGG V AAA TTTT L GGGG T CCCC A C I	EAGG L LCGCCR R CCCCR P CCCCCC GGCCCCR R CCCCCR P	A GGGGG P GTCC CCCCC P GGGGC L GGGGC A GGGGC A GGGGC GGGGC A GGGGC GGGC A GGGC A GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGGC GGCC GGCC	V GCCGG GGCC R GGGAC CCCTC S CCTGT V C C C	C TCAT TAGTA D GGAGC S TCCA S TTGGG TTGGG G AACCC G L G W A TTGG W A TTGGG T ACCC G L G W A TTGG T T T T T T T T T T T T T T T T	15780 - 15840 - 15900 - 15960
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8 8 abc	15781 15841 15901 *	GCCTGA CGGACT H R TGGTGA ACCACT N T CCACGT GGTGCA G V GACGCA T H R T L R CCGGCCA	L A CCTGC+ CGGACC V Q ACCAC F W TCGGCC N P ACCGGCC P G R C G G C V P	K GGTC CCAG T CAGG GGCGT L GGCGA R GGCGG R GGCGG GGCGGG GGCGGGGGGGGGG	A AGC TCG. L GCGC R CCCC R CCCC R CCCC GGGG G GCCG R AGCC AGC AGC AGC AGC AGC AGC AGC AGC AG	A TCG+ AGC+ AGC+ AGG+ TGC+ TGC TGC TGC TGC TGC	V GCC GGG A AGC TCG TTGC H ATG TTAC C M M M M AGG	Q ACA TTGT V CGCCC A ACA CGGCC R A P A CCCGC	I CGGGCC R CCCGG GGGCC GGGCC A FT C C GGGCC T	G GCC + CGGG A TCG + AGC D TCG E AGCA + AGCA R ACGA	T TCG AGC' E ATC TAG I CTG GAC S CGC A ' R V	T AAC' F CGT F GCA R AAG TTC F CGA CGCT F GCT GCA CGCT GCCT CGCT CGCT CGCT CGCT C	TCCC -+- AGGG E TCCC -+- AGGG E TAGG -+- CGA CGCT E R -+- V	CGG GCC R ATC TAG M GGC CCG P AGA TCT E R F	ACCO TGG V AAAA TTTT L CGTG CCC A C GGGG A L AGGGG	EAGG L ACGCC R CCCCA P CCCCA G G G G G G CCCCA CCCCCA CCCCA CCCCCA CCCCCC	A GGGGG P GTCC CCCCC P GGGGC D GGGGC GGGGC GGGGC GGGGC GGGGC GGGGC GGGGC GGGC ACCCCC ACCCCC ACCCCC ACCCCCC ACCCCCC ACCCCCC	V GCCGG G GGCGG R GGGCG R CCTC S GGAC	C TCAT CAGTA D GGAGC S TCCA S TTGGG C AACCC G L G W A TCGAA	15780 - 15840 - 15900 - 15960 - 15960

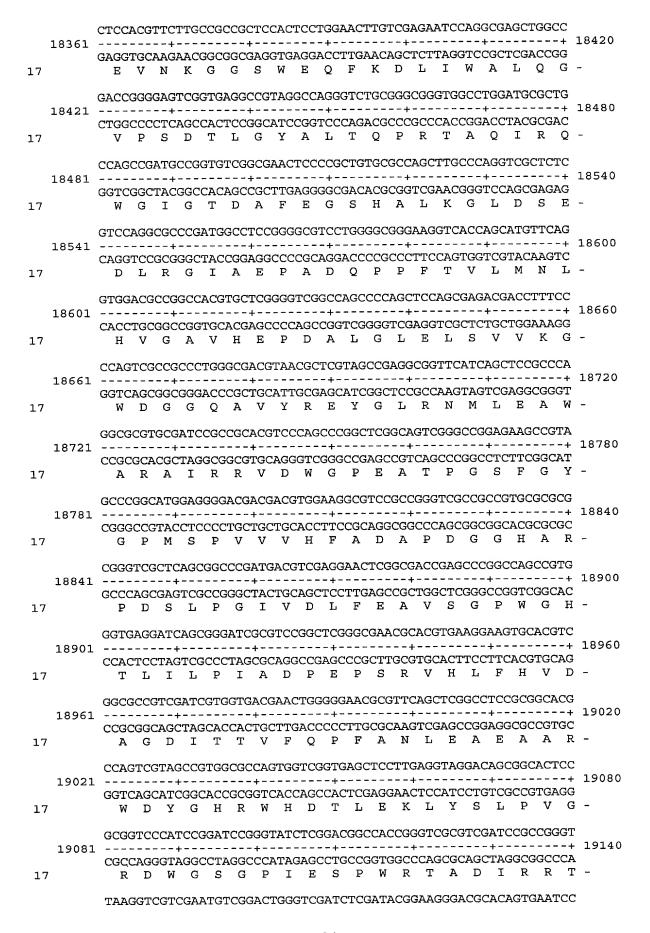
7		GGCCGGCAGCCGGTCGGCCTTCGGCCTGCTTGGCGTGGCTCTCGGGGTGGCTAAGCTT R G D A L R F G P V F R V S L G V S E F	_
		GGCGTCGGTGTACTGCTCGCGGGTGAAGAGGCTGGAGGTCAGGACCTCGGAGAACTCTCT+	16080
7		A D T Y Q E R T F L S S T L V E S F E R GAAGCCGGAGCGTCCGGAACCGGACCTCCAGACGTGACTTGTCGCCCTGGCG	
7	16081	CTTCGGCCTCCGCAGGCGCTGGGCCTTGGCCTGGAGGTCTGCACTGAACAGCGGGACCGC F G S A D A V R F R V E L R S K D G Q R	
	16141	CACGGAGTGCGTCATCCGCGTGATGACACGGCCCTCCTCCTGGTGCAGATGGCCGCCGAC	16200
7		$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	-
7	16201	TACGGGCAGCTCCTTCAAGAGCCCCTTTATGGTCCCAAGCCGCTGCTCCTGAGGGGGCCC H G D L F N E P F Y W P E A V L V G G P	
,	16261	GTTCAGGTGGTGGGCCATGGCCGACACCGCGGCCTTGAGCTCGGTGACGGACCCCATCTC	
7		CAAGTCCACCACCGGTACCGGCTGTGGCGCCGGAACTCGAGCCACTGCCTGGGGTAGAG N L H H A M A S V A A K L E T V S G M E	
7	16321	GCCGAGCGCGTTGCCCATGCAGGTGATCGCGTCGAAGGTGCGGCCCAGGTCGAACGAA	
	16381	CATGTCACCGGCGTGCAGCGGGACGCCGGGAAGCCGGCCG	16440
7		M D G A H L P V G P L R G A A Q E L M A GGGCGCGTACTCGAGGCCCTCCACATGGCCGAAGAGCGTGGCGAGCGTCTCCAGATGGGC	-
7	16441	CCCGCGCATGAGCTCCGGGAGGTGTACCGGCTTCTCGCACCGCTCGCAGAGGTCTACCCG PAYELGEVHGFLTAL	
7	16501	TCCGGTGCCGCAGGCGACGTCCAGGAGCGACACGGCGTCGGGGCGGGGGGGG	
	16561	CTCGGTGAGCCCGCGGGCCTCCAGGTCGAAGTCCTTGCCGCGGCTGCGGAACACGAGGTC+ GAGCCACTCGGGCGCCCGGAGGTCCAGCTTCAGGAACGGCGCCGACGCCTTGTGCTCCAG	
7	16621	E T L G R A E L D F D K G R S R F V L D GTAGAACTTCGCGTGCTCGGGGCCGTACTCCATCAGACGAGCTCCTTCGCAGACTGGGCG	
7- 6-	<	CATCTTGAAGCGCACGAGCCCCGGCATGAGGTAGTCTGCTCGAGGAAGCGTCTGACCCGC Y F K A H E P G Y E M - * V L E K A S Q A	
	16681	GAGATGATTCTGGGCTCCGGGATGGGAACGATGAACTTCCCTCCC	
6	16741	S I I R P E P I P V I F K G G A E L F R CGCTCCTTGCGGACGACCTCGTCGGTGTAGTTCCAGGCGAGGAGGAGGTAGTAGTCCGGC	

6		R E K R V V E D T Y N W A L L L Y Y D P	-
	16801	TCGGTGGCAGCGACCTCCTCCGGAGGAAGGACCGGGATGCGGTTCCCCGGCAGCAGTTTG	16860
6		AGCCACCGTCGCTGGAGGGGCCTCCTTCCTGGCCCTACGCCAAGGGGCCGTCGTCAAAC ETAAVEEPPLVPIRNGPLKK	-
	16861	CCGTGCTTGAGGCTGGTGTCGCCGCAGACGGTGATGTCCTGATCCGTCAGACCGCAG	16920
6		GGCACGAACTCCGACCACAGCGGCGTCTGCCACTACAGGACTAGGCAGTCTGGCGTC G H K L S T T D G C V T I D Q D T L G C	-
	16921	GCCATCAGCAACTGGGTCCCCTTGGACGGTGCTCCGTAGCCGGCCACGCGGTGGCCGTCC	16980
6		CGGTAGTCGTTGACCCAGGGGAACCTGCCACGAGGCATCGGCCGGTGCGCCACCGGCAGG A M L L Q T G K S P A G Y G A V R H G D	<u>-</u>
	16981	GCGGCCAGACCGCGAACGAGCGTACGGATCGCTTCGGTCACGCGCGCG	17040
6		CGCCGGTCTGGCGCTTGCTCGCATGCCTAGCGAAGCCAGTGCGCGCAGTGGGCGAGCCGC A A L G R V L T R I A E T V R T V R E A	-
	17041	AACGCCCGGTAGGGGGCATCCGTCAGCAGTCCGCGCTCCTCCAGGCCGAGCAGCAGCCCC	17100
6		TTGCGGGCCATCCCCGTAGGCAGTCGTCAGGCGCGAGGAGGAGGAGGTCCGCCGCGC F A R Y P A D T L L G R E E E L G L L A	-
	17101	GCGACCGAGGGCTCCGGGACCCGTGCGGCCGACTCGCGCGCG	17160
6		A V S P E P V R A A S E R A A V V A I S	-
	17161	CCGCCGTGCACGCGACCCGCTCCACGTCGATGATCCGCAGGCCGTGCGCGCGAAGAGG+ GGCGGCACGTGCCGCTGGCGAGGTGCAGCTACTAGGCGTCCGGCACGCGCGCTCTCCC	17220
6		G G H V A V R E V D I I R L G H A G F L	-
	17221	TGGCGCAGTGTGCAGGGAGAAGTACGACAGGTGCTCGTGGTAGATCGTGTCGAACTGG+ ACCGCGTCACACACGTCCCTCTTCATGCTGTCCACGAGCACCATCTAGCACAGCTTGACC	17280
6		H R L T H L S F Y S L H E H Y I T D F Q	-
	17281	TTCTCGTCGAGCAGGTTCAGCAGGTACGGCACCTCGATGACCAGGACGCCGTCGTCGTCGTCGTCGTCGTCGTCGTCGTCGTCGTC	17340
6		NEDLLNLLYPVEIVLVGDDD	-
	17341	AGCACTGCGTCGACGCCGTCCAGGATGCGGTGCACGTCGTCGATGTGCGCGAAGCACTGG+ TCGTGACGCAGCTGCGGCAGGTCCTACGCCACGTGCAGCAGCTACACGCGCTTCGTGACC	17400
6		LVADVGDLIRHVDDIHAFCQ	-
	17401	CGGCCGATGACGGCCTTGGCCCTGCCCTAAGGGCGATGCGGCCCGCGGGCTCCGGG+ GCCGGCTACTGCCGGAACCGGGACGGGACGAGTTCCCGCTACGCCGGGCGCCCGAGGCCC	17460
6		R G I V A K A R G Q E L A I R G A P E P CCGAAGAAGTCCGGGGTCCGTGGGGATCCCCCGGGCGTTGGCGATCTCGGCGAGGTTGGCC	-
	17461	GGCTTCTTCAGGCCCAGGCACCCCTAGGGGGCCCGCAACCGCTAGAGCCGCTCCAACCGG	
6		G F F D P D T P I G R A N A I E A L N A GCCGGGTCGACCCGGCACCCGCATGCCCGCCCGGAACATCGCGAGCTGGGTGCCG	-
	17521	GCCGGGTCGACCCCGGCCACCCGCATGCCCGCCGGAACATCGCGAGCTGGGTGCCG++++++++	

Literary Argusting 1 March

6		A	P	D	V	G	A	V	R	M	G	P	7	A	R	F	M	A	I	_	Q	Т	G	_
	17581							-+-				+				+			+	-			+	17640
6		TGCA V	ACC N	S S	G G	rcg. L	AGG E	TGC V	TGG V	TCC	AG(D	CGG	CC 3	TC(S	CGC A	TCC L	GA <i>I</i> S	ACG A	GG(CCG R	CCC R	'AG(T	CGG A	-
	17641	AGCC			+			-+-	- 		- - -	+				+				+			+	17700
6		TCGG L																					TGC V	-
	17701				+			-+-	-			+				+		-		+			+	17760
6		ATCA Y	.CG <i>P</i> H	AC: K	rtg: F	TCA L	CAG T	GGC G	CCC P	AGC D	CTG(V	CTC	ST <i>P</i> J	H H	GCT R	TCG L	CA(T	TE M	GT(CGG L	R R	FTG C	CTG S	-
	17761	CGGC			+			-+-		- 		+			-	+				+			+	17820
6		GCCG R	TG7 C	rgc: V	TAC' I	TGC V	AGC D	TCG L	CCC P	TTC F	CTG V	CAC	GG <i>I</i> O	ACG Q	CGG A	AGT E	AG(D	CCG A	CA(GCC D	GGC A	P	AGC D	-
	17821				+			-+-		- - -		+- •				+				+			+	17880
6		TGCT V																						-
	17881	CCGC	. .		+			-+-				+				+				+	- - -		+	17940
6-4	<	GGCG G																	CA	GAG	GA(CTG	CCG	
	17941	GAAT			+			-+-				+-				+	- -		-	+			+	18000
		GGCG																						
	18001	CCGC			+			-+-				+-	 GT	CGC	CTC	GCC	TG	CC	CAC	+ TT(GCG	 GGC	+ ATG	18060
17	- *												*	R	. 1	. F	•	ĸ	T	F.	Α	К	. V	_
	18061	GTC			+			-+-				+-				+				+			+	18120
17		CAG(GAG E	CTA I	CTC L	CTC	CGAC	CGC() I	CGA(CGA Q	GGT E	'AC M	CG(A	GCG A	CTI . F	CAC F	CGG I	GC(G	G G	GC(R	CAG D	CTI F	'GAG 'E	-
	18121	GGT			+			+-				+-	- -			-+	- 		- 	+	- 		+	18180
17		CCA(GT W	GGC R	GCA T	GT(CCCI	AGC(CGT(?	CCT. L	ACG I	GG G	AG E	CCG A	CTI	rgc'i	rgg S	CC'. R	rag I	P	3GC R	T T	CCG A	-
	18181	GTC			+			+				+-				-+				+-			+	18240
17		CAG(CAG D	GCC P	CTI F	GTO	GGC(ECT(GCG(V	GCT G	GCC V	P P	CG A	GCA T	GTO	CGC(L I	CGG P	TC W	CCG P	G G	CGG G	GGI V	CCA T	-
	18241	GCG	-		+			+				+-				-+-				+-			+	18300
17		CGC(CCG A	CTT F	'CAC	GGC(GGTA A N	ACG(GCG(CTC R	GGC A	CTG S	AG E	CAI Y	CA.	IGT: Y 1	rga L	CT Q	CGC A	GA S	CCT S	TGC	CCG A	-
	18301				+			+				- + -				-+-				+-			+	18360
17		GCA	GCG	CCA	GTC	CGG'	rca:	rct.	AGT.	AGT	GCF	4CC	CA	CTC	CGT	CGG	CCA	.GG	GCC	CT	CTA	CCC		

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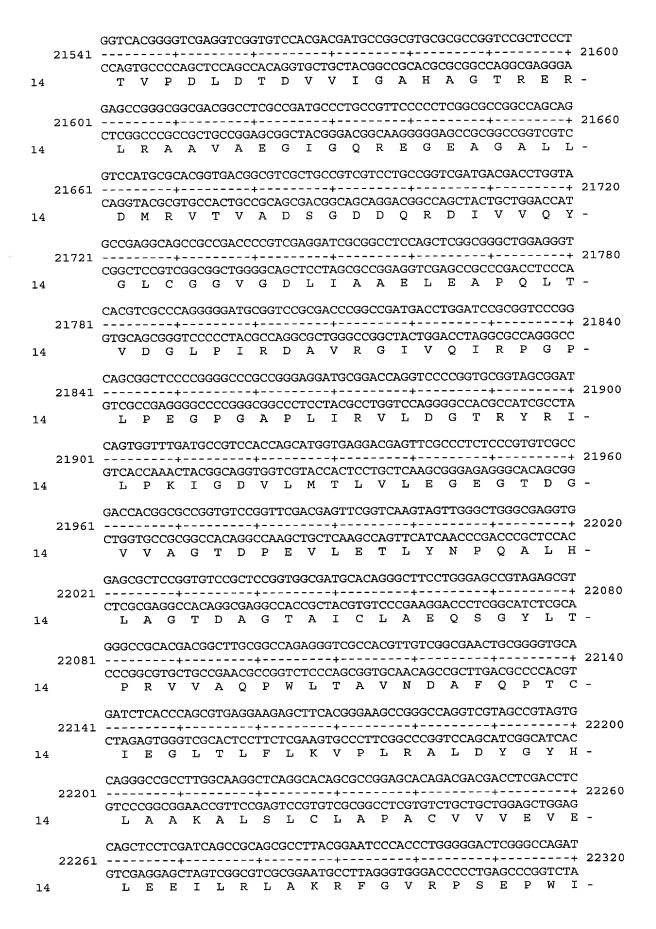


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	19201	ACCCTCGTGATTGTGGGAGCGGGGGGGGGGGGGGGGGGG	19260
16-		CGTGTCTCAGGCCGGTTCGGCCGGCGGCGGCGCGCGCCTTCCCGTGCGGAGAAGGACCGCAC GCACAGAGTCCGGCCAAGCCGGCCGGCGCGGGGAAGGGCACGCCTCTTCCTGGCGTG * A P E A P A A A G E R A S F S R V	
10		GGAGGACAGGAAGTTGCGGATCATCGGCATGCCGTGTTCGGTCCGGAAGCTCTCCGGATG	
16	19321	CCTCCTGTCCTTCAACGCCTAGTAGCCGTACGGCACAAGCCAGGCCTTCGAGAGGCCTAC S S L F N R I M P M G H E T R F S E P H	
16	19381	GAACTGGACGGACTCCACCGGCAGCGAACGGTGGCGCAGGCCCATCACGTACCCGTCGTC+ CTTGACCTGCCTGAGGTGGCCGTCGCTTGCCACCGCGTCCGGGTAGTGCATGGGCAGCAG F O V S E V P L S R H R L G M V Y G D D	
16	19441	CGTGGAGCGCCCGGTGACCTCGAGGGACGGCGGGACCGTGCCCTCCGGCACGATCAGTGA GCACCTCGCGGGCCACTGGAGCTCCCTGCCGCCCTGGCACGGAGGCCGTGCTAGTCACT T S R G T V E L S P P V T G E P V I L S	
16	19501	GTGGTAGCGGGTCGCGAAGAACCCCGCGGGGCAGCCCGGTGAACACTCCGCGCCCGTCGTG CACCATCGCCCAGCGCTTCTTGGGGCCCCCGTCGGGCCACTTGTGAGGCGCGGGCAGCAC H Y R T A F F G A P L G T F V G R G D H	19560
16	19561	CGTGATCCGGCTCGTCTTCCCGTGCATGAGATGCCGGGCGGG	19620
16	19621	GGCGCGGGCGACGGCCTGATGCCCCAGACAGACCCCGAGCAGCGGGACCCGGCCGAA CCGCGCCCGCTGCCGGACTACGGGGTCTGTCTGGGGCTCGTCGCCCTGGGCCGGCC	
	19681	GGCCTGGACGATCTCGACGTGCCCGGAGGTGTCGGGGTGGCCGGGGCCCCAGCAG+ CCGGACCTGCTAGAGCTGCACGGGCCTCCACAGCCCCACCGGCCCCGGGCCGGGTCGTC	19740
16		A Q V I E V H G S T D P H G P G P G L L GACCGCGTCCGGCCGCATCAGCCCCATCTCGTCCGGGGTCATGAGATGCGACCGCACCAT	
16		CTGGCGCAGGCCGGCGTAGTCGGGGTAGAGCAGGCCCCAGTACTCTACGCTGGCGTGGTA V A D P R M L G M E D P T M L H S R V M GACGGGCTCCGCGCGGCGGACATCAGATACTGGCGCAGGATGTCGACGAAGCTGTCGAA	
16		CTGCCCGAGGCGCCGCCTGTAGTCTATGACCGCGTCCTACAGCTGCTTCGACAGCTT V P E A G A S M L Y Q R L I D V F S D F	
16		CGCGTCGACCACCAGGACCCGCGGGGCCTCGGTGCCTGCGCCGGATCCGTCGGGAGACCA+ GCGCAGCTGGTGGTCCTGGGCGCCCCGGAGCCACGGACGCGCCTAGGCAGCCCTCTGGT A D V V L V R P A E T G A G S G D P S W	
	19921	CAAGCTCACAGCAACTCCTCTCCGGTGACCGCCCAGTGAGTG	

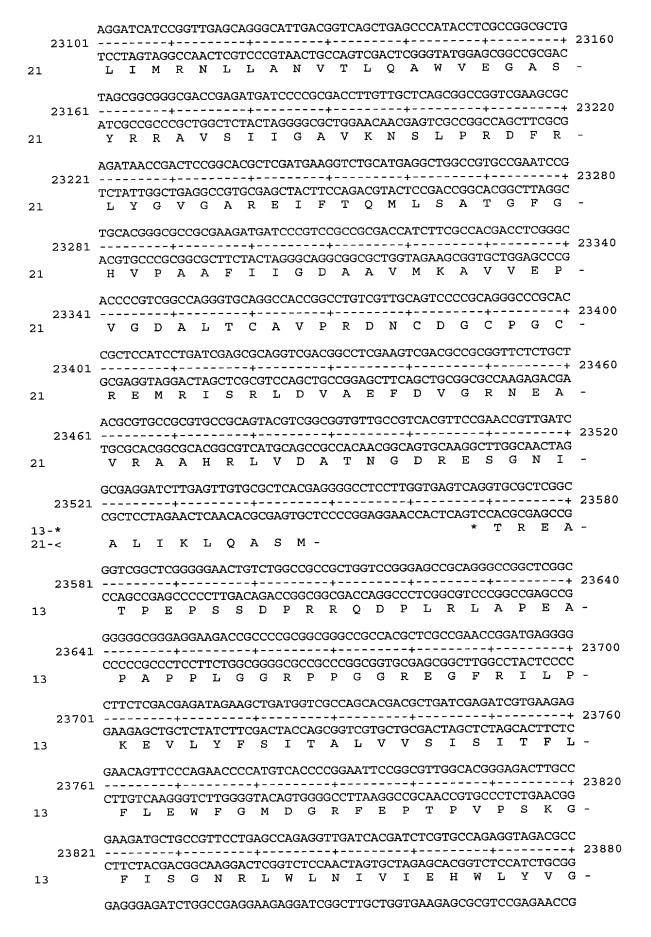
16- 15-		L S M - * L L E E G T V A W H T A S M K A L	-
15	19981	GTCTCGGTCCACTCCGCCCCCGGTTCGGAATCGGCGACGATTCCGGCCGAGGCCCGGGTG CAGAGCCAGGTGAGGCGGGGGCCAAGCCTTAGCCGCTGCTAAGGCCGGCTCCGGGCCCAC T E T W E A G P E S D A V I G A S A R T	20040
15	20041	CGGTAGACGCCCTCGTGGTGGAAAAGGGTCCGGATGCACAGCGCGAGGTTGGTGTACCCG+ GCCATCTGCGGGAGCACCACCTTTTCCCAGGCCTACGTGTCGCGCTCCAACCACATGGGC R Y V G E H H F L T R I C L A L N T Y G	20100
15	20101	CCCACGTCGAGGAGGCCGAGCGCCCCGGCGTACAGGCCGCGGCGGCTGCGTTCGACGGAC GGGTGCAGCTCCTCCGGCTCGCGGGGCCGCCATGTCCGGCGCCGCCGACGCAAGCTGCCTG G V D L L G L A G A Y L G R R S R E V S	20160
15	20161	TCGATGATCTCCATGGCGCGGATCTTCGGCGCGCGCCCGTCATGGTGCCGGCGGGAACAGG AGCTACTAGAGGTACCGCGCCTAGAAGCCGCGCGGGGAGTACCACGGCCGCCCTTGTCC E I I E M A R I K P A G T M T G A P F L	20220
15	20221	GCGGCGATGGTGTCGAAGGCATCGGTGTCCACCCGCGCCCGGCCGACGACCGTGGAGACC CGCCGCTACCACAGCTTCCGTAGCCACAGGTGGGCGCGGGCCGGCTGCTGGCACCTCTGG A A I T D F A D T D V R A R G V V T S V	20280
15	20281	AGGTGCAGCACGTGGGAGTAGCCCTCCACGTCCAGCTGGTCGGGTACGTCGAGCGTGTTC+++++ TCCACGTCGTGCACCCTCATCGGGAGGTGCAGGTCGACCAGCCCATGCAGCTCGCACAAG L H L V H S Y G E V D L Q D P V D L T N	20340
15	20341	GGCCGGCCATCCGTCCGATGTCGTTGCGGCAGAGGTCCACCAGCATGGTGTGCTCGGCG CCGGCCCGCTAGGCAGGCTACAGCAACGCCGTCTCCAGGTGGTCGTACCACACGAGCCGC PRAIRGIDNRCLDVLMTHEA	20400
15	20401	ATCTCCTTGGGATCCGACCTCAGCCGGACTCCCGCGGCGATGCCGCCGTCCGCCGGAC TAGAGGAACCCTAGGCTGGAGTCGGCCTGAGGGCGCCGCTACGGCGGCAGGCGCCTG I E K P D S R L R V G A A I G G D A G S	20460
15	20461	CGCGGCACCGTGCCCGCGATCGGCCGCATCGTGACCTCGCCGTCCTCGATGCGTACGAAC GCGCCGTGGCACGGGCGTAGCCGGCGTAGCACTGGAGCGGCGAGGAGCTACGCATGCTTG R P V T G A I P R M T V E G D E I R V F	20520
15		AGCTCGGGGCTGGCGCCGATCAGACGGTGCCCGTCGATGCCCGCCAGATACATGTACGGG TCGAGCCCCGACCGCGGCTAGTCTGCCACGGGCAGCTACGGGCGGTCTATGTACATGCCC L P S A G I L R H G D I G A L Y M Y P	20580
15		GAGGCGTTCCGCCCGCGCAGGCGCTGGTAGACGTCCGCGGGGTCGGCCGTCGAGCGGATG CTCCGCAAGGCGGCGTCCGCGACCATCTGCAGGCGCCCCAGCCGGCAGCTCGCCTAC SANRGRLRQYVDAPDATSRI	20640
15		GAGAGCTCGTGACCGATCTGCACCTGGTAGATGTCGCCGACGGCGATGTGCTTCAGACAC 1+ CTCTCGAGCACTGGCTAGACGTGGACCATCTACAGCGGCTGCCGCTACACGAAGTCTGTG S L E H G I Q V Q Y I D G V A I H K L C	- 20700 }
	20701	CGCTCGACGTCGTTCGCGAACACTTCGGGGGCGCTGTCGTCGGTGACCGCGGAGGCGGGGGGGG	20760

15		R	E	V	D	N	A	F	V	E	P	A	S	D	D	Т	V	A	S	Α	P	-
	20761	AAG		- - -	+			-+-			+		- -		+			+-			- +	20820
15		TTC F	GGC2 G	AGA(D	CGC(A	CTG(S	P	AGC D	CCG P	GTC W	CGG. A	ACG/ Q	AGG: E	rgc? V	AGC(CGCI A	L L	rcg(G G	CACI T	rgc V	-
	20821	GTC CAG		 CCG(+ : CGC:	rcc	GGC	- + - CCG	 GTC	 ATG	+ CCC	 CTG.	AGC	ACC	+ : FCG:	rcgi	CA/	- + - AGC(GTA	GCC	GC	
15		_	E																		G	-
	20881	GTG		-	+			-+-	-		+			-	+	·		-+-			+	20940
15		_	A	L	D	T	V	V	S	G	R	Н	L	V	M	R	V	D	P	L	G	-
	20941	GGC			+		-	-+-			+				+		-	-+-	-	-	+	21000
15		CCG P																		GGC'		-
	21001	AAC			+			-+-			+				+			-+-			+	21060
15			GGC G																	AGG(D		-
	21061	ATG	GCC	CGC.	AGC.	AGC	GGC	CAC	AAC	CCG	CCC	GCG	GTA	CGC	AGC	CGC	AGC	CCC -+-	TGG	GGG	CCG	21120
15	21001	TAC	:CGG	GCG	TCG	TCG	CCG	GTG	TTG	GGC	:GGG	CGC	CAT	GCG	TCG	GCG'	rcg	GGG	ACC	CCC	GGC G	
	21121	TCC	TCC	AGG	AGC	GCG	CCG	GCC	:CGC	TCC	AGG	AGC	AGG	ccc	CGC.	AGG	GCG	GGT	ACG	CCC	TCG	21180
15	21121	AGG	AGG	TCC	TCG	CGC	GGC	CGG	GCG	AGG	TCC	TCG	TCC	GGG	GCG	TCC	CGC	CCA	TGC	GGG. G	AGC	-
	21101	ACC	cgc	ACC	ACC	CGG	TCG	GTG	ACC	GAG	AGC	GAG	AGC	AGC	GCG	CCG.	AAG	CCG	ACG	AAC	TGG	21240
15	21101	TGC	GCG R	TGG	TGG	GCC	AGC	CAC	TGC	CTC	CTCG	CTC	TCG	TCG	CGC A	GGC	TTC	GGC	TGC	TTG	ACC Q	-
	21241	TGC	CTG	CGG	TCG	CGG	GCC	GGG	CCC	GCC	CGCC	GAC	TCC	'AGG	AGG	TAG	ACC	TCG	TCG	GGG	CCG	21300
15		ACC	GAC	:GCC	'AGC	:GCC	:CGG	CCC	CGGC	CCGC	3CGC	CTC	AGG	TCC	TCC	ATC	TGG	AGC	AGC	CCC	GGC G	
	21301				+			-+-							+			-+-			CGT	21360
15		TT(CACO H	AGC E	CGG A	TCC L	CGC A	CGCC R	CAT(Y	CCG(A	CCC(P	TC(L	CCGC A	GGG G	CAG T	AGG E	AAG K	TGT V	AGC D	TCC L	:GCA R	-
	21361			- -	+			-+-				+			+			-+-				21420
15	-<		ACAC T																CCCF	AGGA	\GGG	
		GGZ	ATC	ACGI	rggī	rga7	rggo	CGT	AGC	GGT	GTG(CCAC	CCTC	ACC	GGC	CGGT	'CAC	CAC	CCGC	CCCG	GTC	21480
14	21421 -*	CC'	rag:	rgc <i>i</i>	ACCA	CTA	ACCO	GCA.	rcg:	CCA	CAC	GGT(3GA(CTG	CCC	CCA	GTC	CGT	GGC	3GGC	CAG R D	
	21481	GG	GGC(CGGI	AGCC	GTT	rgto	CGA(CGA	CGC(GCG(CGG(CCT	rcc <i>i</i>	AGC7	GAC	GAZ	AGG2	AGC	CGGT	GTG	21540
14		CC	CCG	3CC1	rcgo	CCAZ	ACAC	3CT(GCT	GCG	CGC	GCC	GGA/	AGG:	rcg <i>i</i>	ACTO	CT	rcc'	rcg	GCCF	ACAC T H	

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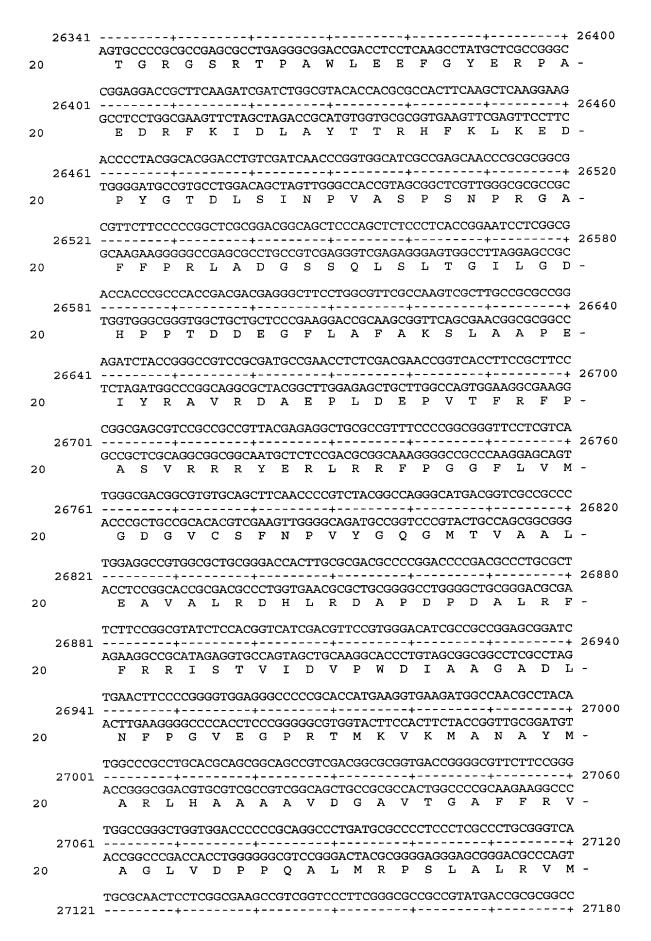
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14		GAACTGCACTGTCCGGGGGTCGAGGCGACGGTGGCGCCACTTGTGCAGGGGCTTGCGCAT K V H C A G L E A A V A T F V D G F A Y -	2300
	22201	CAGCTCCGACGGCCCCATCAGGCCCACGACGGCATCCGCCCCCGAACCTCGCTTCCAG	2440
14		GTCGAGGCTGCCGGGGTAGTCCGGGTGCTGCCCGTAGGCGGGGGGCTTGGAGCGAAGGTC L E S P G M L G V V P M R G G F R A E L -	
		CATGCGGCGCCAGGACTCCCGGACGGCGATGTTGCTGGTCGCGATGTCCTTCTCGCCGCG	
14		GTACGCCGCGGTCCTGAGGGCCTGCCGCTACAACGACCAGCGCTACAGGAAGAGCGGCGC M R R W S E R V A I N S T A I D K E G R -	2500
		TGGGCACGGGGTGGCCCCCGGTGGTCCCGGTGGTCTCGTAGTAGATGCGTGCTTCGTG	
	22501	ACCCGTGCCCCACCGGCGGGCCACCAGGGCCACCAGAGCATCATCTACGCACGAAGCAC	2560
14		P C P T A A G T T G T T E Y Y I R A E H -	
	22561	CAGCGGGCCCGACAGGACGTCGTGCATCTCCCGCCGCAGGTCGTCCTTGGTGGTGAAGGG	2620
1.4	22301	GTCGCCCGGGCTGTCCTGCAGCACGTAGAGGGCGGCGTCCAGCAGGAACCACCACTTCCC L P G S L V D H M E R R L D D K T T F P -	
14		CAGGTCCGCCAGGTTCGCGGGGGTGACGGCCTCGACGTCCACGCCTGCCAGATGGCGGCG	
	22621	+	2680
14		GTCCAGGCGGTCCAAGCGCCCCACTGCCGGAGCTGCAGGTGCGGACGGTCTACCGCCGC L D A L N A P T V A E V D V G A L H R R -	
		GTAGAACGGCGAGCGGCGGTGACGTGGCGCAGTACGGCCGTCAGCCGTTCGCCCTCCCA	2740
	22681	CATCTTGCCGCTCGCCGCCCACTGCACCGCGTCATGCCGGCAGTCGGCAAGCGGGAGGGT	
14		Y F P S R R T V H R L V A T L R E G E W -	
	22741	GCGCTCGCGGTCGGCGGCGGTGAGTTCGCCGCGGTAGAACGCGTCGCTCACCTGCCCGTA	2800
14		$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	
		GGCGGACCAGAACTCGCTGTCCGCGTCGGGGTCCAGCGGCCCGGTCCCGCCGGGACCGGG	
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14		A S W F E S D A D P D L P G T G G P G P -	•
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		GGCGGCCGGCAGAGTGCCGACACGGACCTCAAGCAACTCGCGCTCCGGCTGGGCGAGC	
14 21		RRGDRM- *PQAQLENLALGVRE-	-
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		AACTGGAGCAACCTCCGGTCGTGCAGGCTTGCCGGCCACTCGGCTGCCACAAGCAGCTCG	
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	22981	AGTTCGATCATGTCCGTCATCCTCTCGACCAGGCGCGAGACGTTGGTGAGGCCCTCCTCG	23040
21		TCAAGCTAGTACAGGCAGTAGGAGGAGCTGGTCCGCGCTCTGCAACCACTCCGGGAGGAGC L E I M D T M R E V L R S V N T L G E E -	-
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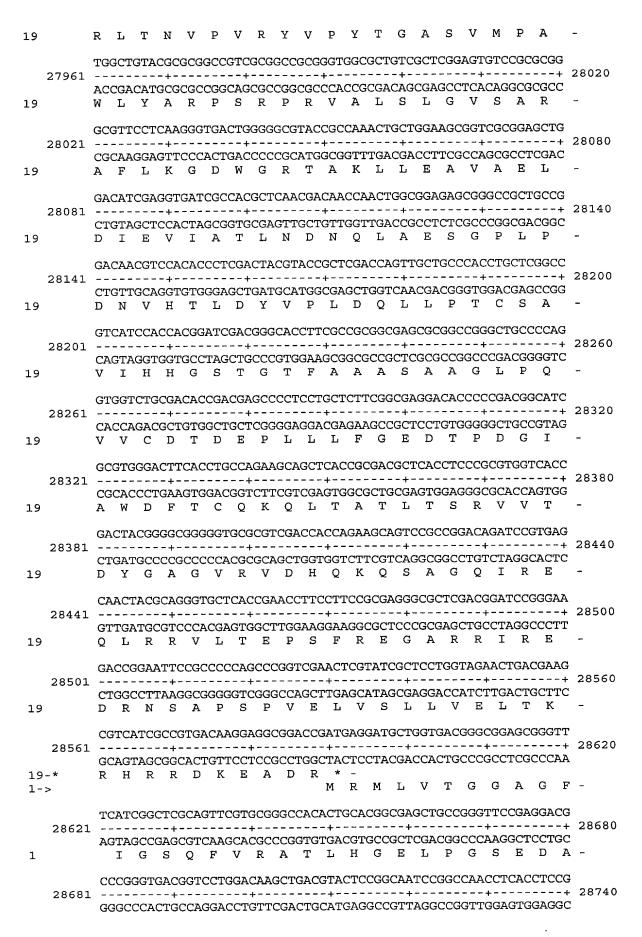
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13		S E A G P V T M P A L L L T F S T L I -	-
	24001	GAAGTGGTCGACGAGCTCCTGGGCCAGGGCCGCGTTGTCGCCCATGCCCGGGATGCCGAT	24060
13		CTTCACCAGCTGCTCGAGGACCCGGTCCCGGCGCAACAGCGGGTACGGGCCCTACGGCTA F H D V L E Q A L A A N D G M G P I G I	-
	24061	GGGCTTGGTGGCGTAGAGGAGGTACAGCGGGATGAGCGGGACCCAGCAGATCAGCGGGCG	24120
13	24001	CCCGAACCACCGCATCTCCATGTCGCCCTACTCGCCCTGGGTCGTCTAGTCGCCCGC P K T A Y L L Y L P I L P V W C I L P R	
	24121	CCGGATCACGAAACGGTAGAAGCCCGGGGTCCCTGGCGTCGCCTCGGCGTACGCGGAGTA	24180
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13	24301	GTCGCGCCGGTCGTCCTCCGCCGCACGGAAGACGGGCCCTCACGACGCGGGGC L A A L P L L R R A K Q G P T S G G R	
	24361	CGCGAGCCGGTGGCCGATCCAGGCGATCAGCGGCAGGCGAGGTAGAACGTGAACTCGGC	24420
13	24501	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	
	24421	GGGGACCGTCCAGGTGGGCTCGATGCCGTGCATCGGCTGGCCCTCGGGCAGATAGAAGTG	24480
13		CCCCTGGCAGGTCCACCCGAGCTACGGCACCGAGCCGGAGCCCGTCTATCTTCAC P V T W T P E I G H M P Q G E P L Y F H	-
	24481	CATGAGCACGGGCCGCAGGACGTCGCTGACGCTGTCGATCTCGAACCAGTTGTAGCC	24540
13		GTACTCGTCGTGCCGGCGTCCTGCAGCGACTGCGACAGCTAGAGCTTGGTCAACATCGG M L L V P R L V D S V S D I E F W N Y G	
	24541	GGGGATTGCGAAGACGAGCAACAGGTAGTAGGCGGGCAGGATGCGCAGGGCCCGGCGTTT	24600
13		CCCCTAACGCTTCTGCTCGTTGTCCATCATCCGCCCGTCCTACGCGTCCCGGGCCGCAAA P I A F V L L L Y Y A P L I R L A R R K	
	24523	GAGGAACCGTCCGGTGGCGGGCCGCTTCGTCCCACTGATGGTGACGCGGGCGTAGGGCTT	24660
13		CTCCTTGGCAGGCCACCGCCGGCGAAGCAGGGTGACTACCACTGCGCCCGCATCCCGAA L F R G T A P R K T G S I T V R A Y P K	
	24661	GTACAGCATCATTCCGGACAGAGCGAAGAAGGGGGAAGGCATACCCCCAGACCGTCCGCG	24720

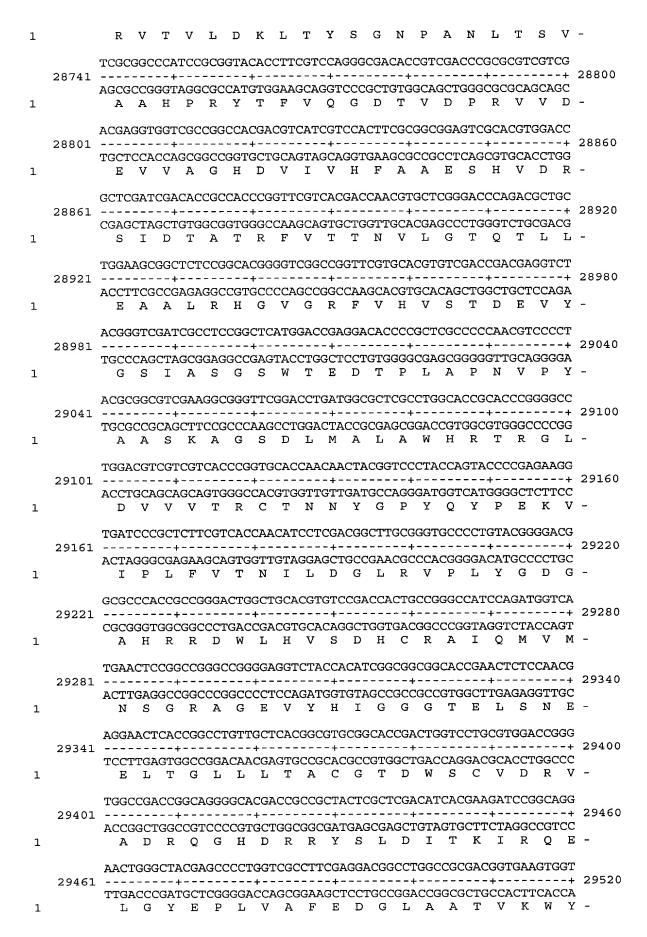
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	24721	AGGACGCCCAGAACGGTTTGCCCGGCTCACCGACGAAGCTGCCCACTCCGGCCTGGAAG+++ TCCTGCGGGGTCTTGCCAAACGGGCCGAGTGGCTGCTTCGACGGGTGAGGCCGGACCTTC	24780
	24781	GCGACGTGGTAGACGACCACACCCAGCGCGAGGACACCTCGCAGTCCCTCGAACTTCGGT+	24840
	24841	ATTCGCTTGCTTTTTGCGCCACCTGCGTCGCGAAGGACGTCCCCATGGAACAGTCCCCT+++ TAAGCGAACGAAAAACGCGGTGGACGCAGCGCTTCCTGCAGGGGGTACCTTGTCAGGGGA	24900
	24901	TTCCCTTGGCACTTGCTCGTTGACTTCCCGAAATAGTCGGGTCTGCGGAGTGTGAGCCGC+++ AAGGGAACCGTGAACGAGCAACTGAAGGGCTTTATCAGCCCAGACGCCTCACACTCGGCG	24960
	24961	ATCTCCAATCGTGCTGTTCCGGTGCTCAGGACGACTTGTTTCGGCCTGAGTGGGAAGGCA+ TAGAGGTTAGCACGACAAGGCCACGAGTCCTGCTGAACAAAGCCGGACTCACCCTTCCGT * S S K N R G S H S P	25020
12-		* S S K N R G S H S P GCCACCCCGCCGCCCGCCTCGGCCAGACCGGGGGCCGAGGAGTCCCGTTCCGAGAGGA	-
12	25021	CGGTGGGGGCGGGGGGGGGGGGGGGGCGGTCTGGCCCCGGCTCCTC	25080
12	25081	TCGGAGTGATCTCCGGCGGCCAGGCGATGCCCACCTCCGGATCCAGCGGATTCAAGCCAT AGCCTCACTAGAGGCCGCCGGTCCGCTACGGGTGGAGGCCTAGGTCGCCTAAGTTCGGTA I P T I E P P W A I G V E P D L P N L G	25140 -
12	25141	GTTCGAGCCGGGGGTCGTAGGCCGCCGAGCACAGGTAGACGATCACCGCCTCGTCGCTCA CAAGCTCGGCCCCCAGCATCCGGCGGCTCGTGTCCATCTGCTAGTGGCGGAGCAGCGAGT H E L R P D Y A A S C L Y V I V A E D S	25200 -
12	25201	GCGTGAGGAATCCGAAGCCCAGCCCCGCGGAGACGTACAGCGCCCGTCCGT	25260 -
12		CGAGCTCCACGGTCCGCCAGCCGCCGAAGGTGGGCGACCCCACCCGGATGTCGACCACGG+ GCTCGAGGTGCCAGGCGGTCGGCGGCTTCCACCCGCTGGGGTGGGCCTACAGCTGGTGCC G L E V T R W G G F T P S G V R I D V V	
12	25321	CGCCGAACACGCTGCCGCGCAGGCAGCTGAAGTACTTGGCCTGGCCGGGTACGCCCCCGG GCGGCTTGTGCGACGGCGCGTCCGTCGACTTCATGAACCGGACCGGCCCATGCGGGGGCC A G F V S G R L C S F Y K A Q G P V G G	25380
12		CGAAGTGGATGCCCCGCAGCACCCCGTGGGAGGAGATCGCGCAGTTCGCCTGCCGCAGGT GCTTCACCTACGGGGCGTCGTGGGGCACCCTCCTCTAGCGCGTCAAGCGGACGGCGTCCA A F H I G R L V G H S S I A C N A Q R L	25440
12		CGAAGGAGTGGCCTACGGTGCGGCGGAAGGGCTCGCCCTGGAACCACTCGCGAAACGAGC GCTTCCTCACCGGATGCCACGCCGCCTTCCCGAGCGGGACCTTGGTGAGCGCTTTGCTCG D F S H G V T R R F P E G Q F W E R F S	
	25501	CCCGTTCGTCACGGAAGACCTGCTTCTCCTCCGTCCACGCTCCCGAGATCCCGATCGGCT	25560

12		GGGCAAGCAGTGCCTTCTGGACGAAGAGGAGGCAGGTGCGAGGGCTCTAGGGCTAGCCGA G R E D R F V Q K E E T W A G S I G I P -	
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	25621	CGCCGGGCCCGTGGGAACGCCGCAGTCTAGATGCGGCGGCACCGGGGGCAGGGGGGTGCG	25680
	25681	GACGACGTCCGCCCCACCTCAGCACACCGGGAGATGCAGGTCGGTGACGGCGACGTGAC+	25740
	25741	GATGCAACGGTCCGAGGCCCGGTTGCCCGGACGACGCCCACAGAGCCATCGGAGCAACG+	25800
		GAGGCGGACCGCAGATGACCAAGCACGCCCGTGACCGCGCGGTAGTCCTCGGCGCAGGGA+++++++	
20-		TGGCGGGGCTCCTGTCCGAGACGTACAAGGAAGTGCTGGTGATCG ACCGCCCCGACGAGGCGCGCGCAGGACAGGCTCTGCATGTTCCTTCACGACCACTAGC A G L L A A R V L S E T Y K E V L V I D	25920
20	25921	ACCGGGACCGGTTGGGCGCACGGAGCACGCCCCGCGTGTCCCGCACGCA	
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20	26041	AACTCACCTCGGCCGGAATCCCCGCCGGGGACCTGCGGGAACCTGCGGTGGTACTTCA TTGAGTGGAGCCGGCCTTAGGGGCGCCCCTGTAGCGGCCCCTTGGACGCCACCATGAAGT L T S A G I P A G D I A G N L R W Y F N	
20		ACGGCCGCCGGCTCCAGCCCTTCGACACCGGGCTGATCAGCGTCTCGGCGACGAGGCCCG TGCCGGCGGCGAGGTCGGGAAGCTGTGGCCCGACTAGTCGCAGAGCCGCTGCTCCGGGC GRRLQPFDTGLISVSATRPE	
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	26221	GGTGCGTGATCCGGGGCCTGACCGCCTCGGCCGACCGCAGCCGCGTCACCGGTGTCGAGG CCACGCACTAGGCCCCGGACTGGCGGAGCCGGCTCGGCGCAGTGGCCACAGCTCC C V I R G L T A S A D R S R V T G V E V	26280
20	26281	TGGTCGACGAGTCGGGTACGGACACCCCGACGCGCCTGGAGGCCGACCTCGTCGTCGACG ACCAGCTGCTCAGCCCATGCCTGTGGGGCTGCGCGGACCTCCGGCTGGAGCAGCAGCTGC V D E S G T D T P T R L E A D L V V D V	26340
∠ ∪		TCACGGGGCGCGGCTCGCGGACTCCCGCCTGGCTGGAGGAGTTCGGATACGAGCGGCCCG	



20-	. *	ACGCGTTGAGGAGCCGCTTCGGCAGCCAGGGAAGCCCGCGGCGCATACTGGCGCGCGGCAGCCAGC	
19	27181	CGTCCGGGGCGGCTGCCGGGGCCAGGAGCCGACATGCGGGTGATGATCACGGTGTTCCCG+ GCAGGCCCCGCCGACGGCCCCGGTCCTCGGCTGTACGCCCACTACTAGTGCCACAAGGGC M R V M I T V F P	27240
19	27241	GCGCGGCCCCCTGCTGCCGCTGGTGCCCTATGCCTGGGCCCTGCAGAGCGCGGGCCAC CGCGCCCGCGTGAAGGACGGCGACCACGGGATACGGACCCGGGACGTCTCGCGCCCGGTG A R A H F L P L V P Y A W A L Q S A G H	27300 -
19	27301	GAGGTATGTGTGGGGGCCCCCGGGCTATCCCACCGGGGTGGCCGACCCCGACTTCCAC + CTCCATACACAGCACCGCGGGGGCCCGATAGGGTGGCCCACCGGCTGGGGCTGAAGGTG E V C V V A P P G Y P T G V A D P D F H	27360 -
19	27361	GAGGCCGTCACCGCGGCCGGCCTGAAGTCGGTGACCTGCGGGCAGCCGCAGCCGCTGGCG CTCCGGCAGTGGCGCCGGCCGGACTTCAGCCACTGGACGCCCGTCGGCGTCGGCGACCGC E A V T A A G L K S V T C G Q P Q P L A	27420 -
19	27421	GTCCACGACCGCGACGACCCCGGCTACGCGGCGATGCTGCCGACCGCGGCGAGTCGGAG CAGGTGCTGGCGCTGCTGGGGCCGATGCGCCGCTACGACGGCTGGCGCCGCCTCAGCCTC V H D R D D P G Y A A M L P T A A E S E	27480
19	27481	CGCTACGTGGCGGCCCTCGGGATCAGCGAGAAGGAGCGCCCCACCTGGGACGTCTTCTAC+ GCGATGCACCGCCGGGAGCCCTAGTCGCTCTTCCTCGCGGGGTGGACCCTGCAGAAGATG R Y V A A L G I S E K E R P T W D V F Y	27540 -
19	27541	CACTTCACCTTGCTGGCGATCCGCGACTACCATCCGCCGCGGCCGGC	27600 -
19		GTGAAGTGGAACGACCGCTAGGCGCTGATGGTAGGCGCGCCGGCGCCGTCCTGCACCTG	-
	27601 27661	GTGAAGTGGAACGACCGCTAGGCGCTGATGGTAGGCGGCGCGCGC	-
19	27601 27661 27721	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	- 27660 - 27720 - 27780
19	27601 27661 27721 27781	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	- 27660 - 27720 - 27780 - 27840
19	27601 27661 27721 27781	GTGAAGTGGAACGACCGCTAGGCGCTGGTATGGTAGGCGGCGCCGGCGCGCGC	- 27660 - 27720 - 27780 - 27840

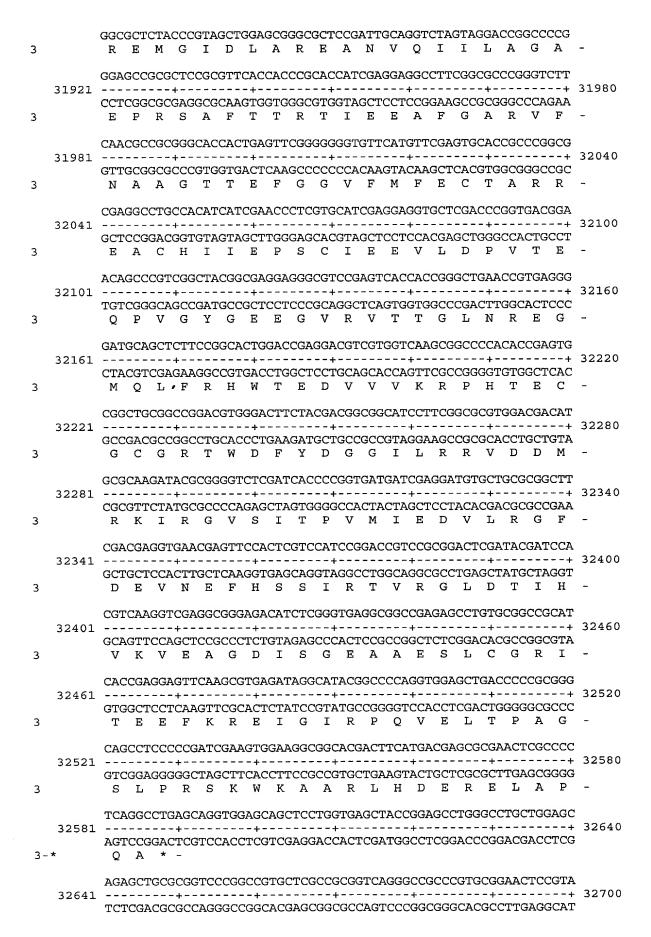




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1		HENRSWWQPLKEAAGLLDAV-	•
	29581	TCGGCTGACGGCAGCCACCGCTAGGAACACCCCAGGAAAGGAGCCACCTCCGTGACAGCA	9640
		$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	_
2-> 1-*		G * -	
	00647	GTCAAGGAGCCGACGTCCCGCGCAGGACGGCGGGAGTGGATCGCTCTCGTCGTCCTCTCC	29700
	29641	CAGTTCCTCGGCTGCAGGGCGCGTCCTGCCGCCCTCACCTAGCGAGAGCAGCAGGAGAGG	25700
2		V K E P T S R A G R R E W I A L V V L S -	-
	29701	TTGCCCACGATGCTGTTGATGCTGGACATCAACGTCCTCATGCTGGCCTTGCCGCAGTTG	29760
•	25701	AACGGGTGCTACGACAACTACGACCTGTAGTTGCAGGAGTACGACCGGAACGGCGTCAAC L P T M L L M L D I N V L M L A L P Q L	_
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	29761	AGCGAGGATCTCGGCGCGAGCAGCACCGCAACAGCTGTGGATCACCGACATCTACGGATTC	29820
2		$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	-
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2		A I A G F L V T M G T L G D R I G R R R	-
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	29881	GAGGACGAGCCCCCGCGCCGCAGAAGCGCCAGCACAGGCAGCA	
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2		TCGCGCCGCTACGAGCAGCAGCCGCGCGCCCCGGTGCCACTAC S A A M L V V S R A V L G V A G A T V M	_
2			
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2		$\begin{array}{cccccccccccccccccccccccccccccccccccc$	-
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2		I A M W A S A M M A G V A L G P A V G G	-
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	30121	GACCAGGAGCGCGCAAGACCACCCCTAGCCACAAGGAGTAGCGGCAAGGCCACTACGAC	
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2		L D L L S A G L S L A T V L P V I Y G L	_

	30301																				GIG	30360
2		TTC	CTC	CGAC	CCGC	GCC	TGG T	CCC	ACC	CTC	GGG	CGAC	CGG	CGC	CCC	CGC	CAC	CAC	GAC	CCC	CAC	-
	30361				-+			-+-							- +			+			CCTC + EGAG	30420
2							F															-
	30421		- - -		-+			-+-							-+			+			CGTC + CAG	30480
2		G	L	F	A	D	R	Т	L	R	A	G	L	Т	V	s	L	V	N	Α	V	-
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2		I	M	G	G G	T	G	L	M	V	A	L	Y	L	Q	T	I	A	G	H	GAGG S	-
	30541				-+-			+-		- ·		+			-+-			+			ACAG	30600
2		GG(L L	CCG(A	GCG(A	G G	CGAC L	W	JGA(L	L L	I I	P	A A	JAC(JATE M	L L	V	V V	G	V V	rgtc Q	-
	30601				-+-			+-				+			-+-			+	- 		GCTG	30660
2							CCG(A														CGAC L	-
	30661				-+-			+		-		+			-+-			+			CCTC	30720
2							TGT(Q													GGA L	GGAG L	-
	30721				-+-			+				+			-+-			+			GGGC	30780
2		TA(CCG(A			GGA(L									G G			GTG T	GTG T	CCCG G	-
	30781				-+-			+				+	-		-+-		- 	+			CACC	30840
2							GCG(A														GTGG T	-
	30841				-+-			+				+			-+-			+			CGTG	30900
2		CC G	GCC G	GCT E	CAA F	GCC G	TCA: V	CCG A	CGA L	GCC G	GTA I	GCG A	CCC G	GGA L	.CCC G	CTC. S	AGA L	CCC G	GTG T	GCA V	GCAC V	-
	30901				-+-			+				+	- 	-	-+-			+	-		GCAG	30960
2		AT Y	GTC S	GCG A	GCC G	CCA V	GCT E	CCA V	CGG P	CCT	GCG A	CCG A	GCC G	CGG P	GCG A	GCT D	GCG A	GCI D	'GCG A	GCG A	CGTC Q	-
	30961		- 		-+-			+				+			-+-			+	. – – –		.CGCC +	31020
2		CT E	CTC S	GTA I	.GCG A	GCC G	GCG A	GGA L	CGT H	'GTG T	CCG A	GCC G	AGT Q	CGA L	A.CCG	TGG P	CCC G	GTC S	:GCG A	GCT D	GCGG A	-
	31021				-+-			+				+			-+-				- 		CTGC	31080
2		GA L	CGA L	CCT D	GAG S	GCG A	CGC R	GCG A	CCG A	GAA F	GTO. T	GTC S	:GCC G	GCA V	CGT Q	'CAG S	GCA V	GCG A	GCC A	GCA V	.GACG C	_

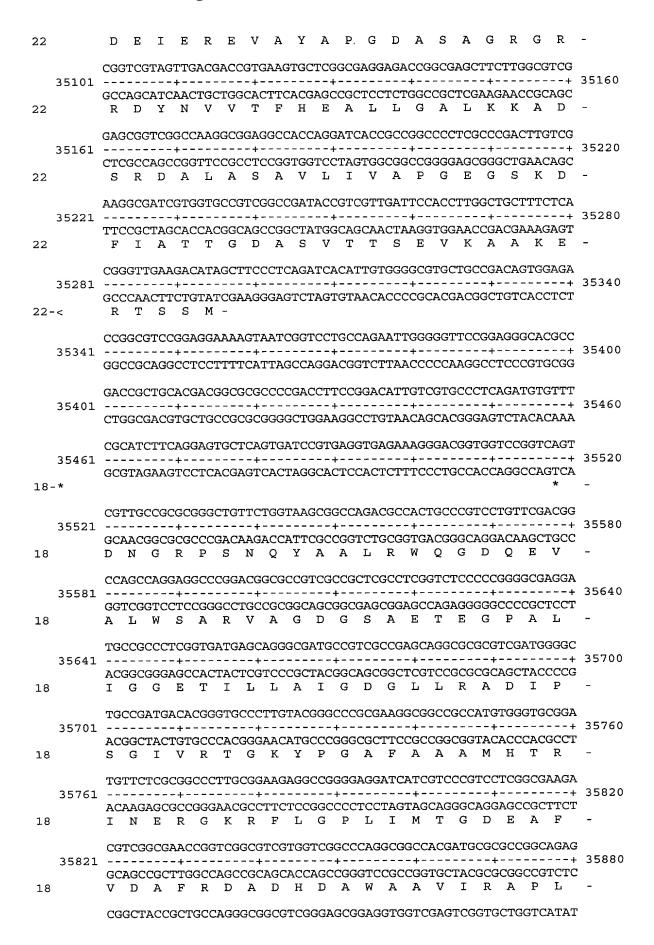
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2	31081	CGG(CAC	AAG	AGC	GAC	CGC	GAC	CGC	CAC	GAC	ATE	GCC	GTG	GG	CCG.	ACG	CC	CTG	TAA	AGG	CGC	-
2-*	31141	ATGO TACO M I	TG	 GTG	+		 GCCG	-+- CTC	CTI	GGC	CCG	 3CT	 CTT	 GCT	GC	 GAG	 TTC	GC	- + - CGG			+	31200
	31201	CAC'	-		+			-+-				+			-+				-+-			+	31260
3->	31261				+		. -	TCF		CA?	rga(+ CCT"	 TGT	 CGI	- + :GC	 CGC	 CT(TG	- + - GTA	.CGG	GGC	CGGT	31320
3	31321	CCT		GCI	+ TG1	CAC	CTT	-+- CAC	CCG	CGT	CCG	+ AGG'	 TCC	 GGC	- + :GG'	 TAC	 CT(3GT	-+- GCG	GT(CTTC		
3	31381		 CGG	GAA	+	CCG		+- TTG(CTG	AGG	: 3GC'	+ FCT	 TGT	AGT	+ rgg.	agg	 TA(CCG	- + - CCI	GAT	rgco	CGGC + GCCG A	31440
3	31441	CGC	 CCA	 CGG	+ BAG		 ACGO	- + - CGTT	rcc	: 3GC'	rgg.	+ AGG	 AGC	GGC	+ CGC	 CTI	CG	 3CG	-+- CAG	AGG	GGG	JAAT	31500
3	31501	GCC	 GTG	GAC	+	- -	GCG/	+ 4CC:	rag(3CC	 GCG.	+ AGC	CTC	'ACC	+ 3CG	GTA	GT(GGT	-+- CTG	GT	CGC	CGTG	31560
3	31561	GTC	 GCC	 LTA	+- GGG	GGG	 3GT2	+ - AGG(CCT	GCA	agc	+ TGT	GGC		+ 3CG	 CTC	AC	CCG	- + - GAC	CA	CGC	rgga + acct D	31620
3	31621	GTG	 CAA	GA(- + - · CGT(GCC	GCG2	+ : AGG'	rgt(CGT	 ACC	+ CGC	ACG		+ 3GC	GTO	 GT	 GTT	-+-	CAG	ACC	rggc + ACCG A	
3	31681	CAA	GCC	GA?	- + - ·	CCG	ACA	+ AGC	GGC	 CAA	 AGA	+ CCC	 CGT	'AC	+ GTG	 ATC		 GGA	-+- .GCT	CG	CGT	TGGG + ACCC G	31740
3	31741	GCG	 GTG	CCZ	- + - AGT2	 AGG(GCC	+ 3GC	 CGC	 CGG.	 AGC	+ TGA	GGG		+ AGG	GCC	CA	 TGC	-+- :CG2	ACG	ACC.	TCGA + AGCT D	31800
3	31801	GAT	 GGI	CT	- + - AGC'	 TCC.	 ACG	+ AGC	 CGG.	 AGT	 GTG	+ GCI	'CGZ	ATA	+ CGC	 TAC	 CGC	CGA	CT	 \GC'	 TCT	CGGC + GCCG A	
	31861																					GGGC	31920

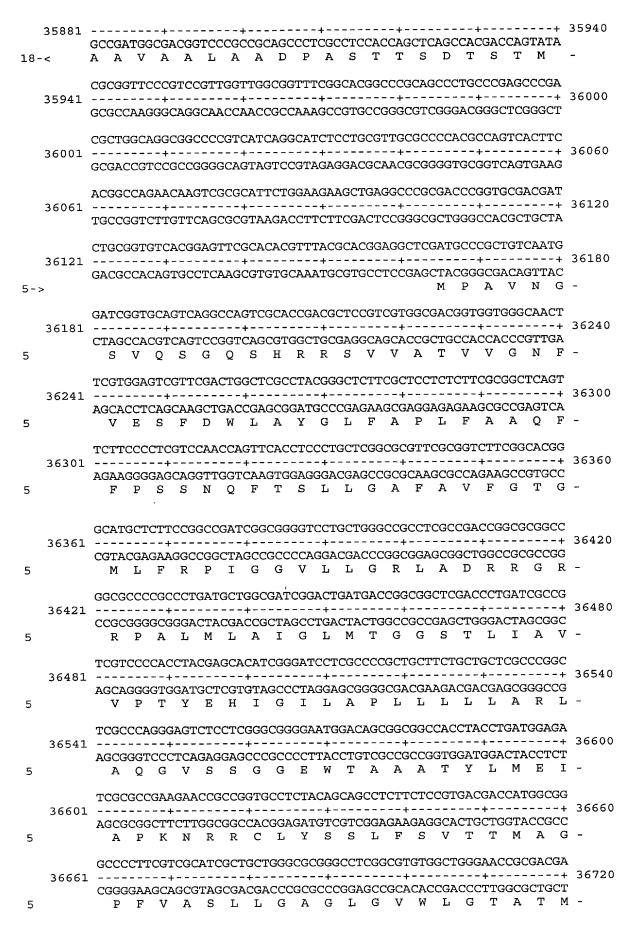


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4->		M P H G	-
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4	3204-	CGTCTCGCGCTTCGCGGCCGGCCGCCGCCGCGCGCGCCGC	-
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4		S L H D P A Q K A V L T A P T D A G D G	-
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4		$\begin{array}{cccccccccccccccccccccccccccccccccccc$	-
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4		CGGGACCACCGGTGGCACGGCTACCTGCGCCTGCCGGAGTTCCACTAGACGGCAGGGAGG A L V A T V P M D A D G L K V I C R P S	33540
4	33541	TACTCCGCAAACGCGGCGACCACGGGCAGCCCGTTCGACAACCCGCTGTCCTCACGGCTG ATGAGGCGTTTGCGCCGCTGGTGCCCGTCGGGCAAGCTGTTGGGCGACAGGAGTGCCGAC Y S A N A A T T G S P F D N P L S S R L	33600
4	33601	GACGAGAACGACGCCATCCTCGTACTCGACCAGGTGCTGATCCCCTGGGAGAACGTGTTC	33660
4		GTCTACGGCAACCTGGGCAAGGTACATCTCCTCGCCGGACAGTCCGGGATGATCGAACGC+ CAGATGCCGTTGGACCCGTTCCATGTAGAGGAGCGGCCTGTCAGGCCCTACTAGCTTGCG V Y G N L G K V H L L A G Q S G M I E R	33720
4	33721	GCCACCTTCCACGGGTGCACCCGGCTCGCCGTGAAGCTGGAGTTCATCGCCGGGCTGCTG+ CGGTGGAAGGTGCCCACGTGGGCCGAGCGGCACTTCGACCTCAAGTAGCGGCCCGACGAC A T F H G C T R L A V K L E F I A G L L	33780
4	33781	GCCAAGGCGCTGGACATCACCGGGGCGAAGGACTTCCGCGGTGTGCAGACCCGGCTCGGA CGGTTCCGCGACCTGTAGTGGCCCCGCTTCCTGAAGGCGCCACACGTCTGGGCCGAGCCT A K A L D I T G A K D F R G V Q T R L G	33840
4	33841	GAAGTCCTGGCCTGGCGCAACCTCTTCTGGTCACTGTCGGACGCGGCCGGC	33900
4	33901	GTCCCCTGGAAGAACGGCACGCTCCTGCCCAACCCTCAGGCGGGTATGGCCTACCGCTGG CAGGGGACCTTCTTGCCGTGCGAGGACGGGTTGGGAGTCCGCCCATACCGGATGGCGACC V P W K N G T L L P N P Q A G M A Y R W	33960
4	33961	TTCATGCAGATCGGCTACCCGCGGGTCCTGGAGATCGTCCAACAGGACGTGGCCAGCGGC AAGTACGTCTAGCCGATGGGCGCCCAGGACCTCTAGCAGGTTGTCCTGCACCGGTCGCCG M Q I G Y P R V L E I V Q Q D V A S G	34020
4	34021	CTCATGTACGTCAACTCCTCCACGGAGGACTTCCGCAACCCCGAGACCGGCCCCTACTTG+ GAGTACATGCAGTTGAGGAGGTGCCTCCTGAAGGCGTTGGGGCTCTGGCCGGGGATGAAC L M Y V N S S T E D F R N P E T G P Y L	
4	34081	GAGAAGTACCTCCGGGGCAGCGACGGCGCAGGCGCCGTCGAGCGTGTCAAGGTGATGAAG+ CTCTTCATGGAGGCCCCGTCGCTGCCGCGTCCGCGGCAGCTCGCACAGTTCCACTACTTC E K Y L R G S D G A G A V E R V K V M K	
4	34141	CTGCTGTGGGACGCGGTGGGATCCGACTTCGGCGGCCGGC	
4	34201	TACTCCGGGAACCACGAGAACACCCGGATCGAGTTGCTGCTGTCGCAGACGGCGAGCGGC ATGAGGCCCTTGGTGCTCTTGTGGGCCTAGCTCAACGACGACAGCGTCTGCCGCTCGCCG Y S G N H E N T R I E L L L S Q T A S G	
	34261	AAACTGGACTCGTACATGGACTTCGCCCAGGCATGCATGGACGAGTACGACCTGGACGGC	

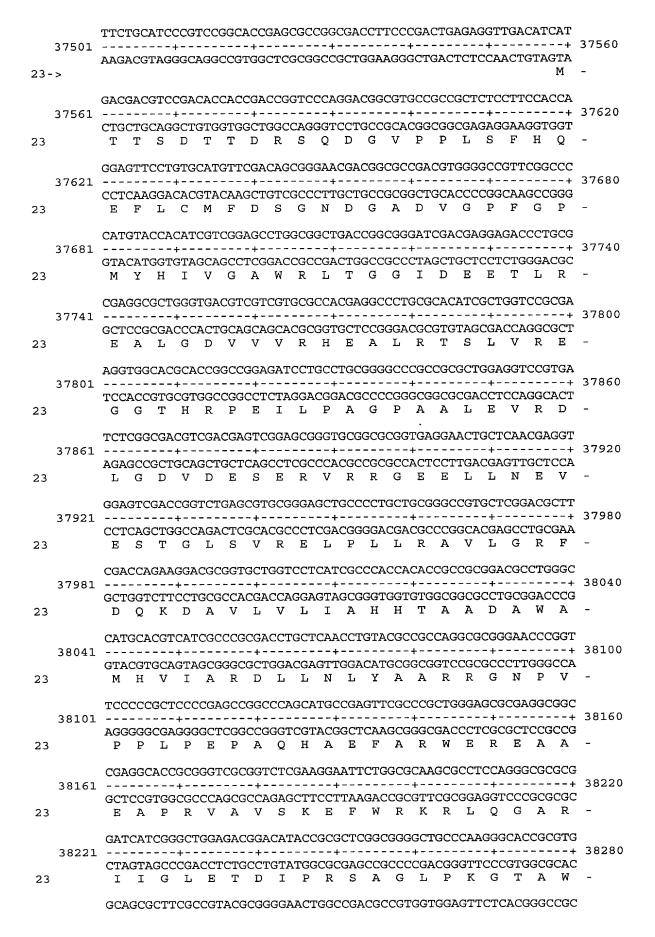
4		TTTGACCTGAGCATGTACCTGAAGCGGGTCCGTACGTACCTGCTCATGCTGGACCTGCCG K L D S Y M D F A Q A C M D E Y D L D G -	-
4		TGGACCGCTCCCGACCTGGAGTCGTTTCACGCGATGCGTTCCGCCTCCCGCGACCTTCTC	34380
4-*		GGAGGGCTGTAGTTCCCCGACGGTGTACTGCGGCCCCCGATCCGGGGGCCGCAGTACACC+	34440
22-		GTCGGGGCGGCTGGTGCTCAGCCGCGCAGGAATCCGATGAGCTCGGGGGCGAGCTTCTTG	
22	34501	GGCGCCATGGCGACGGCACCGTGGTTGAGCCCGTTCAGGGTGCGGTGGCTCGCGTCGGGG CCGCGGTACCGCTGCCGTGGCACCAACTCGGGCAAGTCCCACGCCACCGAGCGCAGCCCC PAMAVAGHNLGNLTRHSADP	34560
22	34561	AGGACTCCGGTGAGTTCCTTCGCGGCACGCTGGAAACCGTCGGGGCTCTTGGAACCGGTC TCCTGAGGCCACTCAAGGAAGCGCCGTGCGACCTTTGGCAGCCCCGAGAACCTTGGCCAG L V G T L E K A A R Q F G D P S K S G T	34620
22	34621	AGCACCAGGGTCGGGGGCCGACGCCGACCACGGCTCGGCGGGGAGCGGCTTGCCCTGC TCGTGGTCCCAGCCCGGCTGCGGCGGGCTGGTGCCGAGCCGCCCTCGCCGAACGGGACG L V L T P A S A A S W P E A P L P K G Q	34680
22	34681	TGGGTGTCGCCCATCACCGCGATGTCGTAGGGAAGCGTGTTGGCCAGACCCTTGAGGTTG	34740
22	34741	GACCAGACACCGGGCATCAGGCGCATGGCGCCGACCATGAAGGAGGGCATGCCCTGTGCC+ CTGGTCTGTGGCCCGTAGTCCGCGTACCGCGGCTGGTACTTCCTCCCGTACGGGACACGG S W V G P M L R M A G V M F S P M G Q A	
22	34801	TTGACCATGAAGGCCTTGACCGCGTCGCTGCGTCGGTCCTCCGCCAGAAGGCTGTCGATC	
22		TGACCGCCGAAGCCGGCGGGCGGGCCGAAGCCGTCCGAGGTGACGGAGAACGGCGGCTCG ACTGGCGGCTTCGGCCGCCCGCCCGGCTTCGGCAGGCTCCACTGCCTCTTGCCGCCGAGC Q G G F G A P P G F G D S T V S F P P E	
22		TAGACCGCGAGCTTGTTCACCTTCAGGCCGGCGGCGGCGGCTCGCAGGGCGAGCACCGCG+ ATCTGGCGCTCGAACAAGTGGAAGTCCGGCCGCCGCCGCCGAGCGTCCCGCTGGCGC Y V A L K N V K L G A A A A R L A L V A	
22		CCGGAAGAGCTGCCGAACAGGGAGGCCGAACCGCCGACCTGGTCGATCAGCGCCGCGATG	
		TCCTCGATCTCGCGCTCGACCGCGTACGCCGGACCGTCGGCGCTGGCGCCCCGA	35100





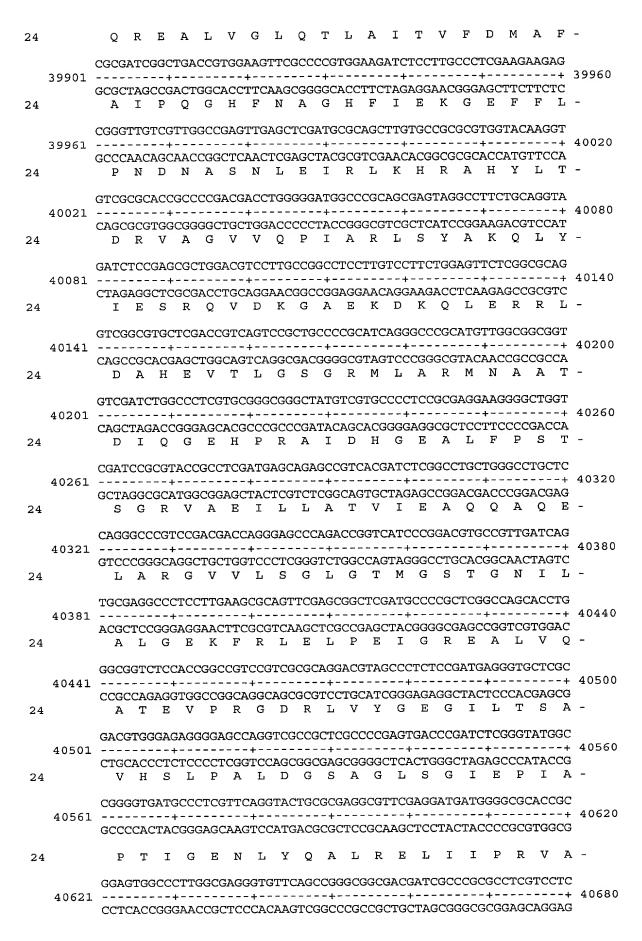
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		TGGAGG	CCTG	GGG	CTGG	3CGC	GTG	CCG	TTC	CTC	CTC	CGG	CGG	CGTC	TTC	CGGC	CGTC	ATC	CTGC	26700
		ACCTCC																		36780
5		E A																		
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		TGATGC'	TGGT	GGC	CGGI	ACTO	3GG(CGT	CATO	CGG	CGG	AAC	GTG	GTC	3AC	CGC	GT(ccc	GCGZ	4
			+				+	- -		+			+				+			- 36960
5		ACTACG. M L																		
3																				
	26061	TGGGCC.																		7 - 37020
	36961	ACCCGG																		
5		G H	R	L	I	G	s	Q	Т	M	F	W	V	V	V	С	V	Т	G S	5 -
		CGGTCA	TCCT	GCT	GCA	GGT	ACC	CAT	AGG	3CT(GCT	CGC	CGA	CCG	GT(GGA.	ACC	GG(CAGG:	ľ
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5		GCCAGT V I																	R I	
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	37001	AGGACT	AGCA	GAG	GTC	GCA(GCA(GAA(GCG(3CG	ACA	CCC	GAG	CAT	GCG(GAT	GGA(GTG	GCAG	
5		L I	V	S	S	V	V	F	A	A	V	G	S	Y	A	Y	L	Т	V	2 -
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5		D S																		
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	37201																			+ 37260
		AGTGGT	'ACCA	CGA	.CGG	CTT	CTA	CGA	GAG	GTC'	TTA	GAA	GGG	AGG	CGT	CTA	TGC	GCC	GGAC	C
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5		GGCGGA A Y																		
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	27201	TCCTGC	TCGC	CCTG	GCC	GGC	CAC	CCT	GTG	GGA	GCG	ACG	GCI	GTT	CCG	CGC	CCG	GAC	GGCC	C + 37440
	3/361	AGGACG																		
5		L I	A	W	P	A	T	L	W	E	R	R	L	F	R	A	R	Т	A	P -
		CGGGAA	.GCG	AGCC	GGT	TCC	CGA	ATC	CGC	CGT	CGC	:CCG	ccc	CGT	CGG	GTG	ACC	GTC	CGCA	С
	37441																			+ 37500
5-	*	GCCCTT G S																UAG.	3031	



23		CGTCGCGAAGCGGCATGCGCCCCTTGACCGGCTGCGGCACCACCTCAAGAGTGCCCGGCGQQRFAVVRGELADAVVEFSRAAA	38340
23	38341	CAAGTGCTCCCCGTTCATGACCATGTTCGCCGCCTACCAGGTGCTGCTGCACCGCAGGAC+ GTTCACGAGGGCAAGTACTGGTACAAGCGGCGGATGGTCCACGACGACGTGGCGTCCTG K C S P F M T M F A A Y Q V L L H R R T	38400
23	38401	GGGCGAGCTGGACATCACCGTGCCGACCTTCTCCGGGGGGGCGCAACAACTCGCGGTTCGA+ CCCGCTCGACCTGTAGTGGCACGGCTGGAAGAGGCCCCCCGCGTTGTTGAGCGCCAAGCT G E L D I T V P T F S G G R N N S R F E	
23	38461	GGACACCGTCGGTTCCTTCATCAACTTCCTGCCGCTGCGTACCGACCTCTCCGGATGCGC CCTGTGGCAGCCAAGGAAGTAGTTGAAGGACGGCGACGCATGGCTGGAGAGGCCTACGCG D T V G S F I N F L P L R T D L S G C A	38520
23	38521	ATCCTTCCGCGAGGTCGTGCTGCGCACCCGCACCACCTGCGGAGAGGCGTTCACCCACGA TAGGAAGGCGCTCCAGCACGACGCGTGGGCGTGGTGGACGCCTCTCCGCAAGTGGGTGCT S F R E V V L R T R T T C G E A F T H E	38580
23	38581	GCTGCCCTTCTCCCGGCTGATCCCGGAGGTGCCGGAGCTGATGGCGTCGGCGGCCTCCGA CGACGGGAAGAGGGCCGACTAGGGCCTCCACGGCCTCGACTACCGCAGCCGCCGGAGGCT L P F S R L I P E V P E L M A S A A S D	38640
23	38641	CAACCACCAGATCTCCGTCTTCCAGGCCGTGCACGCGCCCGCGTCCGAGGGGCCCGAGCA GTTGGTGGTCTAGAGGCAGAAGGTCCGGCACGTGCGCGGGCGCAGGCTCCCCGGGCTCGT N H Q I S V F Q A V H A P A S E G P E Q	38700
23	38701	GGCCGGGGACCTGACGTACTCGAAGATCTGGGAGCGCAGCTGTCGCAGGCGGAGGGCTC CCGGCCCCTGGACTGCATGAGCTTCTAGACCCTCGCCGTCGACAGCGTCCGCCTCCCGAG A G D L T Y S K I W E R Q L S Q A E G S	
23	38761	CGACATCCCCGACGGGGTGCTGTGGTCGATCCACATCGACCCCTCGGGCTCCATGGCCGG GCTGTAGGGGGCTGCCCCACGACACCAGCTAGGTGTAGCTGGGGAGCCCGAGGTACCGGCC D I P D G V L W S I H I D P S G S M A G	
23	38821	CAGCCTCGGGTACAACACCAACCGCTTCAAGGACGAGACGATGGCGGCCTTCCTGGCCGA GTCGGAGCCCATGTTGTGGTTGGCGAAGTTCCTGCTCTGCTACCGCCGGAAGGACCGGCT S L G Y N T N R F K D E T M A A F L A D	
23		CTACCTCGACGTGCTCGAGAACGCGGTGGCCCGGCCGGACGCCCCCTTCACCTCCTGAGA+ GATGGAGCTGCACGAGCTCTTGCGCCACCGGGCCGGCCTGCGGGGGAAGTGGAGGACTCT Y L D V L E N A V A R P D A P F T S * -	38940
26		CAGTTCCGGCGGCGAACCCGCCCGAAGAAAGGAAAGCCAGTGTCCACCGTTTCCGAC+ GTCAAGGCCGCCGCTTGGGCGGGCTTCTTTCCTTTCGGTCACAGGTGGCAAAGGCTG M S T V S D	
26		ACAGCGGCCGGCTCCTCCCTGGAGGAGAAGGTCACCCGGATCTGGACGGGTGTTCTCGGC+ TGTCGCCGGCCGAGGAGGACCTCCTCTTCCAGTGGGCCTAGACCTGCCCACAAGAGCCG T A A G S S L E E K V T R I W T G V L G	39060
	39061	ACGTCCGGTGAGGAAGGCGCGACGTTCATCGAGCTCGGAGGGCAGTCGGTCTCGGCCGTG	39120

26		TGCAGGCCACTCCTTCCGCGCTGCAAGTAGCTCGAGCCTCCCGTCAGCCAGAGCCGGCAC T S G E E G A T F I E L G G Q S V S A V -	
26	39121	CGCATCGCCACGCGTATCCAGGAGGAGCTCGACATCTGGGTCGACATCGGCGTCCTCTTC	180
26	39181	GACGACCCGGATCTGCCTACCTTCATCGCGGCGGTCGTCCGGACGCCGACGCCGCGGGC CTGCTGGGCCTAGACGGATGGAAGTAGCGCCGCCAGCAGGCCTGCCGGCTGCGGCCCCC D D P D L P T F I A A V V R T A D A A G -	9240
26 -		GGCGAGGGCTCCGGAACGCAGTGAGACTCGCCGGGCGCCCGTCTCCCCGCGGCGCCCGGTT	9300
	39301	TCACATGGCTGAGGCGGTTCACCCGGTACCGGGTGAACCGCCTCAGCCATGTGAAACCGG	9360
24-		GCCTGGTCAGCGCAGCTGGATGTCCGTCTCCCGGGCGATCGCCCGGAGGAACTCGCCGCG ++++++	9420
24	39421	GGACAGCGCGTCGGCGACCAGCTCGATGTCGTCGGCCATGTACCGGTCGACGCCCAGCGT CCTGTCGCGCAGCCGCTGGTCGAGCTACAGCAGCCGGTACATGGCCAGCTGCGGGTCGCA S L A D A V L E I D D A M Y R D V G L T -	9480
24	39481	CGGAACCAGCCGGCGCACCGCTTCGTACGTGGCCTTCGCCGCCGGGCTCAAGCCGTCGAA GCCTTGGTCGGCCGCGTGGCGAAGCATGCACCGGAAGCGGCGCCCGAGTTCGGCAGCTT PVLRRVAEYTAKAAPSLGDF-	9540
24	39541	CCGGCCGGAGATGTCGACCGCCTGGGCGGCGGCCAGGTACTCCACCGCGAGGATCTTGTT GGCCGGCCTCTACAGCTGGCGGACCCGCCGCCGCCGTCCATGAGGTGGCGCTCCTAGAACAA R G S I D V A Q A A A L Y E V A L I K N -	9600
24	39601	GTTGTTCGACAGGACCCGGCGGGCGTTGCGGGCCGAGATCAGGCCCATGCTCACCACGTC CAACAAGCTGTCCTGGGCCGCCCGCAACGCCCGGCTCTAGTCCGGGTACGAGTGGTGCAG N N S L V R R A N R A S I L G M S V V D -	9660
	39661	CTGGTTGTCGCCGTTGGACGGACGCTCTGGGTGCTGGCCGGGCCGATCGTCCGGTTCTC 1+	9720
24		Q N D G N S P V S Q T S A P G I T R N E - GGCCACCAGTGCGGTGCCGGGTACTGGGCGCGGCGAATCCGCTGTGCAGCCCCGGGTC	
24		CCGGTGGTCACGCCACCGGCCCATGACCCGCGGCCGCTTAGGCGACACGTCGGGGCCCAG A V L A T A P Y Q A G A F G S H L G P D -	
24	39781	CCCGGAGACGAGAACTCCGGGAGGCCGTAGCTGAGGTGCCGGTTCAGGACCCGGTTGAT 1	
	39841	CTGCCGCTCGGCCAGGACGCCGAGCTGGGTGAGCGCGATGGTCACGAAGTCCATCGCGAA 1+ 3 GACGGCGAGCCGGTCCTGCGGCTCGACCCACTCGCGCTACCAGTGCTTCAGGTAGCGCTT	9900

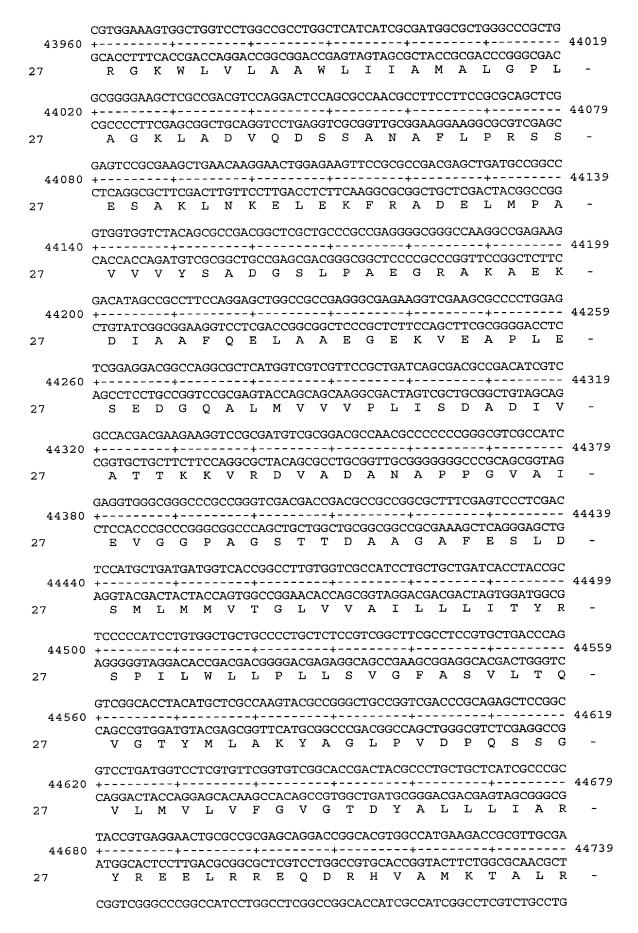


24		s	H	G	K	A	L	T	N	L	R	A	A	V	I	A	R	A	E	D	E	-
	40681			+				+			-+-			+				+			+	40740
24		CCGC A													L L					AGC1 E		-
24	40741	AAGG.	 AAG	+ CTG	AAC	AGC	TGG	+ ACG	TAC	CATO	- + - CTAC	TAG	AGC	GGC		 GGC	 CAC	+ CAG	 TGG		- + T	
24	40801	CTAC	ccc	+ TAC	. – – – !AAG	 ACA	AGC	+ :CGC	TAC	GG <i>F</i>	+ - \AGC	TTC	TAG	+ SAGC	GCC	 GAG	 AAG	+ ACC	 CGG		-+ CG	
24	40861	CTAC	 CTA	+ AGC	CGG	CCA	TGC	+ AGC	TGC	 GCAC	+ - 3CGC	CGCA	AGG	+ 3AG0	 GCGC	TGC	 GCC	+ !GCA	 TGC		-+ AG	
24	40921	CTGC	CAG	TCC	CAG	AGC	 CGGC	+ AGC	TGC	CCT	+- TTG0	 3CC0	TGC	+ CTAC	 BAGC	CAG	 AGC	+ TGA	ACT		-+ TT	
24-	40981	ACGG		+ TG#				+			+-			1	- - -			+			-+	41040
25-		TGAG ACTC					GCC	+ CACI	CGC	GCC	+- CTC	GCCI	 4GG:	rgg <i>i</i>	+ \AGC	GGC	GCC	+ CGCA	ACG		-+ CG	41100
25	41101	TCCG	CAC	 TTC	CAGT	rcco	3CCC	+	TC	rgc	+ · CGC(CCG	rca(CGC:	+ rccc	CCC	GCI	+ TGA			-+ CG	41160
25	41161	TCTA		 CTTC	+ 3CG(GTC	CGGC	- + 3CCI	raci	AGG	+ · CGC(GCG	rgcz	AGGZ	+ AGG(3CCC	GGZ	+ \GAC	GAA		-+ CG	41220
25	41221	TGGT	GC	TG:	+ rgc(CGC:	 rcco	- + 3CC(CAG'	 TGGʻ	+ TCC	GGG	ACC	GGC	+ : AAC:	rgc <i>i</i>	ACCO	-+- CCCI	. – – . CCI		-+ GC	41280
25		GTG	 AGGT	CC'	+ · TGG(CTC	CCC	- + - · AGT(3CC	 AAGʻ	+ TCG	CGC	CGG.	AGC'	+ · TAG	AGC	CAC	- + CA <i>I</i>	AGGI		-+ CC	41340
25		TTCC AAGC N	: 3GC:	 rtg'	+ rcg2	AAC'	TGG	- + - ·	rtc.	 AGG	+ AAC	 GCC	 GGG	GCC	+ TTA	AGG	rcc	-+- CGA(GGCI	- \GCT	-+ TG	
25	•	CGTA GCA:	rgg(GCG	+ CGG'	 TCT	AGG	- + - · GGC(CAG	 GCC	+ ATG	 GTG	 GCC	 AGT	+ GGC	AGG	: CCC(- + - CGC:	rcc	 GGCC	-+ :GC	41460

	47.467	AGGGGCGCGAACAGCGCGCTGTGGTCCGGGCCGCCCTCGACGCGAGATAACCCGGCGTC+ TCCCCGCGCTTGTCGCGCGACACCAGGCCCGGCGGAGACTGCCGCTCTATTGGGCCGCAG													
25	41461	CCCCGCGCTTGTC L P A F I	CGCGCGACACC	AGGCCCGGC	GGAGCTGCCGCT	CTATTGGGCCGCAG									
	41521		+		-	CTCGGCCGGTCGTCC									
25					rgcggccgcccc V G A P	AGCCGGCCAGCAGG SPRDD	-								
	41581					GGGCCGGGCCCGTG + CCCCGGCCCGGGCAC	41640								
25		A D V V I	V Q R	G P L	P Y G I	P A P G T	-								
25	41641		AGAGCACGGTC	CCAGCGCCGC	ragcagagccac	P G Y L N	41700								
	41701					rcgccgggcagcgcc + Agcggcccgtcgcgg	41760								
25		ILRT	RPLA	A R L	G D V L	E G P L A	-								
	41761	AGCGGGTAGTCCT	CGTCCACCGG	TCCCACGGC	CCGGCTAGCGGG	GGTCGGAGGCGGTG + CCAGCCTCCGCCAC	41820								
25		ATCACTCCCAGGA	GTCCCGGGC	GAAGCTGGGC	PRDG ACGGTCTGGAGA'	IGAGTGATCCGCTCC	- !								
25	41821	ragtgagggtcct(I V G L	CCAGGGCCCG	CTTCGACCCG F S P	TGCCAGACCTCT	ACTCACTAGGCGAGG	41880								
	41881					TCCGGCACCGGACAC ++ AGGCCGTGGCCTGTG	41940								
25		Q V L W	PVLK	D P N	V R V R	E P V P C	-								
	41941					GGGTCGTGCTCCGGG + CCCAGCACGAGGCCC	42000								
25						PDHEP									
	S	Q ID No. 2. C-	1027 gene cl	uster DNA	sequence from	1 41,980 to 63,164									
	41980	+				CCCGGCCCCATCGCC	42039								
25						GGGCCGGGGTAGCGC . G P G M 1									
23		AACCGTTCGCCCA	TCCAGCCCGC	GAACTGGCCC	AGCGCGGCATGC	GACTGGGCGATCCC	C								
25	42040	TTGGCAAGCGGGT	AGGTCGGGCG	CTTGACCGGG	TCGCGCCGTACG	CTGACCCGCTAGGG	3								
	42100	+				TCTGCCAGGCCCGGCC	- 42159								

25		K	P	R	G	T	S	G	S	Т	F	Α	V	Y	A	L	D	A	L	G P	-
	42160	CCCC			 -			-+		- -	+				+			-+-		AGGC	G 42219
25																				S .	
25	42220	+	CGT	CCCZ	+ ·	GCG(GGC	-+	TG	 3CC	+ 3CA'	 TGGʻ	 TCT	 CGC	+ GGT	CGC	CTA	-+- GGA	GGA	CGCCT	42279
	42280	+		- -	+			-+			+				+			-+-			42339
25			GCA(G					C												CAAGC R E	
25	42340	+	 GCG	 GCA(+ GGC	 CTC	 GCT'	- + - - TGGT	rgc	GGT	+ CCA	 CCC	 GCG	 GGC	+ GGA	- GGT	 CCT	-+- GAG	GGT	GCAGC CGTCG L L	42399
25	42400	+ TGGC	 GCT	AGG	+ CCG	 CCC(GCG(-+ GGC(CGA	 CGT	+ AGG	 CGT	GGC	 GGT	+ 'GGC	 CGC	 TCG	-+- GCA	CGG	CCGCG GGCGC G A	42459
25	42460	+ GGCC	 GGC	GCC	+ ACT	CCC	 GGC'	-+-	GCT	 GCG	+ CCC	 GGC	GCA	.GGC	+ :GCC	AGT	 CAA	-+- GGC	GCC	rcagt agtca T L	42519
25		+	 GCA	TCG	+ AAC	 ACG	 CGC.	-+- ACG	 GCG	 GCT	+ TGC	 TCT	GCC	GCT	+ GTG	 GCA	 GCA	-+- .CAA	 GGC	CGTGG GCACC H -	42579 !
25	42580	+	 CCT	 GGC'	+ TCC	 GCA	 CGT	-+- GGC	 CGG	 CGC	+ AGT	ACA		GCG	+		-	-+-		CCGAA GGCTI	42639
	42640				+			-+-			+			- - -	+			-+-		GTGTG CACAC	42699
	42700	+			+			-+-			+				+			-+-		CCCGG	42759
	42760	+			+			-+-	- 		+				- +			-+-		CGGTC GCCAC	42819
	42820	+	-		+			-+-			+				- +	- 	- 	-+-		GCCT(CGGA(42879
	42880	+			+			-+-			+	-			-+	-	-	-+-		CGGC(.GCCG(- 42939
	42940	+			+			-+-				- - -			-+			+ -		GGAAG	- 42999

43900	CCGAAGGAGCCCCCTTGCAAGCCTGGTTCAAGCGCACCAGTGGTGTGCCCGGTGACAGA +++++++	43959
43840	AACGGCCGTGCAGTGCGGCCCTCTCCAACACTCCCGGCCATCGCGGAATCCGAGACGTGC +	43899
43780	GGAAGTTGACGTCTTCCAGGTGGAGTTGGGAACGATGGAGGCCCCCGCCGGCCG	43839
43720	CTCGAGGCGCATGCCGCGGCTTCGCAAGCCAGTTTGGAACAAGCTGGTTGACGCGCGCG	
	AGTAAGCGGTGCTGCGGGTAGAACCCCGCCGCGCGTGTCCTGCGAAGAGGAACTCACGC GAGCTCCGCGTACGGCGCCGAAGCGTTCGGTCAAACCTTGTTCGACCAACTGCGCAATCT	
43660	TCATTCGCCACGACGCCCATCTTGGGGCGGCGGCGCACAGGACGCTTCTCCTTGAGTGCG	
43600	++	43659
	TGCAACACCGCAGATCCGGAGGCCGGCGAAGGCCCCCGGTCCGGCTCGGTCACAGTCACG	
43540	CAGGCCGTCCGCAGGCCTCCAGGTCGAGTCCGCCGCTCACGTGGTAGGCCGCGTACGGG +	43599
43480	+++	43333
	CCGCACTCCTCGTCGGCACTCGGCCCGTCCACGGCGGCCCCGTCTCCGGCGGCGGCCCCGC	42E20
43420	++	43479
	GCGGGCGCTCACCCTCGGCGAAGGGACGGGCCCCAGCGAGCG	
43360	TGCGAGGCCACCAGGAGAAGCAGGTGCGCGCGTGGGCCTCTGCGGGCGATGTGGAGCCGT +++++	43419
	TCCATGCGCAAGCTGTTGCGCCGGCCGTCCCCGGGCCTGCCGGGCAGGGTCCGAGGCCTC	
43300	AGGTACGCGTTCGACAACGCGGCCGGCAGGGCCCGGACGGCCCGTCCCAGGCTCCGGAG	43359
43240	+	43299
	GGCGCCCGGGGCGTTTGCGGCGCGGGGCGCAGTGGGAGGCCGGCC	42200
43180	+	43239
	CGGTTCCGGTCGCCGGCAGGACCTGTCCGGAGGCGGTCGCCGAAGACTCCTCGCTCCCG	
43120	+	43179
	CGGTCGTGGTGCCGGCCCCCCCGCGAAGGCGAAGCCGCTCGGCCCGGCATGCCTTC CCGAGGTCCGGACTCCAGCCGAAGGCGACGGTGCTCCCCGCGTGCGAGGCAGGTGCGGG	
43060	GCCAGCACCACGGCGGCGGCGGTGACCCCTTCCGCTTCGGCGAGCCGGGCCGTACGGAAG	43119
	CGCCACCGGCCGCACGGCAGGACAGGCCGCATCGCGACCTGGTCCCGAGCCTTAGGC	
43000	GCGGTGGCCGGCGTCCTGTCCGGCGTAGCGCTGGACCAGGGCTCGGAATCCG	43059



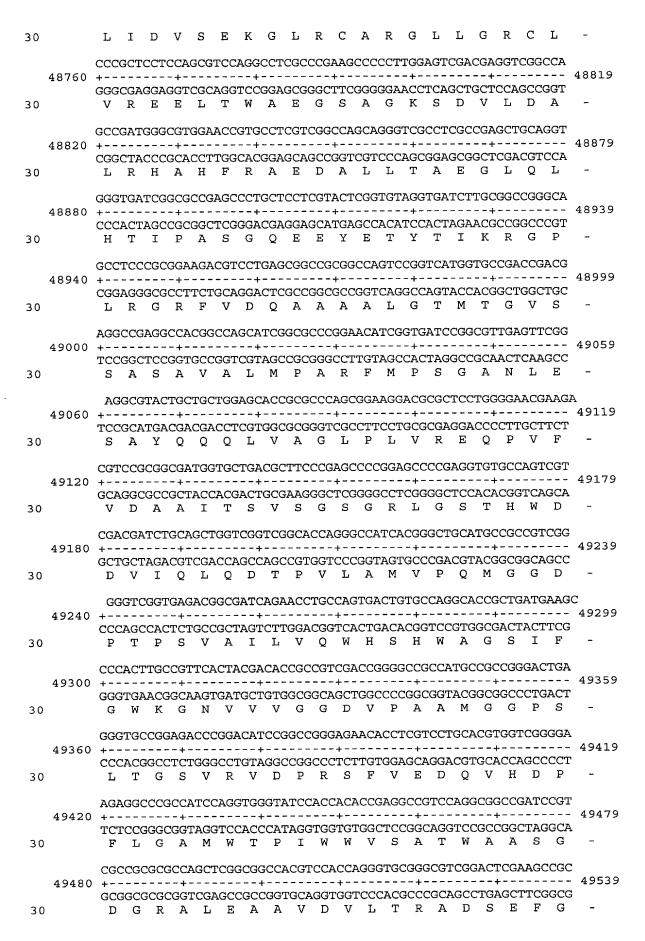
27		GCCAGCCCGGGCCGTAGGACCGGAGCCGGAGCAGACGGAC R S G P A I L A S A G T I A I G L V C L	44799 -
27	44800	GTCCTCGCGGACGTCAACTCCTCCCGCTCCATGGGCCTGGTCGGCGCGATCGGCGTGGTC ++ CAGGAGCGCCTGCAGTTGAGGAGGGCGAGGTACCCGGACCAGCCGCGCTAGCCGCACCAG V L A D V N S S R S M G L V G A I G V V	44859 -
27		TGCGCCCTCCTCGCCATGGTCACGATCCTGCCCGCGCTGCTGGTCATCCTGGGCCGCTGG ++ ACGCGGGAGGAGCGGTACCAGTGCTAGGACGGCCGACGACCAGTAGGACCCGGCGACC C A L L A M V T I L P A L L V I L G R W	44919
27		GTGTTCTGGCCCTTCGTTCCCCGCTGGACGCCGGAGTCGGCCGCGGGCCCCCGAGGCACCG ++++++	44979
27		GCGTCCCACAGCCGCTGGGAGCGCATCGGCTCCGTCACGGCCGCCCGGGCCGCCGCGCCGCCCGC	45039 -
27		TGGGTGCTGTCCTTGGCCGCGACGGGGCTTCTCGCCCTCAGTTCCCTCGGCCTCGACATG ++ ACCCACGACAGGAACCGGCGCTGCCCCGAAGAGCGGGAGTCAAGGGAGCCGGAGCTGTAC W V L S L A A T G L L A L S S L G L D M	45099 -
27		GGACTCACCCAGAGCGAACTGCTCCAGACGAAGCCCGAGTCCGTCGCCCAGGAGCGGGCCGGCC	45159 -
27		ATCTCCGCCCACTACCCGTCCGGCTCCTCCGACCCCGCCACCGTCGTCGCACCCAGCGCGCGC	45219 -
27		GACGTGGCCGAGGTCCGCCGGGCCGCCGAGGGGACCGACGGAGTGGTCTCCGTCCAGGAC ++ CTGCACCGGCTCCAGGCGGCCCCGGCGCTCCCCTGCTGCCTCACCAGAGGCAGGTCCTG D V A E V R R A A E G T D G V V S V Q D	45279 -
27		GGCCCACCACTCCGACGGAGAGCTGACCATGCTGTCCGTGGTGCTGAAGGACGTTCCC ++ CCGGGGTGGTGAGGGCTGCCTCTCGACTGGTACGACAGGCACCACGACTTCCTGCAAGGG G P T T P D G E L T M L S V V L K D V P	45339 -
27		GACAGCAGCGGGGCCAAGGACACCATCGATGCACTGCGGGACAACACGGATGCTCTCGTG +	45399 -
27		GGGGGTACGACGGCCCAGAGCCTGGACACCCAGCGCGCCTCGGTCCGTGACCTCTGGGTC +	
2.		ACCGTCCCCGCGGTCCTGCTGGTGGTCCTGCTCGTCCTGATCTGGCTGCTGCGCTCGGTC + TGGCAGGGGCGCCAGGACGACCACCAGGACGAGCAGGACTAGACCGACGACGACGACCACCAG T V P A V L L V V L L V L I W L L R S V	
	45520	ACCGGACCGCTGATCATGCTCGGCACCGTGGTCGTGTCGTTCTTCGCGGCCCTGGGGGCC	45579

27	TGGCCTGGCGACTAGTACGAGCCGTGGCACCAGCACAGCAGAAGCGCCGGGACCCCCGC T G P L I M L G T V V V S F F A A L G A	-
45580 27	TCCAACCTGCTCTTCGAGTACGTGATGGGGCACGCCGGCGTCGACTGGTCGGTGCCGCTT +	45639 -
45640 27	CTCGGGTTCGTGTACCTGGTCGCCCTCGGAATCGACTACAACATCTTCCTCATGCACCGG +++	45699 -
45700 27	GTGAAGGAGGAGGTCGCTCTGCACGGCCATGCCAAGGGCGTGCTCACCGGCCTGACCACC +	45759 -
45760 27	ACCGGGGGGCGTCATCACCAGTGCCGGCGTGGTCCTGGCCGCGACGTTCGCCGTCATCGCC ++++++	45819
45820 27	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	45879
45880 27	GACACCTTCCTCGTCCGGACGATTCTTCTGCCGGCCCTGGCGCTCGATCTGGGGCCCCGG ++++++ CTGTGGAAGGAGCAGGCCTGCTAAGAAGACGGCCGGGACCGCGAGCTAGACCCCCGGGGCC D T F L V R T I L L P A L A L D L G P R	45939 -
45940 27	TTCTGGTGGCCGGGCGCGCTGTCGAAGACGTCCGGGGGACCGGCCCCCGTCCGCGAGGAC +++++++	45999 -
46000 27	CGCACGTCCCAGCCCGTGGGCTGAGACCCGTCCCGACGAGACCCGTACGGCGGCGGCCGCGCCGCCGCCGGCGGCGGCGGCGGCAGGGCAGGGCAGGGCAGGGCAGGGCAGGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGA	46059
46060	GTTCCCCCGGGCCGTACGACTGAGCAACCCAGAAGATGGGCCGCCCGC	46119
46120 28	CGATGGTGGCCCACCGGCCGCAGGCCGATCTCCCGGAAGGAA	
46180 28	AGGACGGCAAGGCCGCCGAGCTGTGGTCGATGGCGAACCTGGGTACACCGATGGCCGTGC ++ TCCTGCCGTTCCGGCGGCTCGACACCAGCTACCGCTTGGACCCATGTGGCTACCGGCACG D G K A A E L W S M A N L G T P M A V	
46240	GCGTCGCGGCGACCCTGCGCATCGCCGACCACATCACGGCCGGAGCGCACACCGCCGGCG ++ CGCAGCGCCGCTGGGACGCGTAGCGGCTGGTGTAGTGCCGGCCTCGCGTGTGGCGGCCGC V A A T L R I A D H I T A G A H T A G	
46300	AAATCGCCGAAGCGGCCGCCGTGCACGAGGAATCCCTCGACCGGCTGCTGCGCTACCTCA) ++ TTTAGCGGCTTCGCCGGCGGCACGTGCTCCTTAGGGAGCTGGCCGACGACGACGCGATGGAGT I A E A A A V H E E S L D R L L R Y L	46359

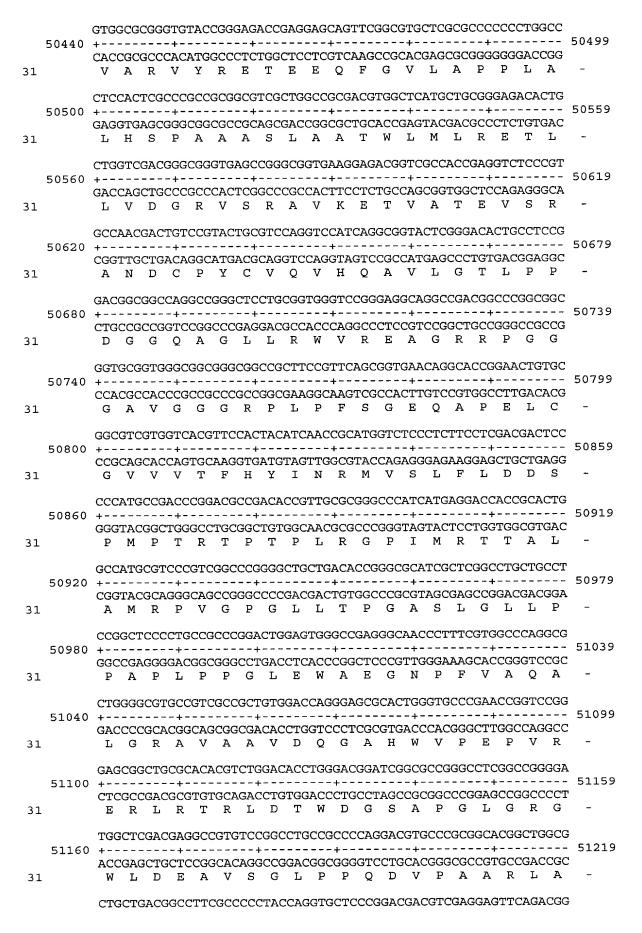
	46360	CCGTC																			4	5419
	46360	+ GGCAG	GCC	+ · CCG(3ACC	3ACC	TTGO	CAC	CTGC	ccc	GAG(CCGC	CCA	TGI	GCC	ACT	'GGC	GGC	3ACC	CGG		3417
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	40420	CCGGC	GAC	ACG	CTC	CTG	TGC	3GG(	CGGC	CCG	CAG(	3CC(	CGGZ	ACCA	AAG	TGT	ACC	CTC	CCT	CGCC		
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28		R	G	Ē	L	S	F	V	D	L	L	Н	S	V	R	T	G	K	Α	A	F.	-
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		AGTCC	TTC	AAC	CGG	CTG	CTC	GC.	CAG	GAC	GTC	GCC	ACT	CGC	GCC	CCG	3CC(	GTG	GTG	GCCG	;	
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28			AAG F	TTG N	GCC R	GAC L	JAG L	G G	0	D	V V	DDJ A	T T	gCG( R	A	P	A A	V	V	A	Ğ	-
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		GCTTC	'GAC'	TGG	GCG.	AGC.	ACC	GGT	CAT	GTC.	ATC	GAC	CTC	GGA(	GGC(	GGC(	GAC(	GGC	TCC	CTGC	; . 4	6719
	46660	CGAAG	ב – – כידים	+ ACC	 CGC	TCG	: TGG(	+ CCA	GTA	cag	-+- TAG	 CTG	GAG	CCT	CCG	CCG	CTG	CCG.	AGG	GAC	;	0719
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28		T	A	L	L	Т	A	С	P	S	L	R	G	Т	V	L	D	L	P	E	Α	-
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28		AGCGC A	G	S	AAG F	F	D	A A	GAG L	P	A	G	A	G	A	Y	V	L	s	L	v	-
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	46900	TCCTC	CAC	GAC	TGG	GAC	GAC	GAG	GCG	TCC	GTC:	:GCG	ATC	CTG	CGG	CGC	TGC	GCC	GAG	GCG	-3 ∠	16959
	46900	AGGAG	CGTG	+ CTG	ACC	CTG	CTG	CTC	:CGC	AGG	CAG	CGC	TAG	GAC	GCC	GCG	ACG	CGG	CTC	CGC	<b>ວ</b> ໍ	.0000
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28		TCAA	JCT( E	E	L GAC	ocici A	G	Doo. R	A	G	L L	R	V V	V	A	V	H	P	A	G	P	-

	47140	CTTCCGCGATCATCCAGATGTCCGCGGTCTGACCGCCCGGAGCCCCGGCCCATCGCGGCG	
28	47140	GAAGGCGCTAGTAGGTCTACAGGCGCCAGACTGGCGGGCCTCGGGGCCGGGTAGCGCCGC S A I I Q M S A V * (ORF28)	47199
	47200	CGGGCCACGGCAGACAAGGAGAGAGAGCGTATGGCCGGCC	47259 -
2:		GCTCGACGCGCTGGGCACGGTGCAGGGGCGTCAGGACCCCTATCCCTTCTACGAGGCGAT ++++++	
29	47320	CCGCGCGCACGGCAGGCGGTCCCCACGAAGCCCGGCCGCTTCGTGGTGGTCGGCCACGA +	47379 -
29	47380	CGCGTGCGACCGGGCGCTGCGGGAACCGGCCCTGCGCGTCCAGGACGCCAGGAGCTACGA ++ GCGCACGCTGGCCCGCGACGCCCTTGGCCGGGACGCGCAGGTCCTGCGGTCCTCGATGCT A C D R A L R E P A L R V Q D A R S Y D	47439 -
29	47440	CGTCGTCTTCCCCTCGTGGCGGTCGCACTCCTCGGTCCGGGGGGTTCACCAGCTCCATGCT +++	47499 -
29	47500	CTACAGCAACCCGCCCGATCACGGCCGGTTGCGCCAGGTGGTGAGCTTCGCGTTCACCCC ++ GATGTCGTTGGGCGGGCTAGTGCCGGCCAACGCGGTCCACCACTCGAAGCGCAAGTGGGG Y S N P P D H G R L R Q V V S F A F T P	47559 -
29	47560	GCCCAAGGTGCGCCGGATGCACGGGGTGATCGAGGACATGACCGACC	47619 -
29	47620	GATGGCCCGGCTCGGCTCCGGCGGCTCCCCGGTCGACCTCATAGCCGAGTTCGCCGCCCG ++	47679 -
29	47680	GCTGCCCGTCGCGGTGATCAGCGAGATGATCGGCTTTCCGGCGAAGGACCAGGTGTGGTT +	
29	47740	CCGCGACATGGCCTCCCGGGTCGCCGTGGCGACGGACGGTTTCACCGACCCCGGCGCGCT +	47799 -
29	47800	CACGGGGGCCGACGCCGCCATGGACGAGATGAGCGCCTACTTCGACGACCTCCTGGACCG +	17859 -
29	47860	TCGCCGCCGCACCCCGGCCGACGACCTGGTCACCCTGCTCGCCGAGGCCCACGACGGCTC +	
		CCCCGGGCGCCTGGACCACGACGACTGATGGGCACCATGATGGTGCTGCTCACAGCCGG	

29		GGGGCCCGCGGACCTGGTGCTTGACTACCCGTGGTACTACCACGACGAGTGTCGGCC PGRLDHDELMGTMMVLLTAG	47979
29	47980	GTTCGAGACCACGAGCTTTCTGATCGGCCACGGGGCGATGATCGCCCTCGAACAACGGGC ++++++	48039
29	48040	GCACGCGGCCCGGCTGCGGGCCGAACCCGACTTCGCCGACGGCTACGTCGAGGAGATCCT +	48099
29	48100	CAGGTTCGAGCCGCCGGTCACCACCCAGCCGGTGGGCTGCCGAGGACCTCGACCTGCT +++++++	48159
29	48160	GGGCCTGTCCGTACCGGCGGGCTCCAAGCTGGTCCTGATCCTGGCCGCCGCGAATCGCGA ++ CCCGGACAGGCATGGCCGCCGAGGTTCGACCAGGACTAGGACCGGCGCGCTTAGCGCT G L S V P A G S K L V L I L A A A N R D	48219
29	48220	TCCCGGCCGCTACCCCGAGCCCGGCCGCTTCGACCCCGACCGCTACGCGCCCGGCCGG	48279
29	48280	CGGGCCGGAGGCCACCAGACCGCTGAGCTTCGGCGCGGGGCGCCACTTCTGCCTCGGCGCCCCCGGCGCCCCCCCGCCGCCGCCG	48339
29	48340	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	48399
29	48400	CCTGGCCGTGTCCGAGCCCCCGTCTACCGCGACCGCTGGGTCGTCCGCGGCCTCGAAACC+	2 48459 -
29	48460	CTTTCCCGTGACCCTCGGGTCCTGAGCCCCGCCGGCCGGAACACGTGACCGTCCCGGCCCCCGCCGGCCG	48519
	48520	GGCGGGTGCGCCCTCTCAGACGTACAGGGTGTTGGGCCCCTGACCACACAGCACCCGG +	48579
3		CCGTACAGCTCCAGGTTGGTGCTCGGGTTCATGCAGGTGCAGCGTGATGCTCTGGGCATC +	48639
30	48640	GCTGCACGCGCTGGATCGGGACGTCGTTGTAGATCGAGGACCCGCCGCTCGCCTGGGCGA +	48699
	48700	GGATGTCCACCGACTCCTTGCCCAGTCGGCACGCCCCCAGCAGGCCGCGCACAGCA +	48759



	49540																		GAC			10500
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30		G	Y	R	A	P	V	R	M	R	F	I	G	A	E	A	M	A	E	V	S	-
		CCT	CGT	GCA	GCCC	acco	GT	гсто	CTC	GGT	rccz	AGG	CCGC	CGTC	GG <i>P</i>	CTC	GAC	CAC	3CGG	CCI	'CA	
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		GGA	GGT	CCT	TAG'	ľAC'.	rct(	3CG(	3GA(	CAG	3CG(	JCA'	PACC	÷CC".	rrcc	FTC(	JGCI	4GA(	CGCG	÷CG1	ľAG	
		GGT																				
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		TCT	TCA	GGT	GCT"	TGC	CAG	GCC	CTT	CGG'	rct:	rac:	ACG	AAG	AGC	CGG	CCT	CAG'	TGC(	CGG	CCG	
		GCC	GGC	GCC	CGT	CGC	CGG'	rca(	CGC	CGG	ACC	ACG	ccc	GGA(	CCG	3TC	ATG	GAG	GCAC	3CC	CAT	
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31	51340	GTCCGTGTCGCACGCTCGTCGTGCCCGACGCCGGGCCGG
	51400	CAACGGCTCGGGAAGGCTGTCTCACGGCCGGAGGCGTACGCCGGTGAGGTGCTCGGACTC +
32	51460	CTCCCAGAGGCGGCCCGGGCCCTGGGGTCGACGGCTGCTCCGCCGGGGCGCACGAGCCC +
32	51520	GGGTGCGCCCCGGGTCTCGGTCACGCCGAGGGGCCCGTAGAACTCGCCCCCGCGCGCG
32	51580	GGGATCGGTGGCCGCCGCAGACCAGGCAGCATCCCCGCCGCGGGGGGGTGCAGGAACAA +
32	51640	CGGGGCGAGCGGGAGCCGAGCCTGCGCACGGGCGCGGGAAAGTCCCGGCCCAGACCGGT +
32	51700	CGCGGTCAGCCCGGGATGAGCGGCGAGCGAGGCCAGTTCCGCGCGGACTCCGCCAGTCT +
32		GTGATGGAGTTCCAGCGCGAACATGAGGTTGGCCAGCTTGGACTGGTTGTAGGCCCGGTA +
32		CCGGCTGTAGCGGCGTTCGCCGTGAAGGTCGCTGAAGTCGATGCGCCCAGCCGGTGCAG +
32		ATAGCTGCTGATCGTCACGACCCGCGCGCGCGCGCGCGCG
32		GCCGGTGAGGGCGAAGTGCCCCAGGTGGTTCGTGGCGAACTGGAGTTCGTGACCGTCCGG +
	52000	GGTGCGGGCCCGGTCGGTCCACATCACGCCCGCGTTGTTGACCAGCAGGTGGATGCGCGG +

32		CCACGCCCGGGCCAGCCAGGTGTAGTGCGGGCGCAACAACTGGTCGTCCACCTACGCGCC T R A R D T W M V G A N N V L L H I R P -	
32	52060	GAAGCGGTCGCGCAGTTCCTCGGCGCCGGCACGCACCGACGCGAGACGGGAAAGATCCAG +	
32	52120	CCGTCTGACCGTCAGTTGCGCCGACGGCACCCGGCTTTGGATGCGGGCCGCCGCGGCGAC +	}
32	52180	CCCGCGGTCCGGATCGCGCACGGCCAGCACCACGTGGGCGCCGTGCCGGGCGAGCTCCTG +	)
32		G R D P D R V A L V V H A G H R A L E Q -	
32	52240	GCGGTCCACGTCAGGCTACGGCCTCGACCGTGGCCACTGGTGGCGCCACCAAGGCCATGC A L H L G I G S S A G T V V A T T G T R -	•
32	52300	GTCCGGGACATCGCCGCGCTCCAGCGTCGCCGCGTTCTCATCGGTCGTCCCTCCC	Э
	52360	GATGCGTCAGCCGGCCTGGGCCATCGCGGCCCGGTAGCCGTTGGCGACGATCTGCCGGGC +	9
	52420	GGAGTGCTCGTAGTACTCGTCGTCGCAGCTCCGTGGCGAGACCGCTGACGTACCG +	9
	52480	GTTGAACATGCAGAACGCGGCGGCGATCAGAACGGTGTCGTGCAGAGCGGTGTCGTCCGC +	9
	52540	TCCCTCGGCCCGCGCGAGGCGATCACCCCTGCGGAGACCGGGCGCGCGC	9
	52600	CTCGGCGGCGACGGCCAGCAGCGCGCGCGTCCTGCCGTCGATGGGCGCGGTGCCGGGGTC +	9
	52660	GGCGAGGACGGCCTCGACGAGCTGCCGGCCGCCGCGGCGGCGAAGGCCCC +	9
	52720	GTGGGAGGCGCGCAGAACTCGGTGGAGTTGAGATGCGAGACGTACGCCGCGATGAGCTC +	9
	52780	GCGTTGCCCCGGTTCCAGCGAGGACGGCGCCCGCAGCAGGGCGTTCGCGAGATCGCCCAG +	9
	52840	CGGTGCTGCGGTGCCGGGGTGGTGAGCCATCAGACCACTGATGCCGGGGAGGTCGTTGTC +	19
	52900	GAGTGCTATGTGGGGCACGGCTCTTCCTTCCGGGTGGACGAGGGGGGGG	59

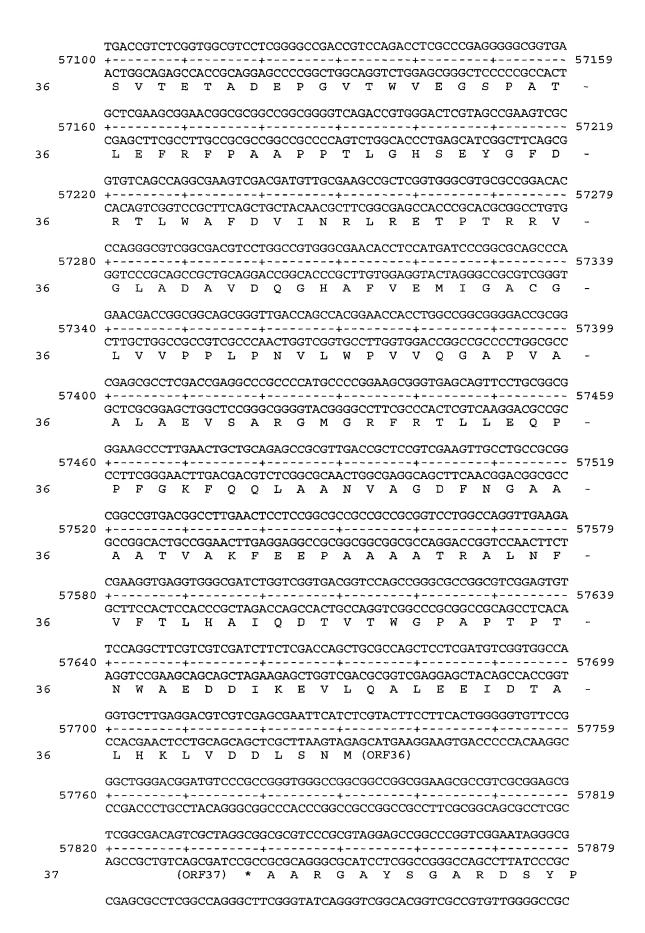
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	-0060	CAGGGCCATTCGACTTCGTCGTCGGCGGCGCGCGCAGATGCGGGTGAAGGGCCATTCCACG
	52960	GTCCCGGTAAGCTGAAGCAGCCGCCGCCGCCGCCGCCTCTACGCCCACTTCCCGGTAAGGTGC
	53020	TCTTCCCCTCCGGTGCGGAGTGGGCGGAGGCCGTGGTGAAGAGGGTGACGAGTCCGAAC +
	53080	GTGCCGAAGAGGAGGACAGTCGGGCAACGTGAAGTGCGGTACCCATGCGAGCTCCTAGC +
	53140	GAGGGCGGCGTGACCGCGGGACGTGAGACCTCGTGATGCCAGGAAGCTAGCGAATCGGA +
33	53200	CTGAGGGTGGCAACGATATGCCAGACTTTGGCAACTTGCCTGTGTATCAGCCGGACTGTC +
33	53260	GGCCGCTGGTAAAGACGGAACGGCGAGATCCCGCGACCGCGTCGCAGAGCAGCAGCAGGGTCT +
33	53320	GCTCACCCAGCGTCGGGGCCGGCCAGCATGTCGCGTACCGGGAGCGTGACGCCCAGCTCGC +
33	53380	GGTTGATCCTGCGGACCAGCCGGGTGATGAGCAGGAGTCGCCGCCGTGGGCGAAGAAAT ++
33	53440	CAGCACCTTCGGAGGGGTCCGGGAAGCCGAGCAGGTCACCCCAGCCGCGCACCAGTACCT ++
33		GGCGGATGTCGCCGGTGGTGACGACCGTGCGCCGGGAGCCCCGACGTGCCGAGCGCAGCC  +
33		GCGAGGCATGCACCAGCGCCACCTGGTCGCCGAGGTTGCGCCGCGACAGCTCGCGCAGCG ) +
33		ACACCGTGACGCCGAACCTCTCGGTGATCCTGCGGACCAGCCGCGTGATCAGCAGCGTGT  1 +
33	53680	CCCCGCCGCGCGAAGAAATCCGAATGCTCGGTGAGGTCGGAGCGGCCGAGGAGCTCGC ) ++
	5374	TCCACGCGCCGACCATGAACTCCCCCACGTCACCGAGCCGGTGCTCGTCGCCGTCGGGGC 0 +

33		H	A	P	$\mathbf{T}$	М	N	S	Ρ '	T S	S I	2 5	R	C	S	S	P	S	G	P	-
	53800	CCTTC	:GGC	:GCG	CCG	GAT(	CCC	GCGG	AAC	GGT	rcco	GCC	GGA	GAC	GCA	GAG(	CGG1	CAC	TGG	; - 5:	3859
33	53800	GGAAG	CCG	CGC	GGC	CTA	GGG(	CGCC A E	TTG:	CCA	AGG	CCGC	CCT	CTGC	CGT A	CTC	GCC	AGTC	GACC L V	:	
	53860	TCACT +		+	. – – –			+	- <b></b>		+			+			+			- 5	3919
33		Т	F	A	Т	S	R	G	M	C :	R I	L I	H F	L	P	A	Т	V	R	Е	-
	53920	AGCAC			<b>-</b>		<b>-</b>	+	<b>-</b>		+		<b>-</b> -	+	<b>-</b> -		+			- 5	3979
33		Н	M	L	H	G	N	T	F	P	S :	R I	M C	) P	s	L	P	S	T	H	-
	53980	ACTG		+				+	<b>-</b>		+		<del>-</del> -	+	<b>-</b> -		+			- 5	4039
33		TGAC	GTC <i>I</i> S	W W	CCGC A	TCT. R	ACA C	V	G G	:AAG I	R	G :	P (	V C	C	Q	A	A	R	D	-
	54040	ACCG		4				+		. – – <del>–</del>	+		- <b></b> -	+			+		<del>-</del> ·	- 5	4099
33		TGGC0	CGT2 H	ATG <i>I</i> T	AGAC L	BACC W	GTG H	TTG/ N	ACCI W	TAC N	TCA E	TCG *	CAC( (OR)	733)	CGGI	.GCC	CCT	GGC	CCG	Ĵ	
	54100	GGGC			+			+		<b></b> -	+		·	-+			+			- 5	34159
		CCCG																			
	54160	+ GAAA			+			+			+			-+			+			- 5	54219
	54220	CGGG	GGA	CGG	CAT	TCI	ACC	GGC	TAT	CTCI	rcgc	GCC	TGG	AGTC	GGG	CGCC	:CGG	;CAG	CCC	T - 9	54279
34	54220	GCCC	CCT	GCC	GTA	CAGA	TGC	CCG	ATA	GAGA	AGCG	CGG	ACC'	TCAG E S	CCC	3CG0	GCC	CGTC	GGG.	A S	
	54280	CCGA		<b>-</b> .	+			-+		<b>-</b> -	-+			-+			-+		<b>-</b>	- !	54339
34		GGCT	'AGC	GCG	GCA	GCGG	GTC	GAC	CGG	CCT	STTC	AGC	CGT	AGTO I S	:GGG	CAG	CCTC	CAAG	CTT	'C G	
	54340	GGTC		<b>-</b>	+			-+	- <b></b>	<del>-</del> -	-+		<b>-</b>	-+			- +			- :	54399
34		CCAG	GGC R	CCG A	GTG T	GAG( S	CGA( L	GCGG A	GTC Q	TAG( I	GAGA L	AGGC S	AGA L	GGT0 S I	AAG S	GGA( L	CCTC E	CAGG S	CTG D	F.	-
	54400	AGAC			+			-+			-+			-+			-+				54459
34		TCTG	GTC S	ACT E	CGA L	AGA(	GCG( A	GCTC E	CGC A	CAT( V	GCA/ R	AGG( S	CGCG A	TACO H	CGGT GQ	CCT D	AGG( P	CTAC M	CGAG L	₽G R	. <del>-</del>
	54460	GCTC			+			- + - <b>-</b>	. <i>– –</i>		-+-			-+			-+-		<del>-</del> -		54519
34		CGAC	ጉርርጥ	reed	GGA	CAC	CGA	CGAC	CCT	GTC.	ACC'	$\Gamma TCC$	GCGG	R I	rgcc	GCT	GAG	CCGC	3CCC	÷C	
	54520	AGCA	- <b>-</b> -	<b>-</b>	+			-+			-+-		<b>-</b> -	-+-			-+-				54579
34		ጥርርግ	racc	GAT	CGA	GGT	CGC	AGAC	CCAC	TGC	GAC'	$\Gamma CA$	CTCC	E :	$^{A}GCC$	'GGA	.CCG	GCT	CAAC	CG.	
		GCG	CACC	GGC	CCI	GAC	CCA	GTT(	CGCC	CGG	TCG	CTĢ	CGGC	TAC'	rgge	CGA	GAT	CGT	TCC	GG	

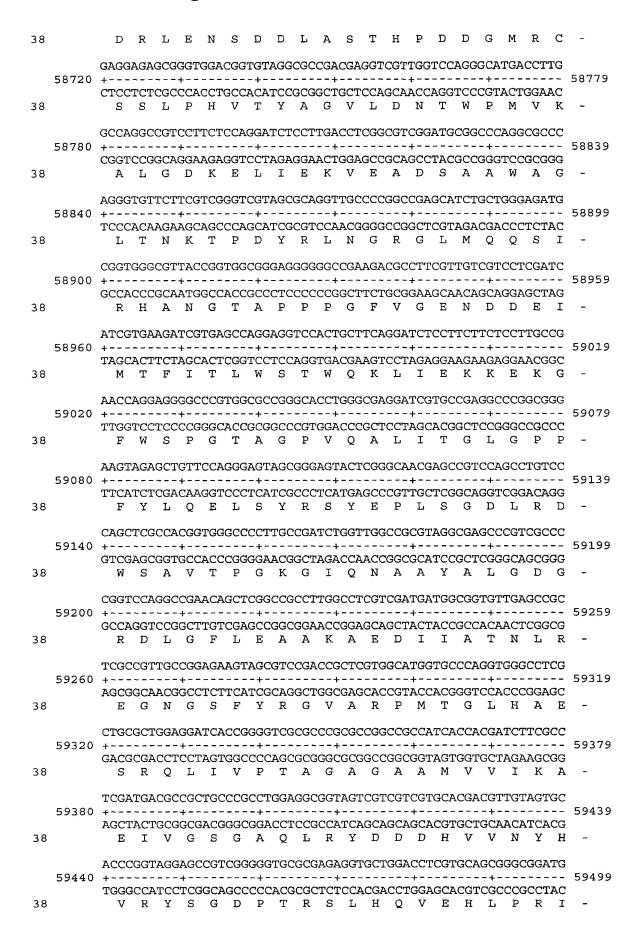
34	54580	CGCGTGCCCGGGACTGAGCGGGCCAGCGACGCCCATGACCCGCTCTAGCAAGGCC A R A L T Q F A R S L R V L G E I V P A -
34	54640	CGGTGGAGGCTGCCGCCGCCCACCGGCTCGCGGTGGACCATGCGCTGTCCAGCCAG
34	54700	ACAGGGCCGCTTCGCTGCTGGTTCTGGTGTCGGTGGAGGCCGAGGCGGGACGGATGCCCG +
34	54760	ACGCCCGGCGCCACGCCGACGAACTGACCGTCCTGGTGAGGGGACGGTCCGACACTCTGT +
34	54820	GGGCCGAGGCGTTGTGGACGGCGGGTGCGTTGAAGGTGCGGCAGGGCGAGTTCGCCGCGGGCGCGCGC
34	54880	CCGAGGTCCTTTTCCAGGAGGCTCTGGACGGGTTCGACAGCCGGGAGAACCTGACGATCT +
34	54940	GGCTGCGGCTGCGCATCGCGATGGCCGAACTCCACCTGCAGAAACTTCCTCCCGAGCCCG +
34	55000	ACGCCGCGCAGCTCTGCATCGAGGCGGCGGAGGCGCCCTTCCCTTTGCCCGCACATCCG +
34	55060	CTCTGGAACAGTCCCTCGCCGCTCTGCGGGCGCGCCTCGCCTTCCATGAGGGCAGGTTCG +
34	55120	CCGATGCCCGCGCGTTGTTGGAGAGGCTCGGCAGGACCGAGCTCCGGCTGCCCTATCAGA +
34	55180	GCCGGATCCGCCTGGAGGTCCTCGGTCATCAGCTGCGCATCCTGAGCGGGGAGGAGGAGGAGGAGGAGCCGCCTAGGCGGGAGCCAGCGCGTAGGACTCGCCCCTCCTCCCCCCCC
34	55240	AAGGCCTGGCCGGCCTCCAGCTCCTGGCCGAGGAGGCGCAGGAGAACTCCAACATCAACC +
34	55300	TCGCCGCGGAGATCTGGCGGCTCGCGGCGGAATGCCTGATGCGGGCGCGCGGGAAGGTCC +
	55360	GCGGCGCCACCGGCGCTGACGCCGCCGCTCGCGAGGTCCACCGCGCCGCCGTGGCC

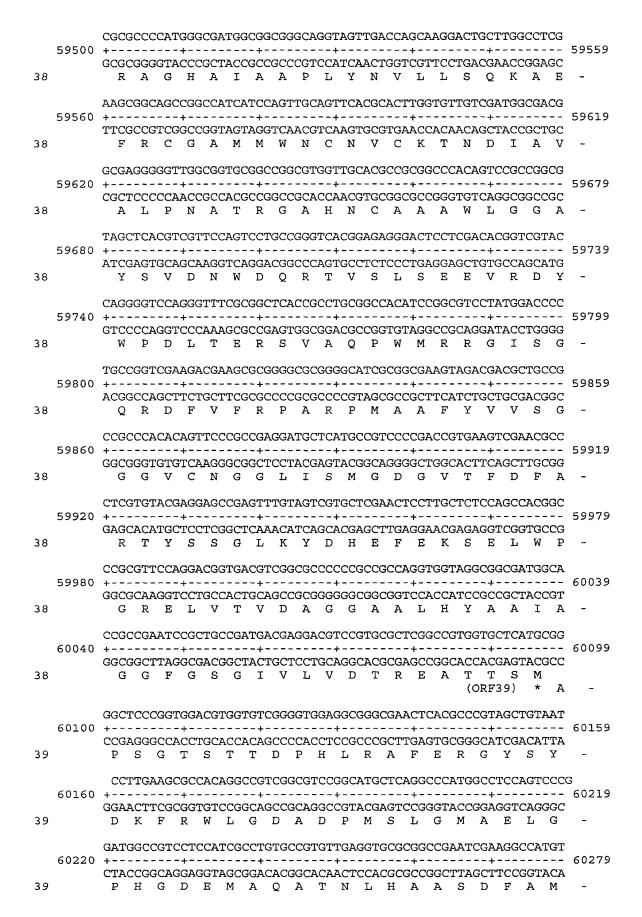
34		GCCGCGGGGCGCCGACTGCGGCGCCGAGCGCTCCAGGTGGCGCGCGC	
	55420	ACCGCCGTCGGCGTGAGGCGCCGCGCGTGTGCCCCCCCACGGTTGCTCGCCCTTGGTGG	55479
		TGGCGGCAGCCGCACTCCGCGGCCGCACACGGCGGGGGTGCCAACGAGCGGGAACCACC	
	EE480	TGCATCTGTTGGCACATGTGTACCTCCTACACAGTCAATTGTTGCCAAAATTGTCGAACC	EEE30
	55460	ACGTAGACAACCGTGTACACATGGAGGATGTGTCAGTTAACAACGGTTTTAACAGCTTGG	22233
		${\tt GAATGGCAATTGCTTGCCTTTGCTGAAGAGGCGTGCTGATATGCAAGTCAAGTAGCCTCC}$	FFF00
	55540	+ + + + + + + + + + + + + + + + + + + + + + + + + + + + +	55599
		${\tt TCCGATCTCGGGCGCCATATGGGAAACATCGAGTTGAGCGCGATGGCGTTCGTCAGTG}$	
	55600	+++++++	55659
		CTGCCGTTCTGGCCAGGCAACTGATGTCGATGGGGATGGCAAGATTTTGCCGAAAACCGA	
	55660	#	55719
		TACATCTCTGTCCGGCACAGCCTTCGCCCCCCGGGTGACACTGCTCCGGCATGGCT	
	55720	ATGTAGAGACAGGCAGGCCTGTCGGAAGCGGGGGCCCACTGTGACGAGGCCGTACCGA	55779
		CCGGTTTCTCGTCGCCCGGCCGACGGACCGCACCGTCCGGAACGAGGCGCCGGTGTGCGT	
	55780	######################################	55839
		CCGCTGATGGGCACAGCGGCCTCGGCCGCAGCAGGTTCCCACCGAGAAGAATGCCGAGGC	
	55840	#	55899
		CCAGCCGTGAACCACGACATGTCCCAGCGTGCCTTGCTGGAGGCGGCGGCCGAGGGGCTG	
	55900	GGTCGGCACTTGGTGCTGTACAGGGTCGCACGGAACGACCTCCGCCGCCGCCTCCCCGAC	55959
		CGGCGGCTGGCCGGCGCGCGCGCGCGCGCGCGCGCCCCCC	
	55960	#	56019
		AGGGACATGTTCTCCCCGCCGCCGCCGGTACGTGCTCGCCTCGGACCGCGCGGGGTTC	
	56020	+	56079
35		(ORF35) M F S P A A R R Y V L A S D R A G F	
	56080	TTCGAGCAGGCTGTCCGGCTCCCGGGGGTACCGGGTGAGCGCGGAGTTCGTCGGC	56139
35	30000	AAGCTCGTCCGACAGGCCGACGCGAGGGCCCCCATGGCCCACTCGCGCCTCAAGCAGCCG F E O A V R L R S R G Y R V S A E F V G	
35		CCCGATCAGGGAGCCACCGACGCCCTCCACGCGGAGCACGTGGTCGAAGAGCACCTGAGG	_
	56140	++	56199
35		$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	-
		CTGCTCGATCAGGAGCCGGCCCCTGACCGGATCGGTGTGGACGTCTCCCGGATCGGCCTC	
	56200	### ##################################	56259
35		L L D Q E P A P D R I G V D V S R I G L	_
	56260	GCCACTCGGCGAGACTGCCTGCGCAACACCGGGCGGCTGCCGCTGCGGCGCTC	56319
	23200	CGGGTGAGCCGCGTCTGACGGGACGCGTTGTGGCCCGCCGACCGA	

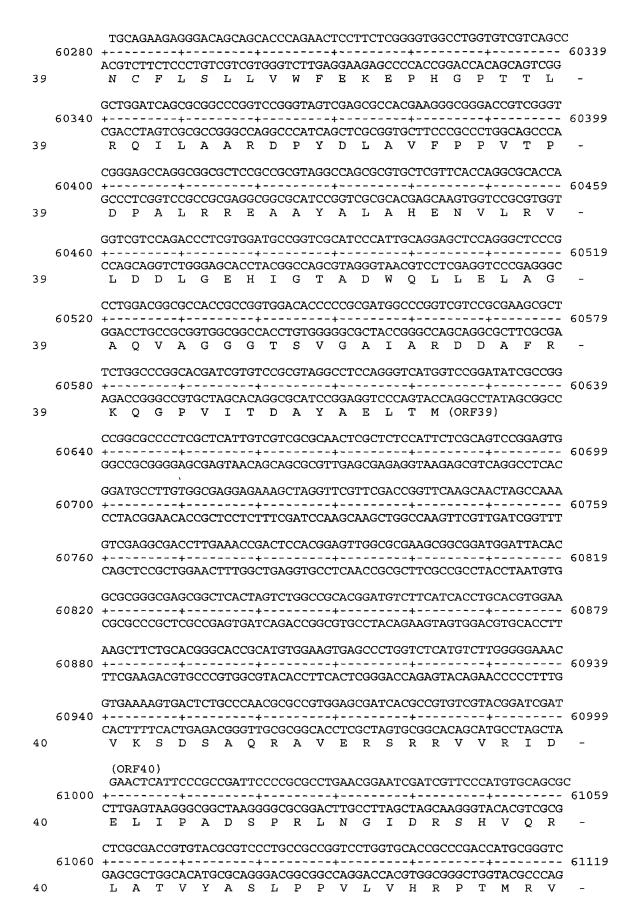
35		A	Н	s	A	Q	Т	A	L	R	N	Т	G	R	L	A	A	A	A	A	L	-
	56320	+	GGGZ  CCC	- <b>-</b> -	+	<del>-</del> -		+-			+	- <b>-</b> -	- <b></b> -		- +		- <b>-</b> -	-+-				56379
35		R	G	S	E	V	V	L	L	M	Е	G	S	Ε	D	Ι	D	Т	V	L	A	-
35	E 6 3 8 U	+	CATO GTAO H	 TG0	- + - ·	GAC	 CAC'	+ TTG(	GCAZ	ATG(	: CTG:	+ - <b>-</b> - TTG(	CAC	CCC	-+-: [AG:	rgg(	JAAC	+- 3TC(	 CGC	 GTG	GAC	56439
	56440	CA	CCG	CAC	CGT	GGA	CGA	CGC	CAT	GGC	GGT	CGC(	GGG	rcc'	TGG(	CCG	CAC	CGT(	GCG	GCT	GGTC	56499
35	56440	GTG H	GCG' R	TGG(	CAC( V	CTG D	CTG D	CGG A	TAC M	CGC A	CAG V	CGC( A	CCA( G	GGA P	CCG( G	GCG' R	T T	CAC V	GCC R	GAC L	CAG V	-
	56500	+	GGC'  !CCG		-+-			+				+			-+-	<b></b> -		+				56559
35		M	G	S	S	A	E	P	A	G	T	Α	L	S	R	G	P	A	L	Ε	D	-
35	56560	+ GCC		 GAA	- + <b>-</b> CTG	 GAG	 CGC	+ CTC		 GAG	CAC	+ CTG	 GCA	 CCG	-+- CAG	 GCC	 GAC	+ TCA	GAC	CGG	TGA	56619
	56620	+ GG(	 :CTG	 CGG	-+- CTC	CAG	GAC	+ CGG:	 CCC	 CGC		+ CTC	 GCA	.CCA	-+- GAC	GAG	 CTT	+ GCG	CAC	GTC	CTG	C 56679
35			D																V :GCC		D CCGC	_
35	56680	<u>+</u>	 GCTC	TAC	-+- GAC	ATO	 GCCA	+		GGG	 CTC	+ GAC	GAC	GCG	-+-	 GTG	 GCC	+ CGC:	CCG	CCC	GCG	56739 -
	56740	+	 3202	 \GCG	-+- 74T	 GTC	 <del>-</del>	+ CCG(	⊦ 3ATG	 3CC0	CTAC	+ TTC	ACC	ACC	- + - CGAA	GGG	GAC	 GAC	 CGC(	CTC	CGAC	56799
35		GCI	C	ממי	ירככ	፥ሮሮር	ገ <b>ጉ</b> ፈር	<b>፡</b> ርጥር	CTC	CAAC	CGC	CTC	GCC	GA	CATO	CGGC	CGG	GAC	CCG	GGA	GCCC	-
35		+ - CG		<del>-</del>	- + -	 :GG(	 CTA(	CCA	+ <b>-</b>	- <b></b> -	GCG(	- + <b>-</b> - 3GA(	CCGC	- <b></b> -	- + - 3ATE	 3CC(	GCC	CCT	+ 3GC	CCT	CGGG P	56859
35		+- CA		 3GT0	+ 3GT(	CCG	CAT	GAC	+ · TGG(	GCG(	GGG(	-+			+-				+	<b>-</b>	GCCC	56919
33		GC	GCC(	GGT(	CAG( + :	CTC	CCG 	GTC	GCC( +	GCG:  CGC'	AAC'  TTG	-+- ACG	GGC	ccg	GAC	GCG	GGGZ	AGC	+ GGG		CCGC	- 56979 <del>3</del>
36		CC	(OR	F36)	عرب -	* ፕርር	S GCG	G ATG	T :	A . AGC	a cac	F (	TCC	g GCC	P 1 TCC	R : TGA:	R (	g : GAC	E GCG	G GTC	A I	? -
36		GC	GCC	ATC	CGG	ACC	CGC	TAC	AGG'	TCG	GTG	AAG.	AGG	CGG	AGG.	ACT'	GG.L.	CIG	CGC	CAG	TCC	- 57039 -
		GC	TCC	agc	+ AGC	GCC	ACC	 GCC	+ <b>-</b> -	 GCC	CAC	-+- TGG	 TCG	 TCC	+ GTC	 TTC	 AGC	acg	+ CGC	CCI	'GGC	- 57099 3
36	5	P	L	D	D	R	H	R	R	. R	: Т	V	L	L		F.	ט	H	L	s E	- 6	-



37	57880	GCTCGCGGAGCCGTCCCGAAGCCCATAGTCCCAGCCGTGCCAGCGGCACAACCCCGGCG A L A E A L A E P I L T P V T A T N P G	57939 -
37	57940	GCATGCAGGCGATGCGCTGGCGTCCCCGCGCCACCAGGGTCTCGCCGCCGTCGTCGCCCA ++ CGTACGTCCGCTACGCGACCGCAGGGGCGCGGTGGTCCCAGAGCGGCGCAGCAGCGGGT R M C A I R Q R G R A V L T E G G D D G	57999 -
37	58000	GCTTGATGTAGTCGAAGGTGAACTCCAGCTGGGTCTGCCGCAGCTCCGAGAGCCTCATCC +	58059
37	58060	GGATCGACAGTTCGTCGAAGGCGGTGATCTCCGCGAAGAACTCGCAGTCCACCTTGAGGG ++ CCTAGCTGTCAAGCAGCTTCCGCCACTAGAGGCGCTTCTTGAGCGTCAGGTGGAACTCCC R I S L E D F A T I E A F F E C D V K L	58119
37	58120	TGAAGAGCTTGAGGTCCTCCTGGACCTCGGCGAGCACCGAAGGCGCCCTCTCCTTGAGAA ++ ACTTCTCGAACTCCAGGAGGACCTGGAGCCGCTCGTGGCTTCCGCGGGAGAGGAACTCTT T F L K L D E Q V E A L V S P A R E K L	
37	58180	AGAGTTCCCGGCAACGCCCCTGCCAACGAAGGTAGTTGACGTAGTAGACGTTGCCGACGA ++ TCTCAAGGGCCGTTGCGGGGACGGTTGCTTCCATCAACTGCATCATCTGCAACGGCTGCT F L E R C R G Q W R L Y N V Y Y V N G V	
37	58240	GGTTCGTCTCCTCGAAGCCGACGGTGTGGCGGAGCTCGAAGTAGTCAGGATTCGTCGCGG ++ CCAAGCAGAGGAGCTTCGGCTGCCACACCGCCTCGAGCTTCATCAGTCCTAAGCAGCGCC L N T E E F G V T H R L E F Y D P N T A	58299
37	58300	TCATAGGTCTGTGCCCTTCGTCGGGGGCCGGTCGTCGCACCGAGTTGCGTGAAGCAAC ++ AGTATCCAGACACGGGAAGCAGCAGCCCCGGCCAGCAGCGTGGCTCAACGCACTTCGTTG T M (ORF37)	58359
	58360	TCACTGGTCGCGATGGCCTGCGGGGTCGGTGGCCCGCGCTCCGGGCGAGAGTGCGGGCG ++ AGTGACCAGCGCTACCGGACGCCCCAGCCACCGGGCGCGAGGCCCGCCTCTCACGCCCGC	58419
38	58420	GGGTGCCGGCCGGGGGTCAGCCGCGCGCGCGAGGGGAAGAACCCTCTCG ++ CCCACGGCCGGCCGCGCGCGCGCGCGCGCGCGCGCGCGC	
38		CGGCCGCTCGTGGAGCCGTCGGGGGCCGGTGCGCCGTAGGTGACGGAGATACCCCGGCTC ++ GCCGGCGAGCACCTCGGCAGCCCCCGGCCACGCGGCATCCACTGCCTCTATGGGGCCGAG R G S T S G D P A P A G Y T V S I G R S	58539
38	58540	TGCGCGGCGCGCACGATCCCCGGCATCGCGCGTTCGGCGAGCGCCGCGATGGTCATCGCG  *	58599
38	58600	GGATTGACCGTCAGCGCGCGGGAACCGACGATCCGTCGGTGACGAAGATCCCCGGGTGG  CCTAACTGGCAGTCGCGGCCCTTGGCTGCTAGGCAGCCACTGCTTCTAGGGGCCCCACC  P N V T L A G P V S S G D T V F I G P H	58659
	58660	TCGCGGAGCTCGTTGCTGTCCAGGGCGGATGTGTGGGGGTCGTCGCCCATCCGGCAG ++++++++	58719







	61100	GTCGACGGCATGCACCGCATCGGCGCGGCCCGCCTGAAGGGGCTGGACACGGTCGAGGTC																				
	61120	CAG																				611/9
40											A								V		V	-
		ACC:	ייריריי	יייריי	ZAC(	3GC	<del>ነ</del> ርር(	ነጋፈና	GAG	ሮልር	ርጥርነ	ጥጥር	ርጥር	CGT	ፐሮሮ	<b>ን</b> ሞርር	3CG(	3CG)	AAC	ATC	ACC	
	61180	+	<b></b> - ·		-+-	<b></b> -	- <b>-</b> - ·	+	<b>-</b>			+		- <b>-</b> -	-+-			+				61239
40		TGG/	AAGZ F	AAG( F	CTC( E		CGG( A		CTC E		CAC. V					CAG( V			TTG: N	ΓAGʻ Ι	TGG T	_
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	61240	TTG																				61299
40		N	G	L	P	L	S	V	A	D	R	K	T	A	Α	А	R	I	L	А	s	-
		CAC	CCG	ACC	CTG:	rcc	3AC	CGC	GCG	GTC	:GCC	GCA	CAC	GTC	GGC	CTC	GAC	GCC2	AAG	ACC	GTG	
	61300	+	- <b>-</b> -	<b>-</b>	-+-			+				+			-+-	- <b>-</b> -	- <b>-</b>	+				61359
40		GTG(	GC'. P								CGG A											_
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	61360	GCG																				61419
	01300																					01417
40		A	G	V	R	Т	C	S	A	A	G	S	P	L	L	N	M	R	T	G	A	-
		GAC	GC(	CGC	GTC(	CAC	CCG:	rtg	GAC	CGC	ACC	GCC	GAA	CGC	CTG	CAC	GCG	GCC	GCG	CTG	CTG	
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40		V	G	P	Α	A	R	P	V	P	K	V	p	P	A	V	A	G	R	P	P	-
		GTG'	rca(	CCG	CGG'	TCC	CGG	GCC	CCG	СТС	GAG	GCG	CTG	CGC	AAG	CTC'	TCC	AAC	GAC	ccc	TCC	
	61720	+			-+-			+	- <b>-</b> -			+			-+-		- <b>-</b> -	+				61779
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40		L	R	H	s	D	Q	G	R	E	$\mathbf{L}$	M	R	W	L	H	N	R	F	V	V	-
		GAC	GAG	CGCGTCCACCGTTGGACCGCACCGCCGAACGCCTGCACGCGGCCGCGCTGCTG +																		
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	61000	GAGCTGGCGCACCACTGCTCGGACGCCTGGCACCGGTTCGCCGAGGAGATGGTTCGGCGC	C1.0F0
40	61900	CTCGACCGCGTCGTGACGAGCCTGCGGACCGTGGCCAAGCGGCTCCTCTACCAAGCCGCG E L A Q H C S D A W H R F A E E M V R R	-
40	61960	CGGCACAGCGCCGCGGCCGACGGCTCCGGACTCCGCACGACTCAGCCAACTCGCCGTTGA  ++ GCCGTGTCGCGGGCGCGGCTGCCGAGGCCTGAGGCGTGAGTCGGTTGAGCGGCAACT  R H S A A A D G S G L R T T Q P T R R *  (ORF40) -	62019
41		CGGCCTACTTCGACAGGAGTTACGGTGACCACGAACACCATCGAGGACGCGGTCCGCCG ++	62079
41		GGTCGTCGAGTACATGCACGTCAACCTGGGTCAGAACCTCACGATCGAT	
41	62140	CACGGCGATGTTCAGCAAGTTCCATTTCACCCGCATCTTCCGCGAAGTCACCGGTACCTC ++ GTGCCGCTACAAGTCGTTCAAGGTAAAGTGGGCGTAGAAGGCGCTTCAGTGGCCATGGAG T A M F S K F H F T R I F R E V T G T S	62199
41	62200	TCCCGGGCGTTTCCTGTCCGCCTTACGGATTCAGGAGGCCAAGAGACTTCTCGTGCACAC  ++ AGGGCCCGCAAAGGACAGGCGGAATGCCTAAGTCCTCCGGTTCTCTGAAGAGCACGTGTG  P G R F L S A L R I Q E A K R L L V H T	6 <b>22</b> 59 -
41	62260	TGCACTCAGTGTGGCCGATATCAGCAGTCAGGTCGGCTACAGCAGTGTCGGTACTTTCAG  ++ ACGTGAGTCACACCGGCTATAGTCGTCAGTCCAGCCGATGTCGTCACAGCCATGAAAGTC A L S V A D I S S Q V G Y S S V G T F S	62319
41	62320	TTCTCGCTTCAAGGCCTGTGTGGGGCTTTCCCCGAGCGCCTATCGCGACTTCGGCGGGGT  ++ AAGAGCGAAGTTCCGGACACCCCCGAAAGGGGCTCGCGGATAGCGCTGAAGCCGCCCA S R F K A C V G L S P S A Y R D F G G V	
41	62380	GCAGCCGGGTTTTCCCTCCGCCGCGCCCGTCTCACTCCCACCGCGCACAATCCCTCCGT ++ CGTCGGCCCAAAAGGGAGGCGCGCGCGGCAGAGTGAGGGTGGCGCGTGTTAGGGAGGCA Q P G F P S A A A R L T P T A H N P S V	62439
41	62440	GCGCGGCCGCATTCACTCCGCCCCGGGTGACAGGCCCGGAAGGATCTTCGTGGGCCTGTT  ++ CGCGCCGGCGTAAGTGAGGCGGGGCCCACTGTCCGGGCCTTCCTAGAAGCACCCGGACAA R G R I H S A P G D R P G R I F V G L F	-
41	62500	CCCCGGCAGGATGCGCCAGGGCCGCCCGGCGCGCGCGCTCATGGAGAGTCCCGGGGC  ++ GGGGCCGTCCTACGCGGTCCCGGCGGCCGCGCGCGCGCGC	
41	62560	CTTCGAGCTCCGGGACGTGCCCGTGGGCACCTGGCACATCCTGGTCCACTCCTTCCCCGC  ++ GAAGCTCGAGGCCCTGCACGGGCACCCGTGGACCAGGTGAGGAAGGGGCG F E L R D V P V G T W H I L V H S F P A	
41	62620	CGGACACCGGCCGCACCAGCTCGACTCCGAACCGCTGTTGCTCGGGCACAGCGGACCGCT  +++	

